Doi:10.31157/an.v27i2.326

Bioinformatic Analysis of Epigenomic Studies for Major Depressive Disorder

Alam Farwah B. 1 | González-Giraldo Yeimy 2 | Forero Diego A.3,4™

- 1. North South University, Dhaka, Bangladesh
- 2. Center for Psychosocial Studies for Latin America and the Caribbean, Universidad Antonio Nariño, Bogotá, Colombia.
- 3. Health and Sport Sciences Research Group, School of Health and Sport Sciences, Fundación Universitaria del Área Andina, Bogotá, Colombia.
- 4. Professional Program in Respiratory Therapy, School of Health and Sport Sciences, Fundación Universitaria del Área Andina, Bogotá, Colombia.

Correspondence

Prof. Dr. Diego Forero, MD, PhD. Professor and Research Leader, School of Health and Sport Sciences, Fundación Universitaria del Área Andina, Bogotá, Colombia.

dforero41@areandina.edu.co

Abstract

Background: Major depressive disorder (MDD) is a common psychiatric entity, being characterized by alterations in mood and in other clinical dimensions. Several epigenome-wide association studies (EWAS) for MDD have been published. Here, we aimed to identify common genes in EWAS and their convergence with multiple lines of genomic evidence. Methods: We carried out a computational analysis using data of EWAS, which included a meta-analysis for brain samples of MDD, a convergence analysis for brain and blood samples, and top results from available genome-wide expression and association data. Functional enrichment and protein-protein interaction network analyses were also performed. Results: The meta-analysis for brain samples detected a significant gene, FAM53B. A list of forty-four top differentially methylated (DM) candidate genes was found, including GRM8, NOTCH4 and SEMA6A, in addition to known druggable genes. The binding-sites for brain-expressed transcription factors, CREB and FOXO1, were enriched in the top DM genes. The protein-protein interaction networks showed that DM genes for MDD, such as RPRM and TMEM14B, play a central role. Conclusion: In this study, we found integrative evidence for the possible role of novel candidate genes and pathways. These genes are involved in mechanisms of synaptic plasticity, which have been associated with several psychiatric disorders. Analysis of epigenetic factors have a great potential for the identification of the mechanisms involved in the pathogenesis of MDD, taking into account their possible role in the interaction between genetic factors and the environment.

Keywords: Epigenomics, DNA Methylation, Psychiatric Genomics, Bioinformatics, Major depressive disorder

Introduction

Major depressive disorder (MDD) is a common psychiatric entity, being characterized by alterations in mood and in other clinical dimensions, which lead to functional impairment in patients. 1 MDD has an average 12-month prevalence of around 6%1 and an estimated heritability of 35-45%.2 A secondary analysis of available global data has shown that the number of incident MDD cases increased from 172 to 258 million in the 1990-2017 period, being one of the psychiatric disorders with the largest impact on burden of disease.3

In recent years, several genome-wide analyses have been carried out to identify the molecular risk factors associated with MDD,² as well as multiple genome-wide association studies (GWAS)⁴ and genome-wide expression studies (GWES).⁵ In this context, epigenetic mechanisms have been of interest in the study of the pathogenesis of MDD, as a possible way of finding the interaction between genetic factors and environmental variables (such as psychological stress).6 Among several epigenetic factors, the analysis of DNA methylation levels has been studied for multiple psychiatric disorders, primarily because of the negative correlation that is found between DNA methylation in promoter regions (in CpG islands) and gene expression.⁷

Epigenome-wide association studies (EWAS) have appeared as important strategies for the analysis of DNA methylation



"© The authors. 2022. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0) license, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. No commercial re-use is allowed."

levels across the genome, based on available microarray platforms that include hundreds of thousands of probes.⁶ Several EWAS for MDD and related phenotypes have been published,^{8,9} but there is the need for a bioinformatic analysis of the convergence of results from several available EWAS with other genomic evidence.^{5,10} In this study, we carried out a computational analysis of available genome-wide DNA methylation studies for MDD and their convergence with multiple lines of genomic evidence. In addition, we performed a meta-analysis for detecting differentially methylated genes in brain samples from subjects with MDD, considering the advantage of this approach to increase statistical power and to obtain more precise results through the combination of individual studies.¹¹

