ANNALS OF HEPATOLOGY

Volume 1

Number 3

July-September 2002

2002

Article:

From lipid secretion to cholesterol crystallization in bile. Relevance in cholesterol gallstone disease

> Copyright © 2002: Mexican Association of Hepatology

Otras secciones de este sitio:

- Índice de este número
- Más revistas
- Búsqueda

Others sections in this web site:

- Contents of this number
- Search





Concise Review

From lipid secretion to cholesterol crystallization in bile. Relevance in cholesterol gallstone disease

Piero Portincasa, 1,2 Antonio Moschetta, 2 Giuseppe Palasciano 2

Abstract

Failure of cholesterol homeostasis in the body can lead to cholesterol gallstone disease, the most common and costly gastrointestinal disease. The *primum movens* in cholesterol gallstone formation is the hypersecretion of hepatic cholesterol; this condition leads to bile chronically surpersaturated with cholesterol which is prone to rapid precipitation as cholesterol crystals in the gallbladder. Essential topics reviewed here deal with pathways of biliary lipid secretion, cholesterol solubilization and crystallization in bile, according to recent advances. Main *in vivo* events in cholesterol gallstone disease are also described.

Key words: Bile salts, crystals, micelles, phospholipids, vesicles.

Introduction

Failure of cholesterol homeostasis in the body can result in cholesterol gallstone disease, the most common and costly gastrointestinal disease. The estimated prevalence of cholelithiasis in adult population is 10-15%. Although there are three predominant types of stones: cholesterol, brown pigment and black pigment, in countries

Department of Internal Medicine and Public Medicine (DIMIMP) University Medical School. Bari, Italy

Address for correspondence:
Piero Portincasa, MD, PhD
Cattedra di Semeiotica Medica–Sezione di Medicina Interna
Dipartimento di Medicina Interna e Medicina Pubblica (DIMIMP)
University of Bari Medical School-Policlinico-70124 Bari-Italy
Tel. +39-80-5478.227-Fax +39-80.5478.232
p.portincasa@semeiotica.uniba.it

Abbreviations: AQPs, aquaporins; BSEP, bile salt export pump; DPPC, dipalmitoyl phosphatidylcholine; CSI, cholesterol saturation index; EYPC, egg-yolk phosphatidylcholine; MDR, multi-drugresistant; MMC, migrating motor complex; PFIC, progressive familial intrahepatic cholestasis; PL, phospholipids; SM, sphingomyelin.

using western-style diet, gallstones are made of 75-80% cholesterol⁴ in up to 80% of patients with gallstones.⁵

The classical *paradigma* on the pathogenesis of cholesterol gallstones relies on the coexistence of at least 3 defects: hypersecretion of hepatic cholesterol resulting in chronically surpersaturated bile, increased crystallization of biliary cholesterol and gallbladder stasis. ⁶⁻⁹ The study of physical-chemical factors and pathways leading to cholesterol crystallization in bile has clinical relevance. A better understanding of early events in gallstone formation, in fact, plays a key role for the identification of potential factors delaying or preventing precipitation of cholesterol crystals and gallstone formation in bile.

In this review, current opinions on the pathogenesis of cholesterol gallstones with respect to lipid secretion, cholesterol solubilization and crystallization in bile will be discussed.

Secretion of lipids in bile

The hepatocyte plasma membrane is functionally divided into a canalicular (or apical) region adjacent to the lumen of the bile canaliculus and a sinusoidal (or basolateral) region in close contact with sinusoidal blood. The process of bile secretion initiates at the hepatic level in the canaliculus, which is a specialized luminal space formed by two hepatocytes. Although the canalicular region comprises only 10-15% of the total plasma membrane, it plays a crucial role in the process of nascent bile formation and biliary secretion of bile salts, water, phospholipids and cholesterol. Bile is a complex fluid in which over 95% is water^{10,11} mixed with three classes of lipids (i.e. bile salts, cholesterol, phospholipids), proteins, bilirubin, and organic anions.⁵ Bile formation starts with active secretion of solutes, followed by osmotic attraction of water. The hepatocyte is the major site for cholesterol synthesis and its elimination in bile. Hepatic secretion of bile salts and cholesterol into bile is the basis for the elimination of excess cholesterol from the body. Molecular mechanisms of the vectorial transport of biliary constituents into the bile are now better understood^{12,13} (Figure 1). Canalicular secretion of bile is a primary active transport mediated by a series of ATP-binding cassette (ABC) transporters. 14 They comprise the sister of P-glycoprotein (Spgp)^{15,16} also known as the bile salt export pump (BSEP, the major

¹ Chair of Semeiotica Medica.

² Section of Internal Medicine.

