

Storage-induced morphological changes in erythrocytes

Raúl Carrillo-Esper,* Carlos Alberto Carrillo-Córdova,**
Jorge Raúl Carrillo-Córdova,** Luis Daniel Carrillo-Córdova**

RESUMEN

Introducción. Los avances en las técnicas de almacenamiento eritrocitario permiten utilizar concentrados eritrocitarios que presentan importantes anomalías morfológicas asociadas a mayor lesión en la microcirculación, disfunción orgánica múltiple e incremento en la morbimortalidad. **Objetivo.** Evaluar las modificaciones de la morfología eritrocitaria relacionadas con el tiempo de almacenamiento. **Material y métodos.** Se analizaron los cambios morfológicos eritrocitarios en relación con el tiempo de almacenamiento de los concentrados eritrocitarios transfundidos en la Unidad de Terapia Intensiva de la Fundación Clínica Médica Sur mediante la evaluación del frotis de una muestra obtenida de los concentrados eritrocitarios obtenidos. Cada uno de los frotis se tiñó con tinción de Wright; dos observadores independientes evaluaron el porcentaje de alteraciones morfológicas eritrocitarias. El tiempo de almacenamiento se definió como el lapso entre la extracción de la sangre y la transfusión del concentrado eritrocitario. **Resultados.** Se recolectaron 67 muestras. El promedio de anomalías eritrocitarias fue de 40%. La media del tiempo de almacenamiento fue de 12.25 ± 6.9 días. El coeficiente de correlación de Pearson demostró correlación entre el porcentaje de anomalías eritrocitarias y los días de almacenamiento. Después de siete días de almacenamiento 26.57% de los eritrocitos de los concentrados eritrocitarios tuvieron anomalías morfológicas eritrocitarias que se incrementaron significativamente en los siguientes días. **Conclusión.** Los eritrocitos que rebasan siete días de almacenamiento presentan alteraciones eritrocitarias significativas.

Palabras clave. Transfusión sanguínea. Anomalías eritrocitarias.

ABSTRACT

Introduction. Advances in storage techniques of erythrocyte concentrates have contributed to develop significant erythrocyte morphologic abnormalities which have been associated with greater damage in the microcirculation, multiple organ dysfunction and increased morbidity and mortality. **Objective.** To evaluate the changes in erythrocyte morphology related to storage time. **Material and methods.** We analyzed the morphologic changes in erythrocyte in relation to the storage time of the globular packages that were transfused. The blood smear was wisdom with Wright's technique and was evaluated by two observers who evaluated the percentage of morphologic alterations. **Results.** 67 samples were collected. The average of abnormalities in the erythrocyte package was 40%. The storage time was of 12.25 ± 6.9 days. The Pearson correlation coefficient demonstrated correlation between the percentages of erythrocyte abnormalities and storage time. After seven days of storage 26.57% of the erythrocytes of the packages had morphological abnormalities which increased significantly in the following days. **Conclusions.** Red blood cells with more than seven days of storage, present significant morphologic alterations, erythrocyte abnormalities.

Key words. Blood transfusion. Abnormal erythrocytes.

INTRODUCTION

Recent investigations in the process of blood transfusion have focused on studying the morphologic changes secondary to conservation and storage in erythrocytes, these morphologic changes are named "storage lesions"

and are related with erythrocyte damage and decrease of the survival after the transfusion. The efforts and investigations made during the last 40 years to maintain the corpuscular integrity and the viability and function after the transfusion have increased the storage time to up to 42 days, this was achieved thanks to the phosphate, ade-

* Unidad de Terapia Intensiva, Fundación Clínica Médica Sur. ** Facultad de Medicina, UNAM.