Methods

Data processing and convergence analysis of EWAS in brain and blood samples

The NCBI GEO database, an online repository for microarray data, ¹² was used to obtain raw data from available epigenomewide association studies for MDD. Data from five EWAS were extracted from the following published articles: Guintivano, 2013; ⁹ Chen, 2014; ¹³ Murphy, 2017BA11 and Murphy,

2017BA25 (both from the same article)¹⁴ and Crawford, 2018¹⁵ (Table 1). The genome-wide DNA methylation data obtained were used to generate two groups for comparison (MDD patients and control subjects), which were then analyzed using the GEO2R tool¹² to identify the differentially methylated (DM) probes for each study. The annotation files from the NCBI GEO database were used for the mapping from microarray probes to human gene identifiers. Convergent differentially methylated genes in these studies were revealed using the Venn diagram tool (http://bioinformatics.psb.ugent.be/webtools/Venn).

Meta-analysis of EWAS in brain samples

Additionally, a meta-analysis was performed using the robust rank aggregation (RRA) method in the R program. ¹⁶ In this analysis, four studies that analyzed DNA methylation in brain tissue samples were included (Table 1). The R package "RobustRankAggreg" was employed following the previously described protocol. ¹⁷ For the current study, the list of significant DM genes identified by GEO2R was used, which were ranked according to their P values. The RRA method allows to integrate data from different studies and methodologies, and uses a prioritized list of genes. ¹⁶ An adjusted P value of <0.05 was considered significant in this analysis.

NCBI GEO Author, Year Tissue Sample size **Platform PMID** Illumina Human Methylation 450K 49 MDD and 49 Guintivano, 2013 GSE41826 Frontal cortex 23426267 Beadchip (GPL13534) controls 17 MDD and 17 Illumina Human Methylation 27K Chen, 2014 GSE38873 Cerebellum 25243493 Beadchip (GPL8490) controls Frontal cortex. 20 MDD and 20 Illumina Human Methylation 450K Murphy, 2017BA11 GSE88890 28045465 Brodmann area 11 controls Beadchip (GPL13534) Frontal cortex, 17 MDD and 18 Illumina Human Methylation 450K Murphy, 2017BA25 GSE88890 28045465 Brodmann area 25 controls Beadchip (GPL13534) 49 MDD and 48 Illumina Human Methylation 450K Crawford, 2018 GSE113725 Whole Blood 29790996 controls Beadchip (GPL13534)

Table 1. Details of EWAS included

Abbreviations: PMID: PubMed identifier; NCBI GEO: NCBI GEO database identifier.

12 | archivosdeneurociencias.org Volume 27, Number 2, year 2022

Arch Neurocien (Mex) Alam Farwah B. et al.

Convergence analysis for common genes in EWAS and other genome-wide studies

Records of significant genes from genome-wide expression studies were extracted from a published meta-analysis of GWES for MDD patients and controls (amygdala, anterior cingulate cortex, cerebellum and prefrontal cortex). 18 Lists of significant genes were also extracted from genome-wide association studies for depressive symptoms, 19 personality traits²⁰ and for the case-control design for MDD.²¹ The significant genes obtained from EWAS for MDD, GWAS for depressive symptoms, GWAS for case-control studies of MDD, GWAS for personality traits and meta-analysis of GWES for MDD were analyzed for their convergence, using an online tool (http://bioinformatics.psb.ugent.be/webtools/ Venn). The genes found on convergence were compared with the following available lists: genes known to harbor mutations for neuropsychiatric disorders²² and genes that are highly expressed in human astrocytes and oligodendrocytes.²³

