



## Concise Review

# The genetics of nonalcoholic fatty liver disease

Christoph H. Österreicher;<sup>1</sup> David A. Brenner<sup>1</sup>**Abstract**

The increasing prevalence of obesity in Western countries has led to a significant increase of nonalcoholic fatty liver disease (NAFLD) over the past decades. Being part of the metabolic syndrome, NAFLD is thought to be the most frequent cause of elevated liver enzymes in the United States affecting up to one third of the population. NAFLD is also proposed to be the major cause for cryptogenic cirrhosis and hepatocellular cancer of unknown etiology, and thus, represents one of the most important problems for hepatologists in the future. However, the natural course of NAFLD is highly variable and is influenced by both environmental and genetic factors. Polymorphisms in specific genes have been proposed to increase the risk of fibrosis in patients with NAFLD. The present review article summarizes currently available data from genotype-pheno-

type studies and defines candidate genes that deserve future investigation.

**Key words:** Nonalcoholic fatty liver disease, nonalcoholic steatohepatitis, single nucleotide polymorphism, genetic risk, susceptibility, steatosis, fibrosis.

**Introduction**

The epidemic of obesity in the Western has increased the prevalence of nonalcoholic fatty liver disease (NAFLD) rendering it now the leading cause of referral to hepatology clinics. NAFLD represents the hepatic manifestation of the metabolic syndrome which is characterized by obesity, type 2 diabetes mellitus, and dyslipidemia, with insulin resistance as a common feature. Indeed, it is estimated that 17-33% of Americans are affected. NAFLD is now believed to be the major cause for abnormal liver function and fibrosis in industrialized countries.<sup>1</sup> NAFLD refers to a spectrum of histological findings ranging from simple and reversible steatosis to steatohepatitis and cirrhosis, and is diagnosed after ruling out other causes, in particular alcoholic liver disease (ALD).<sup>2</sup> NAFLD may account for the majority of cryptogenic cirrhosis cases. In this respect, features suggestive of NAFLD such as obesity or insulin resistance are more frequently observed in patients with cryptogenic cirrhosis than in age- and gender-matched patients with cirrhosis of known etiology. Remarkably, hypertriglyceridemia and diabetes are independent risk factors for the development of hepatocellular cancer (HCC) in patients with cryptogenic cirrhosis, suggesting that HCC represents a complication in an unknown percentage of cases of nonalcoholic steatohepatitis (NASH) induced cirrhosis.<sup>3</sup>

While the majority of obese people only develop simple hepatic steatosis, around 10-20% of morbidly obese patients progress to more advanced disease.<sup>4,5</sup> The current concept of liver fibrosis suggests that the natural history of liver diseases in general is influenced by both environmental and genetic factors that act in concert.<sup>6</sup> The former are not sufficient to explain the wide variety of phenotypes among individuals subjected to profibrogenic insults, and the latter are largely unknown. While the risk factors for NAFLD are well established, little is known about genetic factors.

Fibrosis in NAFLD is thought to be the result of a non-specific wound-healing response to inflammation and re-

<sup>1</sup> University of California, San Diego, Department of Medicine.

## Abbreviations

Nonalcoholic fatty liver disease (NAFLD), alcoholic liver disease (ALD), hepatocellular cancer (HCC), tumor necrosis factor alpha (TNF- $\alpha$ ), free fatty acids (FFA), reactive oxygen species (ROS), triglycerides (TG), microsomal triglyceride transfer protein (MTP), very low density lipoprotein (VLDL), alanine aminotransferase (ALT), phosphatidylethanolamine N-methyltransferase (PEMT), carnitine palmitoyl-transferase 1A (CPT-1A), manganese-dependent superoxide dismutase (MnSOD), hereditary hemochromatosis (HHC), glutathione-S-transferase (GST), myeloperoxidase (MPO), microsomal epoxide hydrolase (MEH), lipopolysaccharide (LPS), Toll-like receptor-4 (TLR4), hepatic stellate cells (HSC), platelet derived growth factor (PDGF), matrix metalloproteinases (MMP), primary sclerosing cholangitis (PSC), nonalcoholic steatohepatitis (NASH), methylene tetrahydrofolat reductase (MTHFR), extracellular matrix (ECM).