Correspondencia:

Dr. Raúl Carrillo-Esper

Unidad de Terapia Intensiva, Fundación Clínica Médica Sur. Puente de Piedra, Núm. 150. Col. Toriello Guerra, C.P. 14050.
Tel.: 5424-7200. Correo electrónico: rcarrillo@medicasur.org.mx

a new solution for preservation of RBC, this is the today well known ACD solution (Acid-citrate-dextrose). ACD solution simplified the sterilization procedure, decreased the volumen of the conservative solution and increased viability of RBC up to 21 days. On the other side, addition of phosphate and 2,3 diphosphoglycerate allow to increase the storage amount of days up to 42 and also increased the viability.⁴⁻⁶

The main result of this investigation is to demonstrate that there is a direct relation between storage time and percentage of abnormalities in the RBC. There is enough scientific evidence that storage time from the transfused RBC during transoperatory or during their stay in the intensive care unit is associated with an increased risk of death, more complications and more hospital stay.⁷⁻⁹

It is known that transfusion of RBC with a long storage time, which have energetic depletion, membrane alterations and distortion, amplify the inflammatory response

when they damage and obstruct the vascular endothelium, this induce more microcirculation alteration and organic dysfunction^{10,11} (Figure 2).

Red blood cell characteristics and changes during storage are characterized by (Table 1):

- **Morphologic changes.** From their normal biconcave disc form, RBC deform during the pass of the storage days. The most important form are the crenocytes which are spheric erythrocytes with spicules, this spicules correspond to lipidic vesicles that protrude from the membrane. This transformation is associated with a loss of the surface/volume of the RBC, this increases the mean corpuscular heamoglobin concentration and osmotic fragility, this leads to a decrease in the capacity of the RBC to deform. There is some evidence that all this morphologic changes correlates with the ATP depletion of the RBC.¹² Nakao¹³ reproduced these

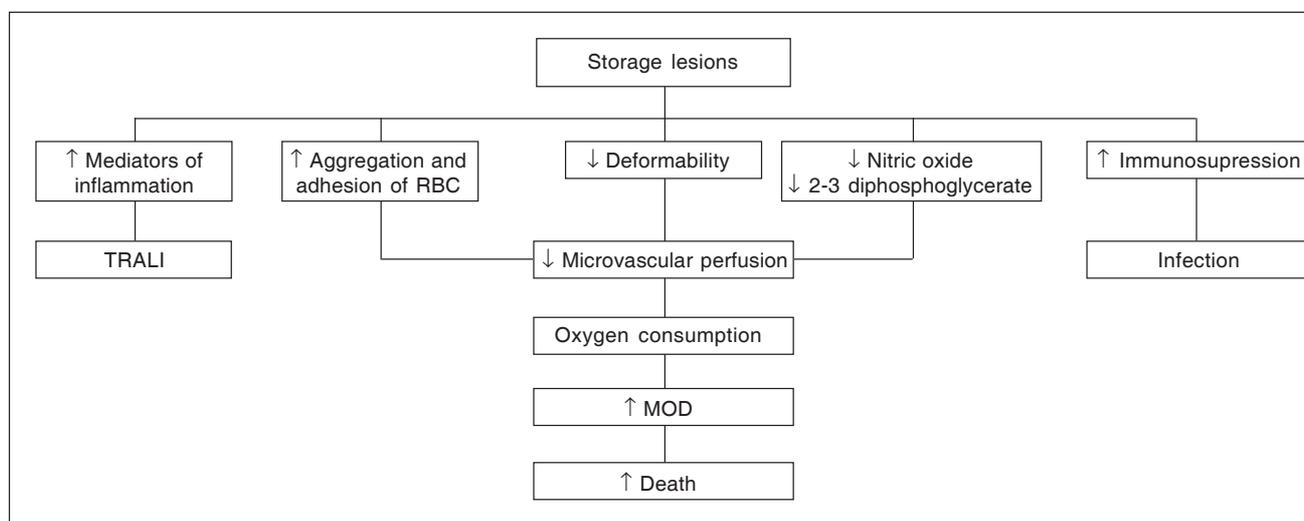


Figure 2. Effects of the red blood cell storage. TRALI: transfusion related acute lung injury. MOD: multiple organ dysfunctions.

Table 1. Changes during storage.

Effects during the storage	Consequences
Decrease 2,3 diphosphoglycerate	Increase of the affinity to oxygen and decrease of tissular contribution.
Depletion of ATP	Changes on the erythrocytes morphology. Increase of the osmotic fragility. Decrease of the deformability.
Microvesiculation	Decrease of the erythrocytes viability.
Lipidic peroxidation	Celular damage or death.

morphologic changes secondary to prolonged storage. He also showed that reestablishing the ATP levels with adenosine made the RBC to obtain their normal biconcave structure after being crenocytes.