Enrichment analysis for convergent genes in EWAS

A functional enrichment analysis was carried out using the DAVID online tool, version 6.8,²⁴ for the following categories: Transcription Factor Binding Sites (TFBS) and Tissue Expression (GNF U133A). The significant genes (that were convergent in five EWAS) were compared with the rest of the genome by using a Fisher exact p value, including a correction for multiple testing using a False Discovery Rate (FDR) method. In the case of TFBS, the significant genes were analyzed for their convergence with transcription factors expressed in the brain.²⁵

Protein-protein interaction network for convergent genes in EWAS

An examination of the experimentally validated protein-protein interactions (PPI) was conducted using the online database of the Human Interactome Project ²⁶ for the significant genes that were convergent in EWAS included in this work. The program, Cytoscape 3.8.0,²⁷ was used to visualize these interactions, in which a connected subnetwork system, using >2 edges, was employed, 28 along with a degree filter (In + Out) of 30-292.

Results

Genome-wide DNA methylation data were extracted from 5 EWAS for MDD, which had samples from different brain regions and whole blood (Table 1). Significant genes from the five EWAS were analyzed; results showed one hundred and seventy-one genes that were differentially methylated in common between the 5 EWAS. Combinations of 4 EWAS identified sixty-five to six hundred and thirty-eight common DM genes (Figure 1, Table S1A). Also, we carried out a meta-analysis for studies performed in brain samples using the robust rank aggregation method. Only one gene, FAM53B (family with sequence similarity 53 member B), was identified as significant (Score: 3.3085, P= 0.0160).

A merging of convergent genes from EWAS for MDD with genes available from 1) GWAS for depressive symptoms, 2) GWAS for case-control studies of MDD, 3) GWAS for personality traits and 4) meta-analysis of GWES for MDD, resulted in a list of 44 top candidate genes (Table 2), including NOTCH4 (Neurogenic locus notch homolog protein 4) and SEMA6A (Semaphorin-6A). A number of these 44 genes have been found to harbor mutations for neuropsychiatric disorders (Table S2), such as COL4A2 (Collagen alpha-2(IV) chain) and RELN (Reelin); and to be enriched in astrocytes and oligodendrocytes (Table S2), such as FAM107B (Protein Family With Sequence Similarity 107 Member B) and TNS3 (Tensin-3).

Guintivano, 2013 Chen, 2014 Murphy,2017BA11 Murphy, 2017BA25 Crawford, 2018 171 DM genes 638 DM genes 67 DM genes 140 DM genes 65 DM genes 232 DM genes

Figure 1. Overview of differentially methylated (DM) genes from five EWAS for MDD.

Table 2. Main top candidate genes. EWAS: EWAS for MDD; GWASD: GWAS for depressive symptoms; GWASM: GWAS for case-control studies of MDD; GWASN: GWASN: GWASN: GWESM: GWE

