

ANNALS OF HEPATOLOGY

Volume **4**

Number **1**

January-March **2005**

Artículo:

Current concepts in autoimmune hepatitis

Copyright © 2005:
Mexican Association of Hepatology

Otras secciones de
este sitio:

-  [Índice de este número](#)
-  [Más revistas](#)
-  [Búsqueda](#)

*Others sections in
this web site:*

-  [Contents of this number](#)
-  [More journals](#)
-  [Search](#)



Medigraphic.com

Concise Review

Current concepts in autoimmune hepatitis

Albert J. Czaja¹

Abstract

Autoimmune hepatitis has a global occurrence, diverse clinical phenotype, and evolving treatment options. The goals of this report are to review the codified diagnostic criteria, spectrum of clinical presentations, proposed pathogenic mechanisms, conventional treatment strategies, and promising interventions. The literature published in English from 1980-2005 was reviewed and an updated current perspective provided. Autoimmune hepatitis affects all ages, may be asymptomatic, frequently has an acute onset, and can present as fulminant hepatitis. Perivenular (zone 3) necrosis is within the histological spectrum. Autoimmune hepatitis can recur or develop *de novo* after liver transplantation. CD4+ T-helper cells and natural killer T cells have been implicated in the pathogenesis, and molecular mimicry may break self-tolerance. *DRB1*0301* and *DRB1*0401* are the susceptibility alleles among white North Americans and northern Europeans, whereas diverse alleles of HLA DR4 have been associated with the disease in Japan, mainland China, and Mexico. *DRB1*1301* is associated with autoimmune hepatitis in South American children, and it may predispose to an indigenous etiologic agent. Antibodies to soluble liver antigen/liver pancreas may have prognostic importance, and cyclosporine and mycophenolate mofetil must be assessed by clinical trial before incorporation into management algorithms. Site-specific interventions are feasible, and they require a confident experimental animal model for evaluation. Variant syndromes lack diagnostic and therapeutic guidelines. In conclusion, autoimmune hepatitis must be considered in all patients with acute and chronic liver disease and those with allograft dysfunction after transplantation. New immunosuppressive agents and site-specific interventions promise to improve care.

Key words: Autoimmunity, hepatitis, genetic predispositions, variants, treatment

Autoimmune hepatitis is an unresolving inflammation of the liver of unknown cause.¹ It is characterized by interface hepatitis on histological examination, hypergammaglobulinemia, and autoantibodies. The *sine qua non* of the diagnosis is the presence of interface hepatitis on histological examination. Lymphocytic, often lymphoplasmacytic, inflammatory infiltrates extend from portal tracts into parenchymal tissue where they are associated with hepatocyte injury (*Figure 1*). Parenchymal inflammation may be limited to periportal areas (interface hepatitis), or it may involve the entire acinus (panacinar or lobular hepatitis) (*Figure 2*). Plasma cells can be seen within the infiltrate (*Figure 3*), but they are not specific or essential for the diagnosis. Plasma cells in groups or sheets in the portal tracts are present in 66% of patients, and this finding in conjunction with moderate-to-severe interface hepatitis and/or panacinar hepatitis has a diagnostic specificity of 81% and positive predictability of 68%.²

Occurrence and ethnic variability

Autoimmune hepatitis has a global occurrence, and it has been described in African Americans, native Alaskans, Arabs, Asians, Europeans, Iranians, South Americans, and subcontinental Indians. Its incidence among white northern Europeans is 1.9 cases per 100,000 persons per year, and its point prevalence is 16.9 cases per 100,000 persons per year.^{3,4} In the United States, autoimmune hepatitis affects 100,000 to 200,000 persons, and it accounts for 5.9% of the liver transplantations in this country and 2.6% of the liver transplantations in Europe.⁵ The frequency of autoimmune hepatitis among patients with chronic liver disease in North America is between 11% and 23%. Its prevalence among Alaskan natives is 43 per 100,000 population and higher than that reported elsewhere.⁶