- **ATP levels.** It is considered that biomechanical surface changes of RBC and cytoplasmic viscosity occurs after the decrease of ATP. There is a low correlation between RBC ATP levels and surviving of RBC, except when ATP levels decrease > 50%. This inconsistency emphasizes in studies that very high levels of ATP have no impact in RBC surviving post-transfusion. Even though ATP depletion does not explain by itself the membrane damage, it has been thought that depletion of ATP increases the levels of secondary mediators like intracellular calcium, protein phosphorylation and membrane kinases which maintain the integrity of cell membrane. The phospholipids analysis shows a change in the phosphorylation/dephosphorylation balance in the RBC during the second and third storage week. The increase in the phosphorylated form of the phosphoinositol-4-phosphate, is accompanied with an appearance of crenocytes, this suggests a direct relation between dephosphorylation and change in the morphology of RBC.^{14,15}
- **Biomechanical changes.** There is a complex interaction between the membrane phospholipids, transmembrane proteins and cytoplasmic components which has an impact in the morphology and deformability of the RBC during storage. This modifications decrease the RBC half life post-transfusion. RBC biomechanical changes are the result of the alterations of the lipidic bilayer, proteins and cytoskeleton. The interactions between these 3 components favor the formation of 20 to 80 nm vesicles, which could be detected by the second storage week. Oxidative damage is another important mechanism of RBC damage, this impacts the biomechanical by damaging the phospholipids of the membrane and the spectrine, this contributes to the formation of squistocytes and spherocytes, also contributes to the osmotic fragility.¹⁶⁻¹⁸

Changes in the RBC during the storage produce RBC with low efficacy and efectivity to improve oxygenation in tissues, this can be related with a depletion of the 2-3 diphosphoglycerate (DPG). This 2-3 DPG depletion is bigger within the seventh day of storage, this implies a shift to the left in the haemoglobin curve with a decrease of the p50.

The clinical impact of the RBC changes induced with the storage reverberates in the poor effectiveness of these

to improve tissue oxygenation this is related with de decrease of 2-3 diphosphoglycerate. This depletion is the mayor alosteric modificador over the affinity to haemoglobin. The deformability reduction of the RBC during storage has been explained like of the physiopathological mechanisms in the development of tissue lesion induced by the stored RBC.^{19,20}

The main objective of RBC transfusions in the critically ill patient is to improve oxygen disponibility in tissues and increase survival and organic function. All this lead to the wrong idea of tranfusing all the critically ill patients and maintain haemoglobin values > 13 g/dL, which is wrong. Different studies has demonstrated that this is not truth and using transfusions in a bad way or abusing of them increases in a significative way complications, specially infections, and morbimortality, specially when the transfused packages has more that 7-10 days of storage.

Based in this, current concepts of RBC transfusions indications is to limit the number of transfusions, follow rigid transfusion politics, set transfusion cutting points in haemoglobin 7-10 g/dL, try to use new RBC, utilize leucoreduced packages in patients with sepsis and when there is evidence of microcirculation damage and trying to avoid the transfusions of RBC with more than 7 days of storage.²¹⁻²⁴

CONCLUSIONS

The morphologic, biological and molecular changes induced by the storage of erythrocytes are subjects of basic and clinical investigation because of their effects in transfused patients. The scientific evidence shows that the erythrocytes with a storage time grater than 14 days induce lesions on the microcirculation and don't increase the consumption of oxygen of cells, which favours the development of multiple organ dysfunctions. Because of that is necessary to develop a specific politic about the transfusions in the ICU's where the use of erythrocyte concentrates with shorter storage life is priority, especially with patients with sepsis and organ or microcirculatory dysfunction.