## Address for correspondence:

David A. Brenner  
University of California, San Diego  
School of Medicine  
Vice Chancellor Health Sciences Office  
9500 Gilman Drive # 0602  
La Jolla, CA 92093-0602  
Phone: 858-534-1501  
Fax: 858-822-0084  
E-mail: dbrenner@ucsd.edu

peated injury, ultimately resulting in the formation of scar tissue instead of parenchyma.<sup>6</sup> Given the histological similarity between NAFLD and ALD it is not surprising that an emerging body of evidence suggests that their pathological mechanisms are very similar and involve cytokine and oxidative stress-mediated injury. Tumor necrosis factor alpha (TNF- $\alpha$ ) is of special interest in the pathogenesis of NAFLD and may be produced by Kupffer cells in response to gut derived endotoxin, but also by hepatocytes in response to an increased supply of free fatty acids (FFA), or by adipose tissue macrophages.<sup>2,7</sup> Reactive oxygen species (ROS) in the setting of NAFLD principally arises via increased oxidation of FFA by mitochondria, peroxisomes, and microsomes. In addition, there is emerging evidence that factors related to obesity and insulin resistance may be directly fibrogenic.<sup>7</sup> These include insulin and adipokines synthesized and released by adipocytes, including angiotensinogen, norepinephrine, and leptin. The anti-inflammatory and anti-steatotic adipokine adiponectin may also contribute to steatosis and inflammation in NAFLD and protects from NAFLD associated carcinogenesis as suggested by studies carried out in mice<sup>8</sup> (Kamada Y, Journal of Hepatology, in press).

### Evidence for genetic factors

There is convincing evidence that genetic factors account for considerable variability in the natural history of NAFLD. Evidence for a contributing effect of genetic factors on the development of advanced fibrosis in NAFLD derive from family clustering studies showing that about one fifth of patients with NASH have a similarly affected first degree relative.<sup>9</sup> In another study, co-existence of NASH and cryptogenic cirrhosis was observed in seven out of eight families studied.<sup>10</sup> In addition there appears to be interethnic variations in susceptibility towards NAFLD. In this respect, it has been reported that the prevalence of cryptogenic cirrhosis in Hispanic and African-Americans was threefold higher and fourfold lower, respectively, compared with Americans of European origin. Assuming that these groups share a similar prevalence of type 2 diabetes, this observation suggests that Hispanics are at particularly high risk.<sup>11</sup>

Compared to chronic HCV infection and ALD,<sup>2,13</sup> only little is known about factors influencing fibrosis progression in patients with NAFLD. With regard to the pathomechanisms underlying NAFLD, variations in genes affecting hepatic lipid metabolism, insulin resistance, ROS formation and degradation, cytokines and endotoxin receptors, and profibrogenic mediators are of special interest.

### Genes influencing lipid metabolism and steatosis

Studies from Japan suggest that genetic predisposition to obesity and inflammation contributes to the de-

velopment of NASH.<sup>4,15</sup> These studies investigated functional genetic variants of  $\beta$ -adrenergic receptors as they play an important role in the regulation of energy expenditure, in part, by stimulating lipid mobilization through lipolysis. Patients with NASH carried significantly more frequently the codon 64 Arginine variant of the  $\beta$ 3-adrenergic receptor than control subjects, which results in hypertriglyceridemia, hyperinsulinemia and obesity.<sup>14</sup> Homozygous carriers of Glycine at codon 16 of the  $\beta$ 2 adrenergic receptor have lower high-density lipoprotein cholesterol level than Arginine homozygotes, and presence of Glutamine at codon 27 is associated with higher concentrations of serum triglycerides (TG) and a higher prevalence of steatosis.<sup>15</sup>

The microsomal triglyceride transfer protein (MTP) regulates synthesis, storage, and export of hepatic TG content and is critical for the synthesis and secretion of very low density lipoprotein (VLDL) in the liver. Low levels of MTP result in failure to excrete triacylglycerol from the liver and hepatic steatosis. Two studies provide evidence that genetic variation of the MTP gene affects susceptibility to development of NAFLD. In patients with type II diabetes, the -493 G/T MTP gene polymorphism was independently associated with elevated an serum alanine aminotransferase (ALT) level as a surrogate marker for steatohepatitis.<sup>16</sup> Accordingly, patients with NASH had a higher frequency of the G allele and the G/G genotype compared to controls. Interestingly, the G/G genotype was also associated with more severe steatosis and more advanced stage of NASH compared to patients with genotype G/T.<sup>17</sup> Noteworthy, the same polymorphism was also reported to affect progression to advanced fibrosis in patients with chronic HCV infection.<sup>18</sup>

A loss-of-function mutation of the phosphatidylethanolamine N-methyltransferase (PEMT) gene, which is required for phosphatidylcholine synthesis, interferes with efficient VLDL synthesis. This mutation was reported to confer susceptibility to NAFLD, and is supported by data from PEMT knock out mice which also develop fatty liver disease.<sup>19</sup>

The genes coding for the LDL receptor, the ApoE protein, the methylene tetrahydrofolat reductase (MTHFR) and the carnitine palmitoyl-transferase 1A (CPT-1A) also display variations that have been proven to affect the course of chronic HCV infection and thus deserve future investigation in patients with NAFLD (*see table II*).