Gene	Protein Name	Evidence	Gene	Protein Name	Evidence
ASIC2	Acid-sensing ion channel 2	GWASD, GWESM	PIEZO2	Piezo-type mechanosensitive ion channel	GWASM, GWESM
C3orf70	UPF0524 protein C3orf70	GWASM, GWESM	PTDSS2	Phosphatidylserine synthase 2	EWAS, GWESM
CDO1	Cysteine dioxygenase type 1	GWASM, GWESM	RCAN2	Calcipressin-2	GWASN, GWESM
CPLX1	Complexin-1	GWASM, GWESM	RELN	Reelin	GWASM, GWESM
COL4A2	Collagen alpha-2(IV) chain	EWAS, GWASM	RPRM	Protein reprimo	GWASM, GWESM
DAD1	Dolichyl- diphosphooligosaccharide protein	GWASN, GWESM	RYR2	Ryanodine receptor 2	EWAS, GWASM, GWESM
FAM107B	Protein FAM107B	EWAS, GWESM	SEMA6A	Semaphorin-6A	EWAS, GWASM
FHIT	Bis(5'-adenosyl)- triphosphatase	GWASD, GWASM	SMARCA2	Probable global transcription activator SNF2L2	GWASM, GWESM
GRM8	Metabotropic glutamate receptor 8	GWASM, GWESM	SSB	SPRY domain- containing SOCS box protein 2	EWAS, GWESM
IGSF21	Immunoglobulin superfamily member 21	EWAS, GWESM	STK39	STE20/SPS1-related proline-alanine-rich protein kinase	EWAS, GWESM
IL17RD	Interleukin-17 receptor D	GWASM, GWESM	TM7SF2	Delta(14)-sterol reductase TM7SF2	EWAS, GWESM
LOC102546299	[Uncharacterized]	GWASD, GWASM, GWASN	TMEM14B	Transmembrane protein 14B	GWASM, GWESM
LPCAT1	Lysophosphatidylcholine acyltransferase 1	GWASM, GWESM	TMEM241	Transmembrane protein 241	GWASM, GWESM
MRAP2	Melanocortin-2 receptor accessory protein 2	GWASM, GWESM	TNS3	Tensin-3	EWAS, GWESM
NCKAP1	Nck-associated protein 1	GWASM, GWESM	TRPM3	Transient receptor potential cation channel subfamily M member 3	GWASD, GWASM
NELL1	Protein kinase C-binding protein NELL1	GWASM, GWESM	TUSC3	Tumor suppressor candidate 3	GWASM, GWESM
NELL2	Protein kinase C-binding protein NELL2	GWASM, GWESM	UBA3	NEDD8-activating enzyme E1 catalytic subunit	GWASM, GWESM
NOTCH4	Neurogenic locus notch homolog protein 4	EWAS, GWASM	UNC13C	Protein unc-13 homolog C	GWASD, GWASM, GWASN
OFCC1	Orofacial cleft 1 candidate gene 1 protein	GWASD, GWASM	WIF1	Wnt inhibitory factor 1	GWASM, GWESM
PCP4	Calmodulin regulator protein PCP4	GWASM, GWESM	ZCCHC14	Zinc finger CCHC domain-containing protein 14	EWAS, GWASM
PEX5L	PEX5-related protein	GWASD, GWASM	ZCCHC24	Zinc finger CCHC domain-containing protein 24	GWASM, GWESM
PFKP	ATP-dependent 6-phosphofructokinase, platelet	EWAS, GWESM	ZIC2	Zinc finger protein ZIC 2	EWAS, GWESM

14 | archivosdeneurociencias.org Volume 27, Number 2, year 2022

Arch Neurocien (Mex) Alam Farwah B. et al.

A functional enrichment analysis found an enrichment of binding-sites for brain-expressed transcription factors (Table 3), such as CREB (cAMP responsive element binding protein), FOXO1 (forkhead box O1), and ZIC1 (Zinc family member 1). In addition, an analysis of the 44 candidate genes showed an enrichment of tissue expression as

Table 3. Functional enrichment analysis of top DM candidate genes from EWAS for MDD. TFBS: Transcription Factor Binding Sites; GNF U133A QUARTILE: Expression in Multiple tissues.