Race may affect disease severity as well as occurrence. Cirrhosis is present at accession more commonly in black North American patients with autoimmune hepatitis than in white North American patients (85% *versus* 38%), and hepatic synthetic function is decreased more frequently.⁷ Alaskan natives have a higher frequency of acute icteric disease, asymptomatic illness, and advanced fibrosis at presentation than non-native counterparts.⁶ Japanese patients typically have mild, late onset disease that can respond to non-steroidal medication such as ursodeoxycholic acid.⁸ South American patients are younger than white

¹ From the Division of Gastroenterology and Hepatology, Mayo Clinic College of Medicine, Rochester, Minnesota.

Address for correspondence:
Albert J. Czaja, M.D.
Mayo Clinic 200 First Street S.W. Rochester, Minnesota 55905
Telephone: 507-284-8118, Fax: 507-284-0538.
E-mail: czaja.albert@mayo.edu

North American counterparts, and they have more severe laboratory abnormalities.⁹ African, Asian and Arab patients have early age onset disease, and they have a higher frequency of cholestatic laboratory findings, greater occurrence of biliary changes on histological examination, and poorer initial response to standard therapy than other ethnic groups.¹⁰ These findings suggest that geographical location and ancestry affect occurrence and behavior of the disease. Interwoven into the natural history of the disease in each racial group and geographical region are cultural and socioeconomic factors that remain unsorted. Differences in the consequences of the liver disease must be correlated with delays in diagnosis or difficulties in accessing medical care that are region-specific.¹¹

Diagnostic clinical criteria

The clinical diagnosis of autoimmune hepatitis has been codified by an international panel, and these criteria must be applied to all patients¹² (*Table I*). An acute, even fulminant, presentation is recognized and important to diagnose quickly and treat promptly.¹³⁻¹⁶ The histological patterns that characterize acute onset autoimmune hepatitis are a panacinar hepatitis that resembles an acute viral or drug-induced hepatitis^{16,17} (*Figure 2*) and a centrilobular or perivenular (Rappaport zone 3) hepatitis that resembles an acute toxic injury^{16,18,19} (*Figure 4*).

Transitions from a perivenular (Rappaport zone 3) hepatitis to an interface hepatitis have been demonstrated



Figure 1. Interface hepatitis. Mononuclear inflammatory infiltrate disrupts limiting plate of portal tract and extends into parenchymal tissue. Hematoxylin and eosin. Original magnification, x100.

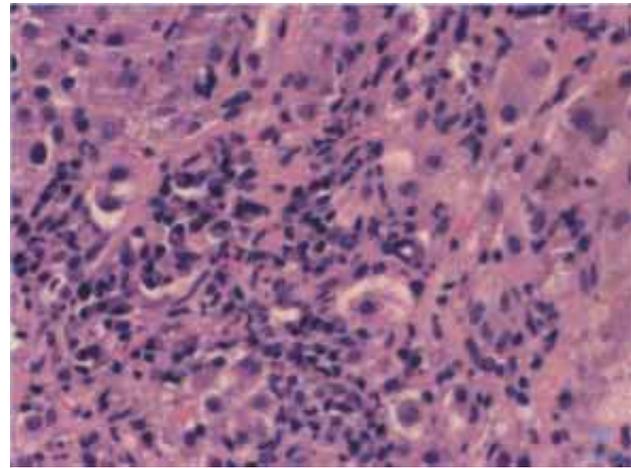


Figure 3. Plasma cells. The hyperchromatic nuclei of plasma cells are attenuated by a cytoplasmic halo. Hematoxylin and eosin. Original magnification, x200.