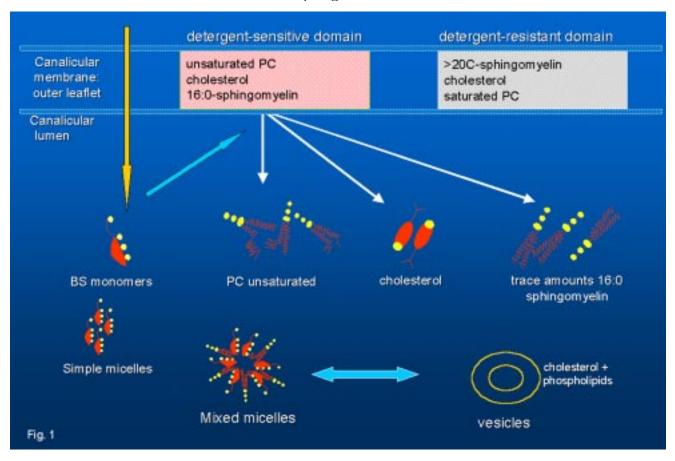


Figure 1. Current views on lipid secretion in bile.⁸¹ The outer leaflet of the hepatocyte canalicular membrane may contain detergent-sensitive domains (with bile-destined phosphatidylcholine species, that have unsaturated acyl chains at the sn-2 position, *plus* small amounts of 16:0 sphingomyelin and cholesterol), and detergent-resistant laterally separated domains (sphingomyelin with long ≥ 20 C atoms saturated acyl chains, disaturated phosphatidylcholine species and cholesterol (modified according to⁸¹)). PC, phosphatidylcholine.

Table I. Lipid transporters involved in bile formations and the phenotype of their mutations.

Name	Gene	Function	Mutation
MDR-3	ABCB4	PC flippase from inner to outer leaflet of the hepatocyte canalicular membrane	PFIC-3
BSEP	ABCB11	Bile salt export pump across the hepatocyte canalicular membrane	PFIC-2
ABCG5	ABCG5	Phytosterols, cholesterol?	Sitosterolemia
ABCG8	ABCG8	Cholesterol?	unknown

Adapted from Oude Elferink and Groen²⁸

Legend: BSEP, bile salt export pump; MDR, multi-drug-resistant; PC, phosphatidylcholine; PFIC, progressive familiar intrahepatic cholestasis

canalicular bile salt export pump of mammalian liver), the canalicular multidrug resistance protein MRP2 (cMOAT/cMRP) for transport of anionic conjugates such as glucuronide and glutathione conjugates such as glucuronide and glutathione conjugates and the canalicular multidrug resistance P-glycoprotein MDR1 for excretion of hydrophobic drugs, amphiphilic organic cations, toxins, metabolites, steroid hormones, hydrophobic peptides or glycolipids. ABC transporters also include the human MDR3 P-glycoprotein (corresponding to the murine mdr2 P-glycoprotein which functions as a "flippase" of phosphatidylcholine molecules from the inner to the outer leaflet of the canalicular membrane. Lipid transporters

in bile and relative genes are listed in *Table I*. The relationship between bile salt and lipid secretion is curvilinear; the secretion of both phospholipids and cholesterol plateaus at high bile salt secretion rates.²³ However, phospholipid secretion rate is always higher than that of cholesterol. Amount of the bile salts-induced biliary lipid secretion is positively related to the hydrophobicity of secreted bile salt species.²⁴ Secretion of "flipped" phosphatidylcholine from the outer leaflet into the canalicular lumen means mainly formation of vesicles from the external hemileaflet of the canalicular membrane.²⁵ Following their secretion into the canalicular lumen, detergent bile salts directly micellize consider-

able amounts of lipid from the membrane. 26,27 Experiments with mdr2 deficient mice (*i.e.* phosphatydilcholine not secreted) show that cholesterol secretion can occur without phospholipid secretion; in this model the infusion of the hydrophobic bile salt deoxycholate is sufficient for the extraction of cholesterol from the canalicular membrane. 26

These findings points to an alternative path of cholesterol secretion. Researches are focusing on the so-called "cholesterol pumping system".28 One theory has considered the possibility that the ABC transporter ABCA1 could play a direct role in cholesterol secretion in bile. ABCA1 regulates serum HDL cholesterol levels and is considered to control the first step of reverse cholesterol transport from the periphery to the liver.²⁹⁻³¹ Recently, Groen et al. 32 have shown that Abca1-/- knockout mice have preserved biliary secretion rates of cholesterol, bile salts, and phospholipid as compared with wild-type animals, in spite of absence of HDL. It appears that plasma HDL levels and ABCA1 activity do not control net cholesterol transport from the periphery via the liver into bile, indicating that ABCA1 has no direct role in cholesterol secretion in bile.

Recently, a possible role for other two ABC trasporters, namely ABCG5 and ABCG8 (both expressed in the intes-

tine and in the liver) in pumping cholesterol molecules in bile, has been advocated.^{28,33} A direct evidence suggests that the two transporters might be involved in the elimination of plant sterols. Accumulation of plant sterols, in fact, has been observed in patients with sitosterolemia, a recessively inherited rare disease in which the disease locus is localized to chromosome 2p21 and *ABCG5* and *ABCG8* genes are involved.^{33,34}

Cholesterol carriers are necessary in bile, as cholesterol is poorly soluble in an aqueous environment (Figure 2). Phenomena of lipid aggregation in bile start immediately after secretion; cholesterol is solubilized in bile in mixed micelles by bile salts and phospholipids. Phosphatidylcholine (PC) is the major phospholipid in bile (> 95% of total: mainly 16:0 acyl chains at the sn-1 position and mainly unsaturated (18:2 > 18:1 > 20:4) acyl chains at the sn-2 position³⁵). In the canalicular space also cholesterol/ PC vesicles exist (ratio ~0.3). During concentration of bile in the bile ducts, however, the cholesterol/PC vesicular ratio increases (> 1), due to progressive micellization of PC by bile salts. In the gallbladder, further concentration of bile results in vesicles aggregates and fusion into multilamellar supersaturated cholesterol/PC vesicles (ratio > 1); such vesicles are unstable due to cholesterol-rich