Category	Term	P value	FDR
UCSC_TFBS	LHX3	9.39E-05	0.009014
UCSC_TFBS	FOXO3	2.92E-04	0.014018
UCSC_TFBS	RP58	6.78E-04	0.015651
UCSC_TFBS	ISRE	6.83E-04	0.015651
UCSC_TFBS	AP2REP	8.15E-04	0.015651
UCSC_TFBS	CDPCR3	0.001284	0.01727
UCSC_TFBS	FAC1	0.001571	0.01727
UCSC_TFBS	CART1	0.001609	0.01727
UCSC_TFBS	HNF1	0.001684	0.01727
UCSC_TFBS	P53	0.00194	0.01727
UCSC_TFBS	IRF2	0.002141	0.01727
UCSC_TFBS	SRY	0.002348	0.01727
UCSC_TFBS	TGIF	0.002659	0.01727
UCSC_TFBS	NFE2	0.002698	0.01727
UCSC_TFBS	CREB	0.00319	0.019138
UCSC_TFBS	AP1	0.003655	0.019969
UCSC_TFBS	IK3	0.004006	0.019969
UCSC_TFBS	ZIC1	0.004134	0.019969
UCSC_TFBS	SREBP1	0.004247	0.019969
UCSC_TFBS	HFH1	0.004501	0.019969
UCSC_TFBS	GATA	0.004576	0.019969
UCSC_TFBS	CDC5	0.004903	0.020465
UCSC_TFBS	TAL1BETAITF2	0.005666	0.022002
UCSC_TFBS	CDPCR1	0.00573	0.022002
UCSC_TFBS	FOXO4	0.006855	0.025311
UCSC_TFBS	STAT3	0.007589	0.026549
UCSC_TFBS	BRACH	0.007743	0.026549
UCSC_TFBS	AREB6	0.009089	0.028731
UCSC_TFBS	FOXO1	0.009868	0.028731
GNF_U133A_ QUARTILE	Olfactory Bulb	1.24E-04	0.002084
GNF_U133A_ QUARTILE	Dorsal root ganglia	6.72E-04	0.007502
GNF_U133A_ QUARTILE	Pituitary	0.005969	0.047689

well, such as Pituitary and Olfactory Bulb (Table 3). A PPI network visualization showed that candidate genes for MDD, such as TMEM14B (transmembrane protein 14B) and RPRM (reprimo, TP53 dependent G2 arrest mediator homolog), play a central role in this network (Figure 2).

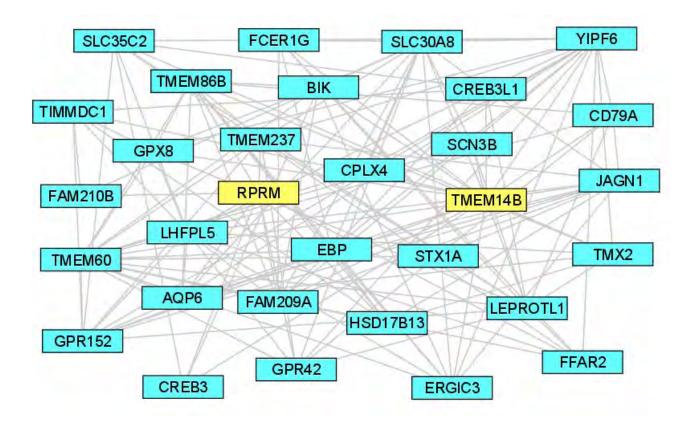
Discussion

Epigenetic factors have been of particular interest in the analysis of the mechanisms involved in the pathogenesis of MDD, considering the possible interaction between genetic factors and the environment.⁶ Multiple epigenome-wide association studies for major depression and related phenotypes have been carried out and published in recent years.⁶ A bioinformatic analysis of the convergence of results from several available EWAS with other genomic evidence 10,29,30 (D. A. Forero et al., 2017; Niculescu & Le-Niculescu, 2010) can be helpful for the identification of novel genes and pathways for MDD.

In this study, we found integrative evidence for the possible role of novel candidate genes and pathways. Key candidate genes such as NOTCH4 and SEMA6A were found in convergence with those identified in GWAS and GWES. These genes are involved in mechanisms of synaptic plasticity, which have been associated with several psychiatric disorders. 18,31,32 Among the candidate genes found in this investigation, genes which harbor mutations for neuropsychiatric disorders, such as COL4A2 and RELN, have been identified; as well as genes that are highly expressed in astrocytes and oligodendrocytes, such as TNS3 and FAM107B. Furthermore, binding-sites for brainexpressed transcription factors, such as FOXO1 and CREB, are of particular importance, given the previous evidence of involvement in pathophysiology of depression^{33,34} — with genes such as TMEM14B and RPRM observed to play a key role in the protein-protein interaction network.