### Genes affecting oxidative stress

Genetic variations of enzymes involved in generation and degradation of ROS are of special interest with regards to NAFLD, as excessive FFA oxidation leads to oxidative stress causing hepatocyte apoptosis and liver injury. Manganese-dependent superoxide dismutase (Mn-SOD) is the main ROS scavenger in mitochondria. A Japanese study has reported that the T/T genotype of the

MnSOD gene which leads to less efficient transport of MnSOD to the mitochondria is a susceptibility allele for the development of NAFLD.<sup>17</sup>

The HFE gene is another attractive candidate to be studied as genetic modifier of NAFLD. Mutations of the HFE gene cause hereditary hemochromatosis (HHC), which is the most common genetic disease in popula-

tions of European ancestry. In addition, HFE gene mutations are common among Caucasians and have been demonstrated to be functional in terms of increased duodenal iron absorption and liver iron deposition. Increased liver iron can act as a catalyst promoting oxidative stress in a steatotic liver. As steatosis has been reported to affect the natural course of HHC<sup>20</sup> one would

**Table I.** Genotype-phenotype studies in NAFLD.

Gene	Protein function	Principal finding	Reference
$\beta 3$ adrenergic receptor	Regulation of energy expenditure	Presence of arginine at codon 64 is more frequently observed among patients with NASH	14
$\beta 2$ adrenergic receptor	Regulation of energy expenditure	Presence of glycine at codon 16 is associated with lower HDL cholesterol levels	15
MTP	Regulation of hepatic TG content	Presence of glutamine at codon 27 is associated with higher serum TG levels and a higher prevalence of steatosis	15
		G allele is independently associated with elevated serum ALT levels	16
		G allele is more frequently observed in patients with NASH	17
		G/G genotype is associated with more severe steatosis and more advanced stage of NASH	17
PEMT	Phosphatidylcholine synthesis	Loss-of-function mutation confers susceptibility to NAFLD	19
MnSOD	ROS degradation	T/T genotype confers susceptibility for the development of NAFLD	17
HFE	Duodenal iron absorption	Heterozygosity for the C282Y mutation is associated with more severe fibrosis	21,22
TNF- $\alpha$	Proinflammatory insulin resistance	-238 polymorphism confers susceptibility towards NAFLD and is associated with insulin resistance	25
IL-1 $\beta$	Proinflammatory	T allele and the T/T genotype at position -511 is more frequently observed in NASH patients	14
CD14	Endotoxin receptor	T allele confers increased risk for the progression from steatosis to NASH	27
TGF- $\beta 1$	Profibrogenic	Presence of arginin at codon 25 is associated with advanced fibrosis	28
Angiotensinogen	Profibrogenic	-6 G $\rightarrow$ A polymorphism is associated with more severe fibrosis	28

**Table II.** Candidate genes for future studies.

Candidate gene	Protein function	Liver disease	Reference
LDL receptor	Lipid metabolism	HCV	30
ApoE	Lipid metabolism	HCV	18,31
CPT-1A	Lipid metabolism	HCV	32
MTHFR	Lipid metabolism	HCV	33
GSTM1	Oxidative stress	ALD	34,35
GSTT1	Oxidative stress	ALD	35
GSTP1	Oxidative stress	HHC	36
MPO	Oxidative stress	HCV, HHC	37,38
MEH	Oxidative stress	HCV	39
IL-1Ra	Proinflammatory	PBC, ALD, HCV	40,41,42
TNF- $\beta$	Proinflammatory	HCV	43
IL-12	Proinflammatory	HCV	44
IFN- $\gamma$	Proinflammatory	HCV	45
IL-10	Antiinflammatory	ALD, HCV	46,47,48
IL-10 receptor	Antiinflammatory	HCV	49
MCP-1	Chemotaxis	HCV	50
MCP-2	Chemotaxis	HCV	51
CCR5	Chemotaxis	HCV	18,51
RANTES	Chemotaxis	HCV	51,52
CTLA-4	Immune response	ALD, HCV	18,53
MMP-1,3,9	Matrix turnover	PSC, HCV	54,55
PDGF	Profibrogenic	HCV post-LTX	56
Keratins	Cytoskeleton	Cryptogenic cirrhosis, end stage liver disease, ALD, HCV	57,58,59,60

expect that this also holds true vice versa. However, similar to other chronic liver diseases discrepant data concerning the contribution of HFE gene mutation to the severity of NAFLD exist.<sup>13</sup> While two studies showed that heterozygosity for the C282Y mutation is associated with more severe hepatic fibrosis,<sup>21,22</sup> three subsequent studies failed to confirm these results.<sup>3,23,24</sup>