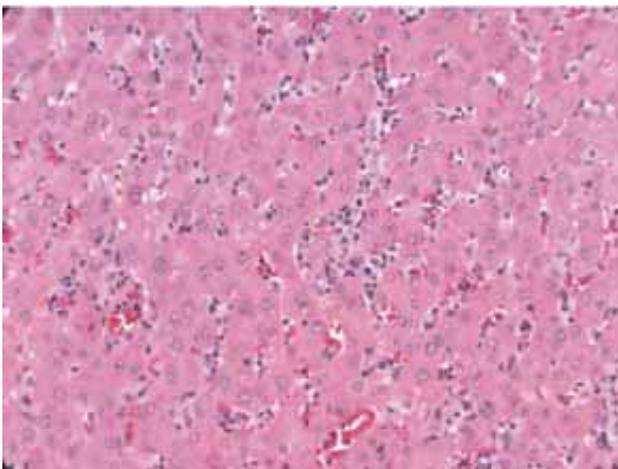


Figure 2. Pancinar hepatitis. Mononuclear inflammatory infiltrates line the sinusoidal spaces in association with liver cell degenerative and regenerative changes. Hematoxylin and eosin. Original magnification, x200.

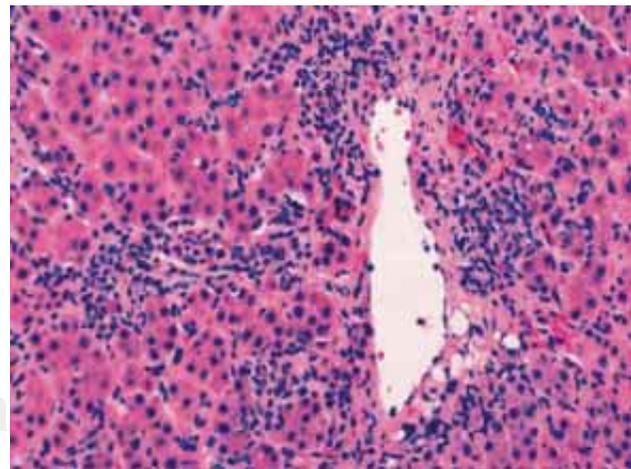


Figure 4. Perivenular (Rappaport zone 3) necrosis. Inflammation and hepatocyte drop out are present around a terminal hepatic venule. Mild diffuse inflammation and disorganization of the hepatic plate architecture are also present. Hematoxylin and eosin. Original magnification, x200.

in successive biopsy specimens from patients with acute onset disease.¹⁹ These observations suggest that the perivenular (Rappaport zone 3) pattern of injury may be an early histological manifestation of autoimmune hepatitis that is unrecognized in specimens obtained later in the course. An exacerbation of a previously unrecognized chronic liver disease may also have an acute presentation, and it should be suspected by the presence of hypoalbuminemia, thrombocytopenia, ascites, esophageal varices, and/or bridging (septal) fibrosis or cirrhosis on histological examination.^{17,20}

There are no disease-specific clinical or histological features. The diagnosis of autoimmune hepatitis requires the confident exclusion of other similar disorders including Wilson disease, genetic hemochromatosis, α -1 antitrypsin deficiency, chronic viral hepatitis, drug-related chronic liver disease, primary biliary cirrhosis (PBC), and primary sclerosing cholangitis (PSC)¹² (Table I). Minocycline is the drug that has been most commonly implicated as a cause of the syndrome.²¹ A cholestatic form of autoimmune hepatitis is not recognized, and the presence of pruritus or hyperpigmentation compels another diagnosis.^{1,12}

Diagnostic scoring criteria

An international scoring system for the objective diagnosis of the disease has been developed and validated retrospectively^{12,22,23} (Table II). Multiple clinical, laboratory and histological features are graded, and a composite score is derived both before and after corticosteroid treatment. The scoring system was developed as a research tool to ensure comparability between patient populations in clinical trials. It is not a discriminative diagnostic index, and it should not be used to distinguish between classical syndromes of chronic liver disease or to deduce that common non-disease-specific clinical and laboratory features connote a mixed or hybrid pathological state.