Previously, Uddin et al found a difference in genome-wide DNA methylation patterns between unaffected and depressed individuals. Functional enrichment showed that methylated and unmethylated genes affect brain development, depending on specific pathways.³⁵ A study involving post-mortem frontal cortex samples found similar results for genes such as CPSF3, LASS2 and PRIMA1 having different methylation profiles.³⁶ Studies with candidate genes have complemented results from EWAS for MDD. A study with MDD patients showed higher levels of methylation at the BDNF gene. 37 Another casecontrol study also showed BDNF, FKBP5, CRHBP and NR3C1 gene promoters to be significantly hypermethylated in MDD.³⁸

Figure 2. Protein-protein interaction network for top candidate genes. Top DM candidate genes from EWAS for MDD were used. A highly connected subnetwork is shown and candidate genes are highlighted in yellow.



It is important for future MDD EWAS to be carried out in other regions of the world (such as Latin America or Africa), ¹⁸ that have millions of depression patients.³

Concerning the meta-analysis performed in this study, a DM gene, the FAM53B, was identified; which encodes a protein that is necessary to regulate the β-catenin-dependent Wnt signal transduction.³⁹ A GWAS has detected a variant in this gene as a risk for cocaine dependence in African-and European-American subjects.⁴⁰ Additionally, other polymorphisms in FAM53B are also associated with MDD and Alzheimer's diseases.⁴¹ Moreover, in a study that analyzed the effects of smoking on DNA methylation, a significant result for 525 genes including FAM53B was found.⁴² These findings suggest that this gene could play an important role in the molecular

mechanisms of different brain disorders. Interestingly, FAM53B was convergent with the study performed by Crawford, 2018, that analyzed DNA methylation in whole blood samples (Table S1). Despite the existence of additional EWAS performed in whole blood samples for depressive symptoms in middleaged and elderly persons,⁸ its raw data is unavailable and it was not possible to include their results in our study.

The number of EWASs included is one of the limitations of this study, as several primary EWAS do not have their data publicly available. Comprehensive meta-analyses of available EWAS could be performed if academic journals request for the public availability of such raw data. Development of user-friendly computational tools would also facilitate such meta-analyses of large volumes of epigenomic data. 44

16 | archivosdeneurociencias.org Volume 27, Number 2, year 2022

Arch Neurocien (Mex)

Alam Farwah B. et al.

References

- Otte C, Gold SM, Penninx BW, Pariante CM, Etkin A, Fava M, et al. Major depressive disorder. Nat Rev Dis Primers. 2016; 2:16065.
- 2. Gonda X, Petschner P, Eszlari N, Baksa D, Edes A, Antal P, et al. Genetic variants in major depressive disorder: From pathophysiology to therapy. Pharmacol Ther. 2019;194:22-43.
- 3. Liu Q, He H, Yang J, Feng X, Zhao F, Lyu J. Changes in the global burden of depression from 1990 to 2017: Findings from the Global Burden of Disease study. J Psychiatr Res. 2020;126:134-140.
- Major Depressive Disorder Working Group of the Psychiatric GC, Ripke S, Wray NR, Lewis CM, Hamilton SP, Weissman MM, et al. A mega-analysis of genome-wide association studies for major depressive disorder. Mol Psychiatry. 2013;18(4):497-511.
- Forero DA, Guio-Vega GP, Gonzalez-Giraldo Y. A comprehensive regional analysis of genome-wide expression profiles for major depressive disorder. J Affect Disord. 2017; 218:86-92.
- 6. Nagy C, Vaillancourt K, Turecki G. A role for activity-dependent epigenetics in the development and treatment of major depressive disorder. Genes Brain Behav. 2018; 17(3):e12446.
- 7. Januar V, Saffery R, Ryan J. Epigenetics and depressive disorders: a review of current progress and future directions. Int J Epidemiol. 2015; 44(4):1364-87.
- Story-Jovanova O, Nedeljkovic I, Spieler D, Walker RM, Liu C, Luciano M, et al. DNA Methylation Signatures of Depressive Symptoms in Middle-aged and Elderly Persons: Meta-analysis of Multiethnic Epigenome-wide Studies. JAMA psychiatry. 2018; 75(9):949-59.
- 9. Guintivano J, Aryee MJ, Kaminsky ZA. A cell epigenotype specific model for the correction of brain cellular heterogeneity bias and its application to age, brain region and major depression. Epigenetics. 2013; 8(3):290-302.
- Niculescu AB, Le-Niculescu H. Convergent Functional Genomics: what we have learned and can learn about genes, pathways, and mechanisms. Neuropsychopharmacology. 2010; 35(1):355-6.
- 11. Ramasamy A, Mondry A, Holmes CC, Altman DG. Key issues in conducting a meta-analysis of gene expression microarray datasets. PLoS medicine. 2008; 5(9):e184.
- Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, et al. NCBI GEO: archive for functional genomics data setsupdate. Nucleic Acids Res. 2013;41 (Database issue):D991-5.
- Chen C, Zhang C, Cheng L, Reilly JL, Bishop JR, Sweeney JA, et al. Correlation between DNA methylation and gene expression in the brains of patients with bipolar disorder and schizophrenia. Bipolar Disord. 2014; 16(8):790-9.