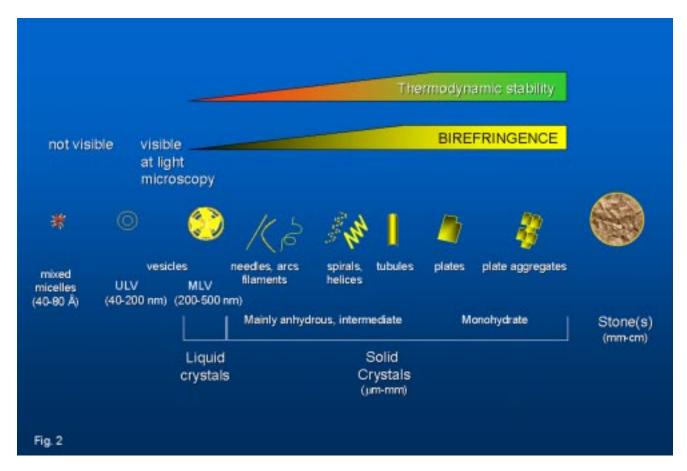


Figure 2. Schematic overview of cholesterol carriers in bile, according to microscopic appearance and crystal shapes. ULV, unilamellar vesicles; MLV, multilamellar vesicles (with typical "maltese cross" birefringent aspect at polarizing light microscopy).

microdomains, prone to precipitate as cholesterol crystals.^{36,37} Events are greatly accelerated in the case of cholesterol supersaturation (see below). The possibility that cholesterol crystals precipitate also from "supersaturated" micelles, has been advocated.³⁸

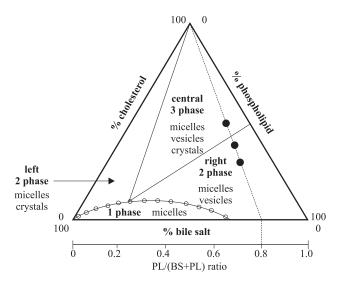


Figure 3. The ternary equilibrium phase diagram³⁹ shows relative concentrations of the three major biliary lipids (cholesterol, phospatidylcholine -PL, bile salts -BS,) and defines the limits of cholesterol solubility. Phase boundaries depend on relative composition of lipids, total lipid concentration (7.3 g/dL), and temperature (37°C). Each triangular apex depicts biles made of a single lipid. Points plotting within the diagram depict biles with different composition. Depicted are a onephase (micellar) zone at the bottom, a left two-phase zone (containing micelles and crystals), a central three-phase zone (containing micelles, vesicles and crystals) and a right two-phase zone (containing micelles and vesicles). The cholesterol saturation index (CSI) has its maximum normal value set at 1 (open dots at the micellar-phase boundary) and is defined as the ratio between molar percentage of cholesterol and the maximum micellar solubility of cholesterol. It can be seen from the diagram that for CSI > 1, biles must contain at least one additional phase of cholesterol at equilibrium. On the basal axis is also depicted an axis with PL/(BS+PL) ratios. The three theoretical model biles (•) plotting on interrupted line have identical PL/(BS+PL) ratios (in this case PL/ (BS+PL) ratio = 0.8).

From cholesterol solubilization to cholesterol crystallization

The conditions when cholesterol crystallization occurs are depicted within the ternary phase diagram in which the behaviour of cholesterol-bile salt-phospholipid aggregates in aqueous solutions are shown (Figure 3). Appearance and transition of micelles, vesicles, and cholesterol crystals are studied between metastable supersaturated systems and thermodynamic equilibrium (at fixed relative total lipid concentration, temperature, and pH). Wang & Carey³⁹ demonstrated that the ternary phase diagram contains a one-phase zone (only micelles), a left two-phase (micelles and cholesterol crystals-containing) zone, a central three-phase (micelles, vesicles and cholesterol crystals-containing) zone and a right two-phase (micelles and vesicles-containing) zone. Accurate quantification of such phases requires isolation of micelles and vesicles by gel filtration 40-43 and repeated observation of crystal growth.³⁹ Distinct sequences of cholesterol crystallization occur in the left two-phase and the central three-phase zones, involving anhydrous cholesterol crystals (needles, arcs, tubules, spirals, mass density ~1.03 g/dL) firtsly described in model biles by Konikoff et al. in 199244 and mature rhomboid birefringent plate-like cholesterol monohydrate crystals (mass density ~1.45 g/dL). Table II depicts the characteristics of different cholesterol crystal forms observed in bile. We also studied this topic⁴⁵ and recently we compared four independent methods to assess cholesterol crystallization in model biles i.e.light microscopy, nephelometry, spectrophotometry and biochemical assay; we found that morphology and size of cholesterol crystals strongly affect turbidimetric estimates of crystal mass. In principle, chemical measurement of crystal mass should be the method of choice for exact quantitation of cholesterol crystallization within the phase diagram.⁴⁶

The study of changes within the ternary phase diagram has pathophysiogical relevance for the prevention of cholesterol crystallization and, ultimately, of cholesterol gall-

Table II. Crystal forms of cholesterol in bile

Form	Crystal configuration	Crystal habit ¹	Unit cell	Mass density g/dL	Birefringence (polarizing light)	Thermodynamic stability at 37°C²
Monohydrate	Triclinic	Romboid, plate-like, fixed angles (79.2 and 100.8 degrees), ften notched corner. Better observed at continuous high humidity	8 molecules cholesterol 8 molecules water	1.045	+++	stable
Anhydrous	Triclinic	Needle-like	8 molecules cholesterol (low (< 31.6°C) temperatur 16 molecules cholesterol (h (> 31.6°C) temperature)	*	-+	unstable
Intermediate (mainly anhyd	lrous)	Filaments, arcs, needles, helices, tubules		~1.03	±	metastable

Note: depending on hydration status and speed of crystallization: 2 indicates that crystal structure is less favourable thermodynamically 82

Table III. Major in vivo events in cholesterol gallstone disease.