A number of studies have provided evidence that polymorphisms of other genes involved in the generation or degradation of ROS such as glutathione-S-transferase-isoforms (GSTM1, GSTT1, GSTP1), myeloperoxidase (MPO) or microsomal epoxide hydrolase (MEH) affect progression to cirrhosis in different liver disease such as ALD, chronic HCV infection or HHC. Given the key role of FFA oxidation and ROS generation in the course of hepatocyte death in NAFLD these polymorphisms should be investigated in the future (*see table II*).

### Cytokine gene polymorphisms

TNF- $\alpha$  plays a major role in the pathogenesis of NAFLD, as increased TNF- $\alpha$  levels favor the development of insulin resistance and impaired glucose tolerance. A promoter polymorphism of the TNF- $\alpha$  gene associated with increased cytokine expression was suggested to affect susceptibility towards NAFLD and to be associated with higher insulin resistance indices and a higher prevalence of impaired glucose tolerance.<sup>25</sup> A Japanese study reported that the frequencies of the IL- $\beta$  -511 T allele and the T/T genotype are significantly higher in NASH patients than in controls providing further evidence for the involvement of cytokine gene polymorphism in the course of NAFLD.<sup>14</sup>

Chronic inflammation and a mixed cellular infiltrate are key characteristics of NASH. It is therefore surprising that TNF- $\alpha$  and IL-1 $\beta$  are the only cytokine gene polymorphisms that have been evaluated in NAFLD so far. *Table II* summarizes data from studies focusing on the role of genetic variants of cytokines, chemokines, their receptors and other proteins involved in immune responses in other liver diseases that might also play a role in the progression from steatosis to NASH and more advanced stages of NAFLD.

### Genes coding for bacterial receptors

There is evidence that gut derived endotoxin leading to cytokine release contributes to the pathogenesis of NAFLD. CD14 is a co-receptor for gram negative bacterial lipopolysaccharide (LPS) expressed on monocytes, macrophages but also on other cells. CD14 lacks an intracellular domain but enhances signaling through Toll-like receptor-4 (TLR4). Expression of CD14 has been suggested to be influenced by a polymorphism within the promoter region. Similar to patients with ALD,<sup>26</sup> the T allele was suggested to confer increased risk for the progression from steatosis to NASH.<sup>27</sup> The Asp299Gly poly-

morphism in the TLR4 gene has been associated with endotoxin hyporesponsiveness in humans. This polymorphism definitely deserves to be tested in larger cohorts of patients with NAFLD.

### Genes influencing extracellular matrix synthesis and degradation

Hepatic stellate cells (HSC) are believed to be the major collagen-producing cells in liver fibrosis. Upon activation by cytokines and other agonists, HSC acquire a profibrogenic phenotype and synthesize extracellular matrix ECM that replaces parenchymal tissue. Consequently, variations of genes that play a crucial role in the activation of HSCs should affect progression to cirrhosis irrespective of etiology and the underlying cause of liver disease. One would expect that variations of these genes affect progression to cirrhosis in a uniform and global way and are therefore primary candidates to be studied in NAFLD-related fibrosis. The only relevant study so far in this regard in NAFLD reported that the combination of high angiotensinogen and TGF- $\beta$ 1 producing polymorphisms is associated with advanced hepatic fibrosis in obese patients with NAFLD<sup>28</sup> which is in accordance with findings in patients with chronic HCV infection.<sup>29</sup>

Genetic variation of the platelet derived growth factor (PDGF), a key molecule in liver fibrosis driving HSC proliferation, has been reported to affect the course of HCV infection after liver transplantation. So far data is lacking if this polymorphism also contributes to the course of other chronic liver diseases including NAFLD. Matrix metalloproteinases (MMP) constitute another group of attractive candidate genes to be studied in NAFLD as they have been shown to affect the course of HCV virus infection and primary sclerosing cholangitis (PSC) by interfering with adequate matrix turn-over. Mallory body formation is a characteristic histopathological feature of NASH that results from misfolding and aggregation of keratins. There is convincing evidence that mutations of keratins can cause cryptogenic cirrhosis and affect the course of liver diseases of different etiology (*see table II*). Results from studies investigating the role of keratin mutations in NAFLD are anticipated with great interest.