- 14. Murphy TM, Crawford B, Dempster EL, Hannon E, Burrage J, Turecki G, et al. Methylomic profiling of cortex samples from completed suicide cases implicates a role for PSORS1C3 in major depression and suicide. Transl Psychiatry. 2017; 7(1):e989.
- Crawford B, Craig Z, Mansell G, White I, Smith A, Spaull S, et al. DNA methylation and inflammation marker profiles associated with a history of depression. Hum Mol Genet. 2018; 27(16):2840-50.
- Kolde R, Laur S, Adler P, Vilo J. Robust rank aggregation for gene list integration and meta-analysis. Bioinformatics. 2012; 28(4):573-80.
- 17. Võsa U, Kolde R, Vilo J, Metspalu A, Annilo T. Comprehensive Meta-analysis of MicroRNA Expression Using a Robust Rank Aggregation Approach. In: Alvarez ML, Nourbakhsh M, editors. RNA Mapping: Methods and Protocols. New York: Springer New York; 2014. 361-73.
- Forero DA, Guio-Vega GP, González-Giraldo Y. A comprehensive regional analysis of genome-wide expression profiles for major depressive disorder. J Affect Disord. 2017; 218:86-92.
- Hek K, Demirkan A, Lahti J, Terracciano A, Teumer A, Cornelis MC, et al. A Genome-Wide Association Study of Depressive Symptoms. Biol Psychiatry. 2013; 73(7):667-78.
- Genetics of Personality C. Meta-analysis of Genome-wide Association Studies for Neuroticism, and the Polygenic Association With Major Depressive Disorder. JAMA psychiatry. 2015; 72(7):642-50.
- Ripke S, Wray NR, Lewis CM, Hamilton SP, Weissman MM, Breen G, et al. A mega-analysis of genome-wide association studies for major depressive disorder. Mol Psychiatry. 2013; 18(4):497-511.
- 22. Forero DA, Prada CF, Perry G. Functional and Genomic Features of Human Genes Mutated in Neuropsychiatric Disorders. The Open Neurology Journal. 2016; 10:143-8.
- 23. Kelley KW, Nakao-Inoue H, Molofsky AV, Oldham MC. Variation among intact tissue samples reveals the core transcriptional features of human CNS cell classes. Nat Neurosci. 2018; 21(9):1171-84.
- 24. Huang da W, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. Nat Protoc. 2009; 4(1):44-57.
- 25. Hawrylycz M, Miller JA, Menon V, Feng D, Dolbeare T, Guillozet-Bongaarts AL, et al. Canonical genetic signatures of the adult human brain. Nat Neurosci. 2015; 18(12):1832-44.
- Rolland T, Tasan M, Charloteaux B, Pevzner SJ, Zhong Q, Sahni N, et al. A proteome-scale map of the human interactome network. Cell. 2014; 159(5):1212-26.
- 27. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D,