Events	Mechanisms	Ref.
Increased cholesterol synthesis	Increased HMGCoA activity	57
Cholesterol crystallization	Precipitation of cholesterol in supersaturated bile	6,44,58
Increased bile salt hydrophobicity	Increased deoxycholate production	59
Protein secretion from gallbladder wall	Increased pronucleating mucin concentration	60
Gallbladder stasis	Absence of gallbladder volume fluctuations in fasting state;	
	Dissociation from MMC;	70
	Impaired gallbladder postprandial emptying;	
	Reduced postprandial turnover of bile	63-65
Defective contractility of gallbladder strips	Smooth muscle disfunction due to miotoxic cholesterol accumulation	62,67
Slow intestinal transit	Impairment of MMC	71

stone formation. When bile is diluted (i.e. lower total lipid concentration) or enriched with disaturated phospholipids or hydrophilic bile salts, the micellar zone is smaller and the right two-phase (i.e. vesicles and micelles-containing) zone expands to the left at the expense of the crystals-containing (i.e. central three-phase and left twophase) zones. The opposite is true with concentrated bile or biles enriched with unsaturated phospholipids or hydrophobic bile salts. We found that in excess of bile salts (i.e. low phospholipid/(bile salts/phospholipid) ratios -referred to as PL/(BS+PL) ratios $\sim \le 0.2$), crystals precipitate at fast rates forming intermediate anhydrous cholesterol crystals and mature rhomboid plate-like cholesterol monohydrate crystals.46 At higher amounts of phospholipids, large amounts of cholesterol are solubilized in vesicles together with phospholipids and crystal precipitation proceeds at slower rates as mainly mature cholesterol monohydrate. In case of excess phospholipids (high PL/ (BS+PL) ratios), solid cholesterol crystals do not occur, and cholesterol is mainly solubilized in vesicular phases. We also showed that phospholipids can interfere within model systems plotting in the left two-phase or central three-phase (crystal-containing) zones. Indeed, speed and extent of crystallization increases with dipalmitoyl phosphatidylcholine (DPPC) or sphingomyelin (SM), as compared with egg-yolk phosphatidylcholine (EYPC). 45 Disaturated phosphatydilcholine (PC) species inhibit crystallization, and PC species with unsaturated acyl chains at the sn-2 position progressively promote crystallization at increasing unsaturation. 47-49 Thus, compared to EYPC-containing systems,³⁹ in SM- or DPPC-containing systems⁵⁰ the right two-phase (vesicles and micelles-containing) zone is greatly expanded to the left at the expense of the crystals-containing (central three-phase and left twophase) zones.

It has been suggested that dietary modification towards more saturated biliary phospholipids might prevent gallstone formation in humans, but effects of dietary modification are expected to be relatively small. In a clinical study, biliary cholesterol crystallization or lipid solubilization did not change with dietary modification.⁵¹

In our studies, the hydrophilic bile salt tauroursodeoxycholate prevented cholesterol crystallization and protected against more hydrophobic bile salt-induced crystallization; 45,52 this is consistent with previous data. 53 Apparently, solubilization of cholesterol in vesicular phases (obtained by enlargement of the central three-phase zone) is a prerequisite for reduced crystallization by tauroursodeoxycholate. Ursodeoxycholate or its tauro-conjugate are used as oral litholitic agents in cholesterol gallstones. When ursodeoxycholate was fed to gallstone patients (~12 mg/Kg b.w./d during 1 mo.), we found that ex vivo incubated fresh bile contained cholesterol crystals which decreased in size or even disappeared at prolonged observation, while ultrafiltered (isotropic) incubated bile remained totally crystal-free.⁵⁴ Newly synthesized fatty acid bile acid conjugates (FABAC) have been shown to dissolve pre-existing cholesterol crystals⁵⁵ and to dissolve stones when given orally to gallstone-susceptible C57J/L mice⁵⁶ but more studies are necessary to translate such observations into clinical practice.