REFERENCES

1. Wolfe LC. The membrana and the lesions of storage in preserved red cells. *Transfusion* 1985; 25: 185-203.
2. Tinmouth A, Chin-Yee I. The clinical consequences of the red cell storage lesion. *Transfus Med Rev* 2001; 15: 91-107.
3. Ho J, Sibbald WJ, Chin-Yee IH. Effects of storage on efficacy of red cell transfusion: when is it not safe? *Crit Care Med* 2003; 31: 687-97.
4. Loutit JF, Mollison PL, Young JM. Citric acid-sodium citrate-glucose mixtures for blood storage. *Q F Exp Physiol* 1943; 32: 183-202.

5. Heaton A, Miripol J, Aster R. Use of Adsol preservation solution for prolonged storage of low viscosity AS-1 red blood cells. *Br J Haematol* 1984; 57: 467-78.
6. Simon TL, Marcus CS, Mythe BA. Effects of AS-3 nutrient-additive solution on 42 and 49 days storage of red cells. *Transfusion* 1987; 27: 178-82.
7. Kuduvalli M, Oo AY, Newall N. Effect of peri-operative red blood cell transfusion on 30 day and 1 year mortality following coronary artery bypass surgery. *Eur J Cardiothorac Surg* 2005; 27: 592-8.
8. Purdy FR, Tweeddale MG, Merrick PM. Association of mortality with age of blood transfused in septic ICU patients. *Can J Anaesth* 1997; 44: 1256-61.
9. Martin CM, Sibbald WJ, Lu X. Age of transfused blood is an independent risk factor for postinjury multiple organ failure. *Am J Surg* 1999; 178: 570-2.
10. Card RT. Red Cell membrane changes during storage. *Trans Med Rev* 1988; 2: 40-7.
11. Chin-Yee I, Arya N, d'Almeida MS. The red cell storage lesion and its implication for transfusion. *Transfus Sci* 1997; 18: 447-58.
12. Card RT, Mohandas N, Perkins HA. Deformability of stored red blood cells: relationship to degree of packing. *Transfusion* 1982; 22: 96-101.
13. Nakao M, Nakao T, Yamazoe S. Adenosine triphosphate and maintenance of shape of human red cells. *Nature* 1960; 187: 945-7.
14. Swietochowska K, Piascik R, Jaroszewicz K. Human stored blood inositol phospholipids. *Acta Physiol* 1991; 78: 283-91.
15. Dern RJ, Brewer GJ, Wiorkowski JJ. Studies of the preservation of human blood. The relationship of erythrocyte adenosine triphosphate levels. *J Lab Clin Med* 1967; 69: 968-78.
16. Greenwalt TJ, Bryan DJ, Dumaswala UJ. Erythrocyte membrane vesiculation and changes in membrane composition during storage in citrate-phosphate-dextrose-adenine. *Vox Sang* 1984; 47: 261-70.
17. Brunauer LS, Moxness MS, Huestis WH. Hydrogen peroxide oxidation induces the transfer of phospholipids from the membrane into the cytosol of human erythrocytes. *Biochemistry* 1994; 33: 4527-32.
18. Wagner GM, Chiu DT, Qju JH. Spectrin oxidation correlates with membrane vesiculation in stored RBC's. *Blood* 1987; 69: 1777-81.
19. Heaton A, Keegan T, Holme S. In vivo regeneration of red blood cell 2,3-diphosphoglycerate following transfusion of DPG depleted AS-1, AS-3 and CPDA-1 red blood cells. *Br J Haematol* 1969; 71: 131-6.
20. Bunn HF, May MH, Kocholaty WF. Hemoglobin function in stored blood. *J Clin Invest* 1969; 48: 311-21.
21. Hebert PC, Wells G, Blajchman MA. A multicenter, randomized, controlled clinical trial of transfusion requirements in critical care. Transfusion Requirements in Critical Care Investigators. *N Engl J Med* 1999; 340: 409-17.
22. Vincent JL, Baron JF, Reinhart K. Anemia Blood transfusion critically ill patients. *JAMA* 2002; 288: 1499-507.
23. Corwin HL, Gettinger A, Pearl RG. The CRIT study: Anemia and Blood Transfusion in the Critically Ill. Current Practice in the United States. *Crit Care Med* 2004; 32: 39-52.
24. Practice Guidelines for the Blood Component Therapy. A Report by the American Society of Anesthesiologists Task Force on Blood Component Therapy. *Anesthesiology* 1996; 84: 732-47.