- et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003; 13(11):2498-504.
- 28. Forero DA, González-Giraldo Y. Convergent functional genomics of cocaine misuse in humans and animal models. Am J Drug Alcohol Abuse. 2020; 46(1):22-30.
- 29. Forero DA, Gonzalez-Giraldo Y. Integrative In Silico Analysis of Genome-Wide DNA Methylation Profiles in Schizophrenia. J Mol Neurosci. 2020; 70(11):1887-1893.
- Ayalew M, Le-Niculescu H, Levey DF, Jain N, Changala B, Patel SD, et al. Convergent functional genomics of schizophrenia: from comprehensive understanding to genetic risk prediction. Mol Psychiatry. 2012; 17(9):887-905.
- 31. Hoseth EZ, Krull F, Dieset I, Morch RH, Hope S, Gardsjord ES, et al. Attenuated Notch signaling in schizophrenia and bipolar disorder. Sci Rep. 2018; 8(1):5349.
- 32. Pasterkamp RJ, Giger RJ. Semaphorin function in neural plasticity and disease. Curr Opin Neurobiol. 2009; 19(3):263-74.
- 33. Cattaneo A, Cattane N, Malpighi C, Czamara D, Suarez A, Mariani N, et al. FoxO1, A2M, and TGF-beta1: three novel genes predicting depression in gene X environment interactions are identified using cross-species and cross-tissues transcriptomic and miRNomic analyses. Mol Psychiatry. 2018; 23(11):2192-208.
- 34. Blendy JA. The role of CREB in depression and antidepressant treatment. Biol Psychiatry. 2006; 59(12):1144-50.
- 35. Uddin M, Koenen KC, Aiello AE, Wildman DE, de los Santos R, Galea S. Epigenetic and inflammatory marker profiles associated with depression in a community-based epidemiologic sample. Psychol Med. 2011;41(5):997-1007.
- Sabunciyan S, Aryee MJ, Irizarry RA, Rongione M, Webster MJ, Kaufman WE, et al. Genome-wide DNA methylation scan in major depressive disorder. PloS one. 2012; 7(4):e34451.
- Na K-S, Won E, Kang J, Chang HS, Yoon H-K, Tae WS, et al. Brain-derived neurotrophic factor promoter methylation and cortical thickness in recurrent major depressive disorder. Sci Rep. 2016; 6(1):21089.
- 38. Roy B, Shelton RC, Dwivedi Y. DNA methylation and expression of stress related genes in PBMC of MDD patients with and without serious suicidal ideation. J Psychiatr Res. 2017; 89:115-24.
- 39. Kizil C, Kuchler B, Yan JJ, Ozhan G, Moro E, Argenton F, et al. Simplet/Fam53b is required for Wnt signal transduction by regulating beta-catenin nuclear localization. Development. 2014; 141(18):3529-39.
- Gelernter J, Sherva R, Koesterer R, Almasy L, Zhao H, Kranzler HR, et al. Genome-wide association study of cocaine

- dependence and related traits: FAM53B identified as a risk gene. Mol Psychiatry. 2014; 19(6):717-23.
- 41. Ni H, Xu M, Zhan GL, Fan Y, Zhou H, Jiang HY, et al. The GWAS Risk Genes for Depression May Be Actively Involved in Alzheimer's Disease. Journal of Alzheimer's disease: JAD. 2018; 64(4):1149-61.
- 42. Dogan MV, Beach SRH, Philibert RA. Genetically contextual effects of smoking on genome wide DNA methylation. Am J Med Genet B Neuropsychiatr Genet. 2017; 174(6):595-607.
- 43. Brazma A, Hingamp P, Quackenbush J, Sherlock G, Spellman P, Stoeckert C, et al. Minimum information about a microarray experiment (MIAME)-toward standards for microarray data. Nat Genetics. 2001; 29(4):365-71.
- 44. Forero DA. Available Software for Meta-analyses of Genomewide Expression Studies. Curr Genomics. 2019; 20(5):325-31.

Article without conflict of interest

© Archivos de Neurociencias