### Conclusion

NAFLD has become the leading cause of referral to hepatology clinics and a major future challenge. Data from family studies and interethnic differences in susceptibility suggest that genetic factors contribute to the variability seen in natural course of NAFLD. Identifying risk genes that predict faster fibrosis progression or susceptibility for HCC development might help managing patients in an individual way. However, data from gene association studies carried out in patients with NAFLD are inconclusive. Most published studies were carried out in

either small cohorts of patients carrying the risk of type I error or with inaccurate study design. Available reports need to be verified by subsequent larger studies. A number of other candidate genes that have been suggested to contribute to fibrosis progression in other chronic liver diseases have not been evaluated. Carefully designed studies according to recently defined guidelines<sup>13</sup> requiring collaboration of different research centers will help to understand the role of genetics of NAFLD.

## Acknowledgements

Christoph H. Österreicher is supported by an Erwin Schroedinger research fellowship kindly provided by the Austrian Science Fund (FWF). David A. Brenner is supported by grants from the NIH.

## References

- Brunt EM. Nonalcoholic steatohepatitis. *Semin Liver Dis* 2004; 24: 3-20.
- Farrell GC, Larter CZ. Nonalcoholic fatty liver disease: from steatosis to cirrhosis. *Hepatology* 2006; 43: S99-S112.
- Bugianesi E, Leone N, Vanni E, Marchesini G, Brunello F, Carucci P, Musso A, De Paolis P, Capussotti L, Salizzoni M, Rizzetto M. Expanding the natural history of nonalcoholic steatohepatitis: from cryptogenic cirrhosis to hepatocellular carcinoma. *Gastroenterology* 2002; 123: 134-40.
- Wanless IR, Lentz JS. Fatty liver hepatitis (steatohepatitis) and obesity: an autopsy study with analysis of risk factors. *Hepatology* 1990; 12: 1106-10.
- Dixon JB, Bhathal PS, O'Brien PE. Nonalcoholic fatty liver disease: predictors of nonalcoholic steatohepatitis and liver fibrosis in the severely obese. *Gastroenterology* 2001; 121: 91-100.
- Battaller R, Brenner DA. Liver fibrosis. *J Clin Invest* 2005; 115: 209-18.
- Day CP. From fat to inflammation. *Gastroenterology* 2006; 130: 207-10.
- Xu A, Wang Y, Keshaw H, Xu LY, Lam KS, Cooper GJ. The fat-derived hormone adiponectin alleviates alcoholic and nonalcoholic fatty liver diseases in mice. *J Clin Invest* 2003; 112: 91-100.
- Willner IR, Waters B, Patil SR, Reuben A, Morelli J, Riely CA. Ninety patients with nonalcoholic steatohepatitis: insulin resistance, familial tendency, and severity of disease. *Am J Gastroenterol* 2001; 96: 2957-61.
- Struben VM, Hespeneide EE, Caldwell SH. Nonalcoholic steatohepatitis and cryptogenic cirrhosis within kindreds. *Am J Med* 2000; 108: 9-13.
- Browning JD, Kumar KS, Saboorian MH, Thiele DL. Ethnic differences in the prevalence of cryptogenic cirrhosis. *Am J Gastroenterol* 2004; 99: 292-8.
- Stickel F, Österreicher CH. The role of genetic polymorphisms in alcoholic liver disease. *Alcohol Alcohol* 2006; 41: 209-24.
- Österreicher CH, Stickel F, Brenner DA. Genomics of liver fibrosis and cirrhosis. *Semin Liver Dis* 2007; 27: 28-43.
- Nozaki Y, Saibara T, Nemoto Y, Ono M, Akisawa N, Iwasaki S, Hayashi Y, Hiroi M, Enzan H, Onishi S. Polymorphisms of interleukin-1 beta and beta 3-adrenergic receptor in Japanese patients with nonalcoholic steatohepatitis. *Alcohol Clin Exp Res* 2004; 28: 106S-110S.
- Iwamoto N, Ogawa Y, Kajihara S, Hisatomi A, Yasutake T, Yoshimura T, Mizuta T, Hara T, Ozaki I, Yamamoto K. Gln27Glu beta2-adrenergic receptor variant is associated with hypertriglyceridemia and the development of fatty liver. *Clin Chim Acta* 2001; 314: 85-91.