The virtues of the scoring system are that it quantifies the diagnosis, facilitates objective comparisons between patient populations, and accommodates individuals with atypical manifestations. Its drawbacks are its complexity, and its failure to consistently distinguish cholestatic syndromes from autoimmune hepatitis. The sensitivity of the scoring system for autoimmune hepatitis ranges from 97%-100%, and its specificity for excluding autoimmune hepatitis in patients with chronic hepatitis C ranges from 66%-92%. Its diagnostic specificity for excluding autoimmune hepatitis in cholestatic syndromes ranges from 45%-65%.¹²

Types

Three types of autoimmune hepatitis have been proposed based on serological markers, but only two types have distinctive clinical phenotypes²⁴ (Table III). None has been ascribed a unique cause, individual management strategy or special behavior, and they have not been endorsed as separate entities by the International Autoimmune Hepatitis Group. The designations are used mainly as clinical descriptors.

Type 1 autoimmune hepatitis is the most common form worldwide, constituting 80% of all cases, and it is characterized by the presence of antinuclear antibodies (ANA) and/or smooth muscle antibodies (SMA) (Table III). Seventy-eight percent of patients are female, and the female:male ratio is 3.5. Earlier reports that autoimmune hepatitis had a bimodal age distribution between ages 10 years and 30 years and between 40 years and 50 years were probably affected by referral patterns to tertiary medical centers.²⁵ Current experiences suggest that autoimmune hepatitis occurs as commonly across all age ranges and that it may be under-diagnosed in the elderly.^{26,27} Forty-eight percent of patients are less than 40 years old, and the disease can affect infants.

Table I. International criteria for the diagnosis of autoimmune hepatitis.

Diagnostic Criteria	
Definite AIH	Probable AIH
Normal α -1 AT phenotype	Partial α -1 AT deficiency
Normal ceruloplasmin level	Nondiagnostic ceruloplasmin/copper levels
Normal iron and ferritin levels	Nondiagnostic iron and/or ferritin changes
No active hepatitis A, B and/or C infection	No active hepatitis A, B and/or C infection
Daily alcohol <25 g/day	Daily alcohol <50 g/day
No recent hepatotoxic drugs	No recent hepatotoxic drugs
Predominant serum AST/ALT abnormality	Predominant serum AST/ALT abnormality
Globulin, γ -globulin or IgG level ≥ 1.5 times upper limit of normal	Hypergammaglobulinemia of any degree
ANA, SMA, or anti-LKM1 $\geq 1:80$ in adults and $\geq 1:20$ in children; no AMA	ANA, SMA or anti-LKM1 $\geq 1:40$ in adults; other autoantibodies
Interface hepatitis, moderate to severe	Interface hepatitis, moderate to severe
No biliary lesions, granulomas or prominent changes suggestive of another disease	No biliary lesions, granulomas or prominent changes suggestive of another disease

AIH = autoimmune hepatitis; α -1 AT = alpha-1 anti-trypsin; ANA = antinuclear antibodies; SMA = smooth muscle antibodies; anti-LKM1 = antibodies to liver/kidney microscope type 1; AMA = antimitochondrial antibodies; IgG = serum immunoglobulin G level.

An abrupt onset of symptoms occurs in 40%, and a fulminant presentation is possible.¹⁶ Thirty-eight percent of individuals have concurrent immune diseases, especially autoimmune thyroiditis, synovitis, or ulcerative colitis, and 25% have cirrhosis already established at the time of presentation.²⁸ Cholangiography is warranted in patients with ulcerative colitis to exclude PSC. Forty-one percent will have abnormal cholangiograms that support the diagnosis of PSC, and this finding may explain a refractory response to corticosteroid therapy.²⁹ The high frequency of cirrhosis at presentation indicates that type 1 autoimmune hepatitis has an indolent, aggressive stage.

Thirty-four percent of patients with type 1 autoimmune hepatitis may be asymptomatic at initial consultation, and they are most commonly men with lower serum aminotransferase and immunoglobulin levels than symptomatic patients.³¹ Histological features are similar between asymptomatic and symptomatic patients, and both groups respond as well to corticosteroids. Most asymptomatic patients become symptomatic during follow-up, and differences between the asymptomatic and symptomatic state may reflect variations in disease activity and patient tolerance.