Cholesterol gallstone disease: an overview of events in vivo

Cholesterol gallstone disease is a multifactorial condition, and in vivo the events are even more complex. Liver (secretion), gallbladder (concentration) and intestine (absorption) are the three organs handling biliary lipids which play a key role in the pathogenesis of the cholesterol gallstone disease. Table III depicts the most important factors which deserve attention. In the liver increased cholesterol synthesis due to increased HMGCoA activity has been reported.⁵⁷ In bile, cholesterol precipitation in crystal forms^{6,44,58} as well as increased bile salt hydrophobicity⁵⁹ are present, together with increased secretion of pronucleating proteins,54 and especially mucin.60 The combination of factors such as cholesterol supersaturation, bile salt hydrophobicity, and pronucleating proteins has been recently investigated by van Erpecum et al.61 in mice susceptible to cholesterol gallstone formation (i.e. C57L inbred mice). It has been demonstrated that both cholesterol supersaturation and

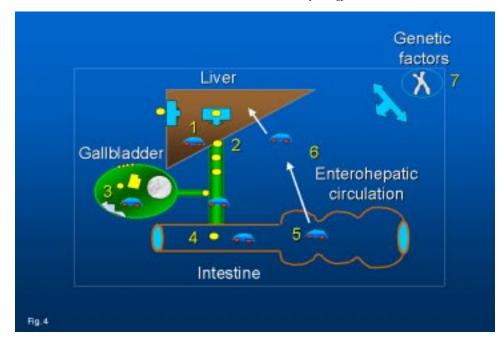


Figure 4. Current concepts on the pathogenesis of cholesterol gallstones: 1, cholesterol hypersecretion by the liver; 2, sustained hypercholesterobilia; 3, events in the gallbladder (colesterol incorporation into the sarcolemma leading to defective smooth muscle contractility, increased mucin secretion, increased deoxycholate, cholesterol crystallization, stone growth); 4, intestinal hypomotility (and delayed transit of bile); 5, increased bacterial production of deoxycholate; 6, increased liver uptake and biliary (re)secretion of deoxycholate; 7, the genetic background can influence several of the above mentioned mechanisms.

crystallization *plus* increased bile salt hydrophobicity and mucin concentration are early pathogenic events; *in vitro* crystallization-promoting immunoglobulins and aminopeptidase-N do not appear to be major factors in murine gallstone pathogenesis. Motility defects of gall-bladder, both *in vivo*⁶²⁻⁶⁶ and *ex vivo*,^{62,67,68} leading to gallbladder stasis are also important risk factors for gallstone disease. Finally, intestine may play an important role,⁶⁹ since intestinal motility disorders are common in gallstone patients,⁷⁰ with an incoordination of the migrating motor complex (MMC), the housekeeper of the intestine.⁷¹ In *figure 4* are reported some of the current concepts on the pathogenesis of cholesterol gallstones.

In conclusion, current concepts on pathogenic factors in cholesterol gallstones are schematically depicted in figure 4. Excess biliary cholesterol represents the primum movens and this condition anticipates increased incorporation of cholesterol into the sarcolemma (hypomotile gallbladder and delayed intestinal transit), sluggish enterohepatic recirculation of bile salts, increased hepatic reuptake and biliary secretion of the hydrophobic bile salt deoxycholate. As a result, more cholesterol secreted in bile becomes available for precipitation as crystals. Events leading to cholesterol crystallization need to be studied both in vitro by artificial model biles plotting in the ternary equilibrium phase diagram and in human bile ex vivo. Cholesterol gallstone formation may be partly genetically determined, as suggested by both epidemiologic studies,⁷² family surveys,⁷³ and clinical studies. 74-78,79 The role of genes in the pathogenesis of gallstones has been recently reviewed. 80 In the murine model, candidate genes for cholesterol gallstones have been identified: defects are found for several intracellular biliary proteins (hepatocyte cytosol); membrane lipid transport proteins (hepatocytes and cholangiocytes), transcription factors (membrane-bound and ligand-activated), lipoprotein receptors and intracellular lipid transporters. Changes in genes for the conformation/activity of the gallbladder smooth muscle CCK receptor and changes in the expression of gallbladder epithelial mucin might represent further pathogenic factors for cholesterol gallstone formation. More research, however, is needed to unravel remaining mechanisms of cholesterol gallstone disease.

References

- Sleisenger MH, Fordtran JS. Gastrointestinal Disease: pathophysiology, diagnosis, management. Philadelphia: W.B. Saunders, 1993.
- Ingelfinger FJ. Digestive disease as a national problem. V. Gallstones. Gastroenterology 1968: 55: 102-4.
- 3. Attili AF, Carulli N, Roda E, et al. Epidemiology of gallstone disease in Italy: prevalence data of the multicenter italian study on cholelithiasis (M.I.C.O.L.). *Am J Epidemiology* 1995: 141: 158-65.
- Bills PM, Lewis D. A structural study of gallstones. Gut 1975: 16: 630-7.
- Sherlock S, Dooley J. Diseases of the liver and biliary system. Oxford: Blackwell Science, 2002.
- Holan KR, Holzbach RT, Hermann RE, et al. Nucleation time: a key factor in the pathogenesis of cholesterol gallstone disease. *Gastroenterology* 1979: 77: 611-7.
- Carey MC. Formation of cholesterol gallstones: the new paradigm. In: Paumgartner G, Stiehl A, Gerok W, eds. *Trends in bile acid research*. Dordrecht, The Netherlands: Kluwer Academic Publishers, 1989: 259-81.
- Wang DQ, Paigen B, Carey MC. Phenotypic characterization of Lith genes that determine susceptibility to cholesterol cholelithiasis in inbred mice: physical-chemistry of gallbladder bile. *Journal of Lipid Research* 1997: 38: 1395-411.
- LaMont JT, Carey MC. Cholesterol gallstone formation. 2. Pathobiology and pathomechanics. *Prog Liver Dis* 1992: 10: 165-91.
- Marinelli RA, LaRusso NF. Solute and water transport pathways in cholangiocytes. Semin Liver Dis 1996: 16: 221-9.
- Marinelli RA, LaRusso NF. Aquaporin water channels in liver: their significance in bile formation. *Hepatology* 1997: 26: 1081-4.