- Bernard S, Touzet S, Personne I, Lapras V, Bondon PJ, Berthezene F, Moulin P. Association between microsomal triglyceride transfer protein gene polymorphism and the biological features of liver steatosis in patients with type II diabetes. *Diabetologia* 2000; 43: 995-9.
- Namikawa C, Shu-Ping Z, Vyselaar JR, Nozaki Y, Nemoto Y, Ono M, Akisawa N, Saibara T, Hiroi M, Enzan H, Onishi S. Polymorphisms of microsomal triglyceride transfer protein gene and manganese superoxide dismutase gene in non-alcoholic steatohepatitis. *J Hepatol* 2004; 40: 781-6.
- Richardson MM, Powell EE, Barrie HD, Clouston AD, Purdie DM, Jonsson JR. A combination of genetic polymorphisms increases the risk of progressive disease in chronic hepatitis C. *J Med Genet* 2005; 42: e45.
- Song J, da Costa KA, Fischer LM, Kohlmeier M, Kwock L, Wang S, Zeisel SH. Polymorphism of the PEMT gene and susceptibility to nonalcoholic fatty liver disease (NAFLD). *Faseb J* 2005; 19: 1266-71.
- Powell EE, Ali A, Clouston AD, Dixon JL, Lincoln DJ, Purdie DM, Fletcher LM, Powell LW, Jonsson JR. Steatosis is a cofactor in liver injury in hemochromatosis. *Gastroenterology* 2005; 129: 1937-43.
- George DK, Goldwurm S, MacDonald GA, Cowley LL, Walker NI, Ward PJ, Jazwinska EC, Powell LW. Increased hepatic iron concentration in nonalcoholic steatohepatitis is associated with increased fibrosis. *Gastroenterology* 1998; 114: 311-8.
- Bonkovsky HL, Jawaid Q, Tortorelli K, LeClair P, Cobb J, Lambrecht RW, Banner BF. Non-alcoholic steatohepatitis and iron: increased prevalence of mutations of the HFE gene in non-alcoholic steatohepatitis. *J Hepatol* 1999; 31: 421-9.
- Chitturi S, Weltman M, Farrell GC, McDonald D, Kench J, Liddle C, Samarasinghe D, Lin R, Abeygunasekera S, George J. HFE mutations, hepatic iron, and fibrosis: ethnic-specific association of NASH with C282Y but not with fibrotic severity. *Hepatology* 2002; 36: 142-9.
- Deguti MM, Sipahi AM, Gayotto LC, Palacios SA, Bittencourt PL, Goldberg AC, Laudanna AA, Carrilho FJ, Cancado EL. Lack of evidence for the pathogenic role of iron and HFE gene mutations in Brazilian patients with nonalcoholic steatohepatitis. *Braz J Med Biol Res* 2003; 36: 739-45.
- Valenti L, Fracanzani AL, Dongiovanni P, Santorelli G, Branchi A, Taioli E, Fiorelli G, Fargion S. Tumor necrosis factor alpha promoter polymorphisms and insulin resistance in nonalcoholic fatty liver disease. *Gastroenterology* 2002; 122: 274-80.
- Jarvelainen HA, Orpana A, Perola M, Savolainen VT, Karhunen PJ, Lindros KO. Promoter polymorphism of the CD14 endotoxin receptor gene as a risk factor for alcoholic liver disease. *Hepatology* 2001; 33: 1148-53.
- Brun P, Castagliuolo I, Floreani AR, Buda A, Blasone L, Palu G, Martines D. Increased risk of NASH in patients carrying the C(-159)T polymorphism in the CD14 gene promoter region. *Gut* 2006; 55: 1212.
- Dixon JB, Bhathal PS, Jonsson JR, Dixon AF, Powell EE, O'Brien PE. Pro-fibrotic polymorphisms predictive of advanced liver fibrosis in the severely obese. *J Hepatol* 2003; 39: 967-71.
- Powell EE, Edwards-Smith CJ, Hay JL, Clouston AD, Crawford DH, Shorthouse C, Purdie DM, Jonsson JR. Host genetic factors influence disease progression in chronic hepatitis C. *Hepatology* 2000; 31: 828-33.
- Hennig BJ, Hellier S, Frodsham AJ, Zhang L, Klenerman P, Knapp S, Wright M, Thomas HC, Thursz M, Hill AV. Association of low-density lipoprotein receptor polymorphisms and outcome of hepatitis C infection. *Genes Immun* 2002; 3: 359-67.
- Wozniak MA, Itzhaki RF, Faragher EB, James MW, Ryder SD, Irving WL. Apolipoprotein E-epsilon4 protects against severe liver disease caused by hepatitis C virus. *Hepatology* 2002; 36: 456-63.
- Huang H, Shiffman ML, Cheung RC, Layden TJ, Friedman S, Abar OT, Yee L, Chokkalingam AP, Schrodi SJ, Chan J, Catanese JJ, Leong DU, Ross D, Hu X, Monto A, McAllister