Type 2 autoimmune hepatitis is characterized by the presence of antibodies to liver/kidney microsome type 1 (anti-LKM1)³¹ (Table III). This disease occurs mainly in children, but 20% of patients with type 2 disease in Europe are adults. Concurrent immune diseases are also common, especially insulin-dependent diabetes mellitus, vitiligo, and autoimmune thyroiditis. Organ specific au-

toantibodies are frequent, including antibodies to parietal cells, islets of Langerhans, and thyroid. As in type 1 disease, a fulminant presentation is possible and important to recognize early.¹⁴

Type 2 autoimmune hepatitis is the only form in which the target autoantigen has been identified. It is the cytochrome mono-oxygenase, CYP2D6, which is an important drug-metabolizing enzyme within the cytosol of the hepatocyte.³²⁻³⁴ The antigen has been sequenced, cloned, and mapped, and five antigenic sites located between peptides 193-212, 257-269, 321-351, 373-389, and 410-429 are recognized by anti-LKM1.³⁵ The amino acid sequence spanning 193-212 of the CYP2D6 molecule is the target of anti-LKM1 in 93% of patients.

Homologies have been recognized between epitopes on the CYP2D6 molecule and the genome of the hepatitis C virus (HCV).³⁴⁻³⁷ The detection of anti-LKM1 in occasional patients with chronic hepatitis C in Europe ($\leq 10\%$) may reflect this molecular mimicry and antibody cross-reactivity. The hexameric amino acid sequence spanning 193-212 of the CYP2D6 molecule is homologous to the sequence spanning region 2985-2990 of the HCV genome and identical to the sequence spanning region 130-135 of the cytomegalovirus (CMV) genome.³⁵ These homologies suggest that multiple exposures to viruses mimicking self may be a mechanism by which to break self-tolerance and induce type 2 autoimmune hepatitis. Antibodies to LKM1 are extremely rare in North American patients with chronic hepatitis C, and this rarity may reflect differences in the indigenous virus or the genetic

Table II. International scoring system for diagnosis of autoimmune hepatitis.

Gender	Female	+2	HLA	DR3 or DR4	+1
AP:AST (or ALT) ratio	>3	-2	Immune disease	Thyroiditis, colitis, others	+2
	<1.5	+2	Other markers	Anti-SLA/LP, actin, LC1, pANCA	+2
γ -globulin or IgG level above normal	>2.0	+3			
	1.5-2.0	+2			
	1.0-1.5	+1			
	<1.0	0			
ANA, SMA, or anti-LKM1 titers	>1:80	+3	Histological features	Interface hepatitis	+3
	1:80	+2		Plasmacytic	+1
	1:40	+1		Rosettes	+1
	<1:40	0		None of above	-5
				Biliary changes	-3
				Other features	-3
AMA	Positive	-4	Treatment response	Complete	+2
				Relapse	+3
Viral markers	Positive	-3			
	Negative	+3			
Drugs	Yes	-4	Pretreatment score		
	No	+1	Definite diagnosis	>15	
			Probable diagnosis	10-15	
Alcohol	<25 g/day	+2	Post-treatment score		
	>60 g/day	-2	Definite diagnosis	>17	
			Probable diagnosis	12-17	

AP:AST (or ALT) ratio=ratio of alkaline phosphatase level to aspartate or alanine aminotransferase level; anti-SLA/LP=antibodies to soluble liver antigen/liver pancreas; anti-LC1=antibodies to liver cytosol type 1; pANCA=perinuclear anti-neutrophil cytoplasmic antibodies; IgG=immunoglobulin G; ANA=antinuclear antibodies; SMA=smooth muscle antibodies; anti-LKM1=antibodies to liver/kidney type 1; AMA=antimitochondrial antibodies; and HLA=human leukocyte antigen.