- Haussinger D, Schmitt M, Weiergraber O, et al. Short-term regulation of canalicular transport. Semin Liver Dis 2000: 20: 307-21.
- Nathanson MH, Boyer JL. Mechanisms and regulation of bile secretion. *Hepatology* 1991: 14: 551-66.
- Meier PJ, Eckhardt U, Schroeder A, et al. Substrate specificity of sinusoidal bile acid and organic anion uptake systems in rat and human liver. *Hepatology* 1997: 26: 1667-77.
- Childs S, Yeh RL, Georges E, et al. Identification of a sister gene to P-glycoprotein. Cancer Res 1995: 55: 2029-34.
- Gerloff T, Stieger B, Hagenbuch B, et al. The sister of P-glycoprotein represents the canalicular bile salt export pump of mammalian liver. *J Biol Chem* 1998: 273: 10046-50.
- 17. Taniguchi K, Wada M, Kohno K, et al. A human canalicular multispecific organic anion transporter (cMOAT) gene is overexpressed in cisplatin-resistant human cancer cell lines with decreased drug accumulation. *Cancer Res* 1996: 56: 4124-9.
- Ishikawa T, Ali-Osman F. Glutathione-associated cis-diamminedichloroplatinum (II) metabolism and ATP-dependent efflux from leukemia cells. Molecular characterization of glutathione-platinum complex and its biological significance. *J Biol Chem* 1993: 268: 20116-25.
- Muller M, Jansen PL. Molecular aspects of hepatobiliary transport. *Am J Physiol* 1997: 272: G1285-G1303.
- Gros P, Ben Neriah YB, Croop JM, et al. Isolation and expression of a complementary DNA that confers multidrug resistance. *Nature* 1986: 323: 728-31.
- Roninson IB, Chin JE, Choi KG, et al. Isolation of human mdr DNA sequences amplified in multidrug-resistant KB carcinoma cells. *Proc Natl Acad Sci USA* 1986: 83: 4538-42.
- Smit JJM, Schinkel AH, Oude Elferink RPJ, et al. Homozygous disruption of the murine mdr2 P-glycoprotein gene leads to a complete absence of phospholipid from bile and to liver disease. *Cell* 1993: 75: 451-62.
- 23. Barnwell SG, Tuchweber B, Yousef IM. Biliary lipid secretion in the rat during infusion of increasing doses of unconjugated bile acids. *Biochim Biophys Acta* 1987: 922: 221-33.
- Verkade HJ, Vonk RJ, Kuipers F. New insights into the mechanism of bile acid-induced biliary lipid secretion. *Hepatology* 1995: 21: 1174-89.
- Crawford JM, Möckel GM, Crawford AR, et al. Imaging biliary lipid secretion in the rat: ultrastructural evidence for vesiculation of the hepatocyte canalicular membrane. *Journal of Lipid Research* 1995: 36: 2147-63.
- Oude Elferink RPJ, Ottenhoff R, van Wijland M, et al. Uncoupling of biliary phospholipid and cholesterol secretion in mice with reduced expression of mdr2 P-glycoprotein. *Journal of Lipid Research* 1996: 37: 1065-75.
- Oude Elferink RPJ, Tytgat GNJ, Groen AK. The role of mdr2 P-glycoprotein in hepatobiliary lipid transport. FASEB Journal 1997: 11: 19-28.
- Elferink RO, Groen AK. Genetic defects in hepatobiliary transport. Biochim Biophys Acta 2002: 1586: 129-45.
- Bodzioch M, Orso E, Klucken J, et al. The gene encoding ATP-binding cassette transporter 1 is mutated in Tangier disease. *Nat Genet* 1999: 22: 347-51.
- Brooks-Wilson A, Marcil M, Clee SM, et al. Mutations in ABC1 in Tangier disease and familial high-density lipoprotein deficiency. *Nat Genet* 1999: 22: 336-45.
- Marcil M, Brooks-Wilson A, Clee SM, et al. Mutations in the ABC1 gene in familial HDL deficiency with defective cholesterol efflux. *Lancet* 1999: 354: 1341-6.
- Groen AK, Bloks VW, Bandsma RH, et al. Hepatobiliary cholesterol transport is not impaired in Abca1-null mice lacking HDL. *J Clin Invest* 2001: 108: 843-50.
- Lee MH, Lu K, Hazard S, et al. Identification of a gene, ABCG5, important in the regulation of dietary cholesterol absorption. *Nat Genet* 2001: 27: 79-83.
- Berge KE, Tian H, Graf GA, et al. Accumulation of dietary cholesterol in sitosterolemia caused by mutations in adjacent ABC transporters. Science 2000: 290: 1771-5.
- Hay DW, Cahalane MJ, Timofeyeva N, et al. Molecular species of lecithins in human gallbladder bile. *Journal of Lipid Research* 1993: 34: 759-68.