- LB, Broder S, White T, Sninsky JJ, Wright TL. Identification of two gene variants associated with risk of advanced fibrosis in patients with chronic hepatitis C. *Gastroenterology* 2006; 130: 1679-87.
33. Adinolfi LE, Inghosso D, Cesaro G, Cimmino A, D'Anto M, Capasso R, Zappia V, Ruggiero G. Hyperhomocysteinemia and the MTHFR C677T polymorphism promote steatosis and fibrosis in chronic hepatitis C patients. *Hepatology* 2005; 41: 995-1003.
  34. Savolainen VT, Pjarinen J, Perola M, Penttila A, Karhunen PJ. Glutathione-S-transferase GST M1 «null» genotype and the risk of alcoholic liver disease. *Alcohol Clin Exp Res* 1996; 20: 1340-5.
  35. Ladero JM, Martinez C, Garcia-Martin E, Fernandez-Arquero M, Lopez-Alonso G, de la Concha EG, Diaz-Rubio M, Agundez JA. Polymorphisms of the glutathione S-transferases mu-1 (GSTM1) and theta-1 (GSTT1) and the risk of advanced alcoholic liver disease. *Scand J Gastroenterol* 2005; 40: 348-53.
  36. Stickel F, Osterreicher CH, Datz C, Ferenci P, Wolfel M, Norgauer W, Kraus MR, Wrba F, Hellerbrand C, Schuppan D. Prediction of progression to cirrhosis by a glutathione S-transferase P1 polymorphism in subjects with hereditary hemochromatosis. *Arch Intern Med* 2005; 165: 1835-40.
  37. Reynolds WF, Patel K, Pianko S, Blatt LM, Nicholas JJ, McHutchison JG. A genotypic association implicates myeloperoxidase in the progression of hepatic fibrosis in chronic hepatitis C virus infection. *Genes Immun* 2002; 3: 345-9.
  38. Osterreicher CH, Datz C, Stickel F, Hellerbrand C, Penz M, Hofer H, Wrba F, Penner E, Schuppan D, Ferenci P. Association of myeloperoxidase promoter polymorphism with cirrhosis in patients with hereditary hemochromatosis. *J Hepatol* 2005; 42: 914-9.
  39. Sonzogni L, Silvestri L, De Silvestri A, Gritti C, Foti L, Zavaglia C, Bottelli R, Mondelli MU, Civardi E, Silini EM. Polymorphisms of microsomal epoxide hydrolase gene and severity of HCV-related liver disease. *Hepatology* 2002; 36: 195-201.
  40. Donaldson P, Agarwal K, Craggs A, Craig W, James O, Jones D. HLA and interleukin 1 gene polymorphisms in primary biliary cirrhosis: associations with disease progression and disease susceptibility. *Gut* 2001; 48: 397-402.
  41. Chen WX, Xu GY, Yu CH, Wang P, Yu M, Xu P, Liu YS, Li YM. Correlation of polymorphism in the interleukin-1 receptor antagonist gene intron 2 with alcoholic liver disease. *Hepatobiliary Pancreat Dis Int* 2005; 4: 41-5.
  42. Bahr MJ, el Menuawy M, Boeker KH, Musholt PB, Manns MP, Lichtinghagen R. Cytokine gene polymorphisms and the susceptibility to liver cirrhosis in patients with chronic hepatitis C. *Liver Int* 2003; 23: 420-5.
  43. Goyal A, Kazim SN, Sakhujia P, Malhotra V, Arora N, Sarin SK. Association of TNF-beta polymorphism with disease severity among patients infected with hepatitis C virus. *J Med Virol* 2004; 72: 60-5.
  44. Suneetha PV, Goyal A, Hissar SS, Sarin SK. Studies on TAQ1 polymorphism in the 3'untranslated region of IL-12P40 gene in HCV patients infected predominantly with genotype 3. *J Med Virol* 2006; 78: 1055-60.
  45. Dai CY, Chuang WL, Hsieh MY, Lee LP, Hou NJ, Chen SC, Lin ZY, Hsieh MY, Wang LY, Tsai JF, Chang WY, Yu ML. Polymorphism of interferon-gamma gene at position +874 and clinical characteristics of chronic hepatitis C. *Transl Res* 2006; 148: 128-33.
  46. Grove J, Daly AK, Bassendine MF, Gilvarry E, Day CP. Interleukin 10 promoter region polymorphisms and susceptibility to advanced alcoholic liver disease. *Gut* 2000; 46: 540-5.