Table III. Types of autoimmune hepatitis

Clinical features	Type 1	Type 2
Signature markers	SMA, ANA	Anti-LKM1
Autoantigen	Unknown	CYD2D6
Age (years)	Infancy to old age	Pediatric (2-14)
Female (%)	78	89
Acute or fulminant onset	Yes	Yes
Immune diseases (%)	38	34
Typical concurrent immune diseases	Thyroiditis Graves' disease Ulcerative colitis	Thyroiditis Vitiligo Type 1 diabetes APECED
HLA associations	B8, DR3, DR4	B14, DR3, C4A-Q0, DR7
Allelic risk factors	<i>DRB1*0301, DRB1*0401</i> <i>DRB1*0404, DRB1*0405</i> <i>DRB1*1301</i> <i>DRB1*1501</i> (protective)	<i>DRB1*07</i>
Autoimmune promoter genes (putative)	<i>Tumor necrosis factor-α</i> <i>Cytotoxic T lymphocyte antigen 4 (CTLA-4)</i> <i>Vitamin D receptor (VDR)</i> <i>Tyrosine phosphatase CD45</i> <i>Fas</i> <i>MHC class I chain-related A (MICA)</i>	Unknown
Steroid responsive	+++	++

SMA = smooth muscle antibodies; ANA = antinuclear antibodies; anti-LKM1 = antibodies to liver/kidney microsome type 1; CYD2D6 = cytochrome 2D6; APECED = polyendocrinopathy-candidiasis-ectodermal dystrophy; MHC = major histocompatibility complex.

susceptibility of the host.^{38,39} Studies in Germany⁴⁰ and Italy⁴¹ have not found an association between structural changes within the viral genome and the presence of anti-LKM1. A host factor for anti-LKM1 expression has been implicated.⁴⁰

Type 2 autoimmune hepatitis occurs in 15% of patients with autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED).⁴² This syndrome consists of multiple endocrine organ failure, mucocutaneous candidiasis, and ectodermal dystrophy in various syndromic combinations that may include autoimmune hepatitis. APECED is caused by a single-gene mutation located on chromosome 21q22.3 which affects the generation of the autoimmune regulator (AIRE).⁴³ AIRE is a transcription factor that is expressed in epithelial and dendritic cells within the thymus, and it regulates clonal deletion of autoreactive T cells (negative selection). APECED has an autosomal recessive pattern of inheritance, and it lacks HLA-DR associations and female predilection. The autoantigens associated with APECED are CYP1A2 and CYP2A6.

Type 3 autoimmune hepatitis is the least established form, and it is characterized by the presence of antibodies to soluble liver antigen/liver-pancreas (anti-SLA/LP).^{44,45} These antibodies are directed against a 50 kDa cytosolic protein⁴⁶ which is a transfer RNA complex (tRNP^{(ser)sec}) involved in the incorporation of selenocysteine into polypeptide chains.⁴⁷ Patients with type 3 disease are mostly women (91%) with a mean age of 37 years (range, 17 to 67 years).⁴⁴ Other autoantibodies, including ANA, SMA and anti-LKM1, can co-exist with anti-SLA/LP, and only 26% of patients have anti-SLA/LP as their sole

serological finding.⁴⁴ Patients with anti-SLA/LP are indistinguishable from patients with type 1 autoimmune hepatitis by clinical or laboratory features, HLA phenotype, or response to corticosteroids.^{48,49} The designation of a type 3 autoimmune hepatitis has been largely abandoned.

Pathogenic mechanisms

The pathogenic mechanisms of autoimmune hepatitis are unknown. The most popular hypotheses evoke a constellation of interactive factors that include a triggering agent, a genetic predisposition, and various determinants of autoantigen display, immunocyte activation, and effector cell expansion.^{50,51} Multiple triggering factors have been proposed, and they include infectious agents, drugs, and toxins. The multiplicity of etiologic agents that have been implicated in the pathogenesis of the disease suggests that the triggering epitope is a short amino acid sequence that is common in many antigens. There can be a long lag time between exposure to the trigger and onset of the disease, and the triggering factor is not needed for perpetuation of the disorder.