- Mazer NA, Carey MC. Quasi-elastic light-scattering studies of aqueous biliary lipid systems. Cholesterol solubilization and precipitation in model bile solutions. *Biochemistry* 1983: 22: 426-42.
- Sömjen GJ, Gilat T. A non-micellar mode of cholesterol transport in human bile. FEBS Lett 1983: 156: 265-8.
- Ahrendt SA, Fox-Talbot MK, Kaufman HS, et al. Cholesterol nucleates rapidly from mixed micelles in the prairie dog. *Biochim Biophys Acta* 1994: 1211: 7-13.
- 39. Wang DQ, Carey MC. Complete mapping of crystallization pathways during cholesterol precipitation from model bile: influence of physicalchemical variables of pathophysiologic relevance and identification of a stable liquid crystalline state in cold, dilute and hydrophilic bile saltcontaining system. *Journal of Lipid Research* 1996: 37: 606-30.
- Donovan JM, Jackson AA. Rapid determination by centrifugal ultrafiltration of inter-mixed micellar/vesicular (non-lecithin-associated) bile salt concentrations in model bile: influence of Donnan equilibrium effects. *Journal of Lipid Research* 1993: 34: 1121-9.
- Donovan JM, Jackson AA, Carey MC. Molecular species composition of inter-mixed micellar/vesicular bile salt concentrations in model bile: dependence upon hydrophilic-hydrophobic balance. *Journal of Lipid Research* 1993: 34: 1131-40.
- Donovan JM, Timofeyeva N, Carey MC. Influence of total lipid concentration, bile salt:lecithin ratio, and cholesterol content on intermixed micellar/vesicular (non-lecithin-associated) bile salt concentrations in model bile. *Journal of Lipid Research* 1991: 32: 1501-12.
- Moschetta A, Eckhardt ER, De Smet MB, et al. Accurate separation of vesicles, micelles and cholesterol crystals in supersaturated model biles by ultracentrifugation, ultrafiltration and dialysis. *Biochim Biophys Acta* 2001: 1532: 15-27.
- 44. Konikoff FM, Chung DS, Donovan JM, et al. Filamentous, helical and tubular microstructures during cholesterol crystallization from bile. Evidence that biliary cholesterol does not nucleate classic monohydrate plates. *J Clin Invest* 1992: 90: 1156-61.
- Moschetta A, vanBerge-Henegouwen GP, Portincasa P, et al. Cholesterol crystallization in model biles. Effects of bile salt and phospholipid species composition. *Journal of Lipid Research* 2001: 42: 1273-81.
- Portincasa P, Venneman NG, Moschetta A, et al. Quantitation of cholesterol crystallization from supersaturated model bile. *J Lipid Res* 2002: 3: 604-10.
- Nishioka T, Tazuma S, Yamashita G, et al. Quantitative assessment of comparative potencies of cholesterol-crystal-promoting factors: relation to mechanistic characterization. *Biochemical Journal* 1998: 332: 343-50.
- Ringel Y, Sömjen GJ, Konikoff FM, et al. Increased saturation of the fatty acids in the sn-2 position of phospholipids reduces cholesterol crystallization in model biles. *Biochim Biophys Acta* 1998: 1390: 293-300.
- Halpern Z, Moshkowitz M, Laufer H, et al. Effect of phospholipids and their molecular species on cholesterol solubility and nucleation in human and model biles. Gut 1993: 34: 110-5.
- van Erpecum KJ, Carey MC. Influence of bile salts on molecular interactions between sphingomyelin and cholesterol: relevance to bile formation and stability. *Biochim Biophys Acta* 1997: 1345: 269-82.
- Pakula R, Konikoff FM, Rubin M, et al. The effects of dietary phospholipids enriched with phosphatidylethanolamine on bile and red cell membrane lipids in humans. *Lipids* 1996: 31: 295-303.
- van Erpecum KJ, Portincasa P, Gadellaa M, et al. Effects of bile salt hydrophobicity on nucleation behaviour of cholesterol crystals in model bile. Eur J Clin Invest 1996: 26: 602-8.
- Nishioka T, Tazuma S, Yamashita G, et al. Partial replacement of bile salts causes marked changes of cholesterol crystallization in supersaturated model bile systems. *Biochemical Journal* 1999: 340: 445-51.
- van Erpecum KJ, Portincasa P, Eckhardt E, et al. Ursodeoxycholic acid reduces protein levels and their nucleation-promoting activity in gallbladder bile. *Gastroenterology* 1996: 110: 1225-37.
- Gilat T, Leikin-Frenkel A, Goldiner L, et al. Arachidyl amido cholanoic acid (Aramchol) is a cholesterol solubilizer and prevents the formation of cholesterol gallstones in inbred mice. *Lipids* 2001: 36: 1135-40.
- Gilat T, Leikin-Frenkel A, Goldiner I, et al. Dissolution of cholesterol gallstones in mice by the oral administration of a fatty acid bile acid conjugate. *Hepatology* 2002: 35: 597-600.