  47. Knapp S, Hennig BJ, Frodsham AJ, Zhang L, Hellier S, Wright M, Goldin R, Hill AV, Thomas HC, Thursz MR. Interleukin-10 promoter polymorphisms and the outcome of hepatitis C virus infection. *Immunogenetics* 2003; 55: 362-9.
  48. Paladino N, Fainboim H, Theiler G, Schroder T, Munoz AE, Flores AC, Galdame O, Fainboim L. Gender susceptibility to chronic hepatitis C virus infection associated with interleukin 10 promoter polymorphism. *J Virol* 2006; 80: 9144-50.
  49. Hofer H, Neufeld JB, Oesterreicher C, Grundtner P, Wrba F, Gangl A, Ferenci P, Gasche C. Bi-allelic presence of the interleukin-10 receptor 1 G330R allele is associated with cirrhosis in chronic HCV-1 infection. *Genes Immun* 2005; 6: 242-7.
  50. Muhlbauer M, Bosserhoff AK, Hartmann A, Thasler WE, Weiss TS, Herfarth H, Lock G, Scholmerich J, Hellerbrand C. A novel MCP-1 gene polymorphism is associated with hepatic MCP-1 expression and severity of HCV-related liver disease. *Gastroenterology* 2003; 125: 1085-93.
  51. Hellier S, Frodsham AJ, Hennig BJ, Klenerman P, Knapp S, Ramaley P, Satsangi J, Wright M, Zhang L, Thomas HC, Thursz M, Hill AV. Association of genetic variants of the chemokine receptor CCR5 and its ligands, RANTES and MCP-2, with outcome of HCV infection. *Hepatology* 2003; 38: 1468-76.
  52. Promrat K, McDermott DH, Gonzalez CM, Kleiner DE, Koziol DE, Lessie M, Merrell M, Soza A, Heller T, Ghany M, Park Y, Alter HJ, Hoofnagle JH, Murphy PM, Liang TJ. Associations of chemokine system polymorphisms with clinical outcomes and treatment responses of chronic hepatitis C. *Gastroenterology* 2003; 124: 352-60.
  53. Valenti L, De Feo T, Fracanzani AL, Fatta E, Salvagnini M, Arico S, Rossi G, Fiorelli G, Fargion S. Cytotoxic T-lymphocyte antigen-4 A49G polymorphism is associated with susceptibility to and severity of alcoholic liver disease in Italian patients. *Alcohol Alcohol* 2004; 39: 276-80.
  54. Satsangi J, Chapman RW, Haldar N, Donaldson P, Mitchell S, Simmons J, Norris S, Marshall SE, Bell JI, Jewell DP, Welsh KI. A functional polymorphism of the stromelysin gene (MMP-3) influences susceptibility to primary sclerosing cholangitis. *Gastroenterology* 2001; 121: 124-30.
  55. Okamoto K, Mimura K, Murawaki Y, Yuasa I. Association of functional gene polymorphisms of matrix metalloproteinase (MMP)-1, MMP-3 and MMP-9 with the progression of chronic liver disease. *J Gastroenterol Hepatol* 2005; 20: 1102-8.
  56. Ben-Ari Z, Tambur AR, Pappo O, Sulkes J, Pravica V, Hutchinson I, Klein T, Tur-Kaspa R, Mor E. Platelet-derived growth factor gene polymorphism in recurrent hepatitis C infection after liver transplantation. *Transplantation* 2006; 81: 392-7.
  57. Ku NO, Gish R, Wright TL, Omary MB. Keratin 8 mutations in patients with cryptogenic liver disease. *N Engl J Med* 2001; 344: 1580-7.
  58. Ku NO, Darling JM, Krams SM, Esquivel CO, Keeffe EB, Sibley RK, Lee YM, Wright TL, Omary MB. Keratin 8 and 18 mutations are risk factors for developing liver disease of multiple etiologies. *Proc Natl Acad Sci U S A* 2003; 100: 6063-8.
  59. Ku NO, Lim JK, Krams SM, Esquivel CO, Keeffe EB, Wright TL, Parry DA, Omary MB. Keratins as susceptibility genes for end-stage liver disease. *Gastroenterology* 2005; 129: 885-93.
  60. Strnad P, Lienau TC, Tao GZ, Lazzeroni LC, Stickel F, Schuppan D, Omary MB. Keratin variants associate with progression of fibrosis during chronic hepatitis C infection. *Hepatology* 2006; 43: 1354-63.