The CD4+ T-helper cell is the principal effector cell, and its activation and differentiation are the initial steps in the pathogenic pathway.^{50,51} (Figure 5). Natural killer T (NKT) cells are abundant in the liver, and they have also been implicated in the pathogenesis of the disease. NKT cells are produced in the bone marrow rather than the thymus, lack antigen-specific receptors, and produce interferon (IFN)- γ and tumor necrosis factor (TNF)- α . They are inhibited by cells with normal expression of the major histocompatibility

complex (MHC) and by inhibitory receptors activated by glycolipid. Conversely, they target cells with aberrant MHC expression, defend against cells altered by viruses or cancer, and seem to promote hepatic regeneration.

The immunoregulatory cytokines orchestrate immunocyte differentiation through cross-regulatory actions and result in cellular and humoral mechanisms of liver cell injury⁵⁰⁻⁵² (Figure 5). Interleukin (IL)-2, IFN- γ , and TNF- α constitute the type 1 (Th1) cytokine response which regulates cellular immune mechanisms by facilitating clonal expansion of cytotoxic T lymphocytes. Interleukin-4, IL-5, IL-6, IL-8, IL-10 and IL-13 constitute the type 2 (Th2) cytokine response which influences the humoral immune response by activating B cells and stimulating autoantibody production. The type 1 cytokine response favors liver injury by expanding sensitized tissue-infiltrating cytotoxic T cells (cellular cytotoxicity), and the type 2 cytokine response favors liver cell injury by generating immunoglobulin complexes on the hepatocyte surface that are targeted by NKT cells (antibody-dependent cell-mediated cytotoxicity). The type 2 cytokine response also has anti-inflammatory effects that counter the type 1 cytokine actions.

Molecular mimicry of a foreign antigen and a self-antigen is the most common explanation for the loss of self-tolerance, but this mechanism has not been established in human disease.⁵¹ Cross-reacting autoantibodies between foreign and self antigens have been described, but cross-reacting immunocytes have been more difficult to demonstrate. Recently, a murine model of type 2 autoimmune hepatitis based on DNA immunization against self-antigens has supported this possibility.⁵³ Most immunized

mice developed peak serum alanine aminotransferase abnormalities 4 and 7 months after the last of three plasmid injections that contained the antigenic regions of human CYP2D6 and human formiminotransferase cyclodeaminase, the target antigen of antibodies to liver cytosol type 1 (anti-LC1). Affected mice expressed anti-LKM1 and anti-LC1, but they also had cytotoxic T lymphocytes within the liver that were sensitized against the antigens in the plasmid constructs. This murine model indicated that DNA immunization against human autoantigens could break self-tolerance and cause liver injury by molecular mimicry between foreign and self-antigens involving cross-reacting humoral and cellular responses.

HLA Associations

Genetic factors affect the occurrence, clinical expression and treatment outcome of type 1 autoimmune hepatitis. HLA DR3 is the main susceptibility factor in white northern European and North American patients, and HLA DR4 is a secondary but independent risk factor⁵⁴ (Table III). Eighty-five percent of white patients from these regions have HLA DR3, DR4 or DR3-DR4. Different geographical regions and ethnic groups have different susceptibility factors. HLA DR3 occurs rarely in the Japanese population, and HLA DR4 is the principal risk factor for autoimmune hepatitis in this ethnic group.⁵⁵ HLA DR4 is also the principal susceptibility factor for autoimmune hepatitis in mainland China.⁵⁶ In contrast, HLA DR3, but not HLA DR4, is the susceptibility factor in Italy,⁵⁷ and HLA DR13 is associated with childhood autoimmune hepatitis in South America.^{9,58-60}