- Maton PN, Ellis HJ, Higgins MJP, et al. Hepatic HMG-CoA reductase in human cholelithiasis: effects of chenodeoxycholic and ursodeoxycholic acids. *Eur J Clin Invest* 1980: 10: 325-32.
- Portincasa P, van Erpecum KJ, Jansen A, et al. Behavior of various cholesterol crystals in bile from gallstone patients. *Hepatology* 1996: 23: 738-48.
- 59. Shoda J, He BF, Tanaka N, et al. Increased deoxycholate in supersaturated bile of patients with cholesterol gallstones disease and its correlation with *de novo* syntheses of cholesterol and bile acids in liver, gallbladder emptying, and small intestinal transit. *Hepatology* 1995: 21: 1291-302.
- Shiffman ML, Shamburek RD, Schwartz CC, et al. Gallbladder mucin, arachidonic acid, and bile lipids in patients who develop gallstones during weight reduction. *Gastroenterology* 1993: 105: 1200-8.
- van Erpecum KJ, Wang DQH, Lammert F, et al. Phenotypic characterization of *Lith* genes that determine susceptibility to cholesterol cholelithiasis in inbred mice: soluble pronucleating proteins in gall-bladder and hepatic biles. *Journal of Hepatology* 2001: 35: 444-51.
- 62. Portincasa P, Di Ciaula A, Baldassarre G, *et al.* Gallbladder motor function in gallstone patients: sonographic and *in vitro* studies on the role of gallstones, smooth muscle function and gallbladder wall inflammation. *Journal of Hepatology* 1994: 21: 430-40.
- van der Werf SDJ, vanBerge-Henegouwen GP, Palsma DM, et al. Motor function of the gallbladder and cholesterol saturation of duodenal bile. *Neth J Med* 1987: 30: 160-71.
- Pomeranz IS, Shaffer EA. Abnormal gallbladder emptying in a subgroup of patients with gallstones. Gastroenterology 1985: 88: 787-91.
- Jazrawi RP, Pazzi P, Petroni ML, et al. Postprandial gallbladder motor function: refilling and turnover of bile in health and cholelithiasis. Gastroenterology 1995: 109: 582-91.
- 66. Moschetta A, Stolk MF, Rehfeld JF, et al. Severe impairment of post-prandial cholecystokinin release and gall- bladder emptying and high risk of gallstone formation in acromegalic patients during Sandostatin LAR. Aliment Pharmacol Ther 2001: 15: 181-5.
- 67. Shulz M, Hanisch E, Guldutuna S. In-vitro-kontraktilitatsverhalten humaner muskulatur von gallenblasen mit und ohne steinerkrankung -Relevanz des prostaglandin-systems für die CCK-regulierte motorik. Z Gastroenterol 1993: 31: 376-87.
- Portincasa P, Minerva F, Moschetta A, et al. Review article: in vitro studies of gall-bladder smooth muscle function. Relevance in cholesterol gallstone disease. Aliment Pharmacol Ther 2000: 14 Suppl 2: 19-26.
- van Erpecum KJ, van Berge-Henegouwen GP. Gallstones: an intestinal disease? Gut 1999: 44: 435-8.

- Stolk MF, van Erpecum KJ, Peeters TL, et al. Interdigestive gallbladder emptying, antroduodenal motility, and motilin release patterns are altered in cholesterol gallstone patients. *Dig Dis Sci* 2001: 46: 1328-34.
- Marcus SN, Heaton KW. Intestinal transit, deoxycholic acid and the cholesterol saturation of bile: three inter-related factors. *Gut* 1986: 27: 550-8.
- Attili AF, Capocaccia R, Carulli N, et al. Factors associated with gallstone disease in the MICOL experience. Hepatology 1997: 26: 809-18.
- Van der Linden W, Westlin N. The familial occurrence of gallstone disease. II. Occurrence in husbands and wifes. *Acta Genet Basel* 1966: 16: 377-82.
- Van der Linden W, Lindelof G. The familial occurrence of gallstone disease. Acta Genet Basel 1965: 15: 159-64.
- Danzinger RG, Gordon H, Schoenfield LJ, et al. Lithogenic diet in siblings of young women with cholelithiasis. *Mayo Clin Proc.* 1972: 47: 762-6.
- Gilat T, Feldman C, Halpern Z, et al. An increased familial frequency of gallstones. *Gastroenterology* 1983: 84: 242-6.
- Van der Linden W, Simonson N. Familial occurrence of gallstone disease. Incidence in parents of young sisters. *Human Heredity* 1973: 23: 123-7
- Kesaniemi YA, Koskenvuo M, Vuoristo M, et al. Biliary lipid composition in monozygotic and dizygotic pairs of twins. *Gut* 1989: 30: 1750-6
- Shoda J, Tanaka N, Matsuzaki Y, et al. Microanalysis of bile acid composition in intrahepatic calculi and its etiological significance. *Gastroenterology* 1991: 101: 821-30.
- Lammert F, Carey MC, Paigen B. Chromosomal organization of candidate genes involved in cholesterol gallstone formation: a murine gallstone map. *Gastroenterology* 2001: 120: 221-38.
- 81. Moschetta A, vanBerge-Henegouwen GP, Portincasa P, van Erpecum KJ, Palasciano G, van Erpecum KJ. Differential distribution of sphyngomyelin and phosphatidylcholine between micellar and vesicular phases: potential implications for lipid secretion. In: vanBerge-Henegouwen GP, Keppler D, Leushner U, Paumgartner G, Stiehl A, eds. Biology of bile acids in health and disease. Dordrecht: Kluwer Academic Publishers, 2001: 121-7.
- Chung DS, Benedek GB, Konikoff FM, et al. Elastic free energy of anisotropic helical ribbons as metastable intermediates in the crystallization of cholesterol. Proc Natl Acad Sci USA 1993: 90: 11341-5.