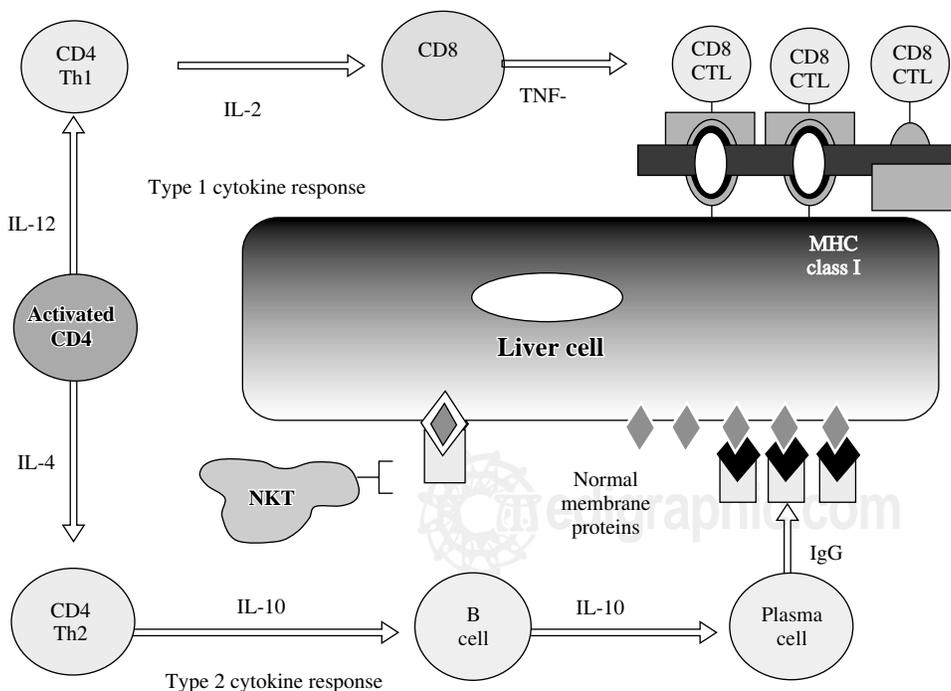


Figure 5. Cytokine pathways of immunocyte differentiation. The principal effector is the CD4+ T helper cell, and its differentiation depends on the counter-regulatory effects of the interleukins (IL) and tumor necrosis factor- α (TNF- α). The type 1 cytokine response promotes clonal expansion of liver infiltrating cytotoxic T lymphocytes (CD8 CTL). The type 2 cytokine response promotes the expansion of plasma cells and immunoglobulin G (IgG) production. The immunoglobulin complexes with normal membrane proteins on the hepatocyte surface, and natural killer T (NKT) cells target these complexes and cause cytotoxicity by an antibody-dependent cell-mediated form of cytotoxicity. Type 1 and type 2 cytokine responses are counter-regulatory.

Patients with HLA DR4 in North America are older and more commonly female than patients with HLA DR3.⁶¹ They also have higher serum levels of γ -globulin and immunoglobulin G (IgG), higher titers of ANA, and a greater frequency of concurrent immune diseases. Patients with HLA DR4 respond more readily to corticosteroid therapy than those with HLA DR3 by entering remission more commonly and failing treatment less often. The bases for these effects on clinical phenotype and outcome are unknown, but they may relate to the diversity of alleles associated with each susceptibility factor.

HLA DR3 is associated with only 2 alleles that might affect autoreactivity, and *DRB1*0301* is the only allele common in the United States.⁶² In contrast, HLA DR4 is associated with 26 alleles that may affect autoreactivity, of which 10 are common in the United States. HLA DR4-positive patients have a greater diversity of susceptibility alleles, and they can present a wider spectrum of antigens to immunocytes than patients with HLA DR3. This diversity of antigenic presentation may in turn enhance the spectrum of clinical manifestations and the frequency of concurrent immune diseases.

The genetic associations of type 2 autoimmune hepatitis are not well defined because the disease is infrequent in some geographical regions and type 1 autoimmune hepatitis is the predominant form in all experiences. Pa-

tients with type 2 autoimmune hepatitis from Germany have *DRB1*03* and *DRB1*04* less commonly and *DRB1*07* more frequently than white North American patients with type 1 autoimmune hepatitis and normal control subjects⁶³ (Table III). *DRB1*07* has also been associated with the disease in Brazil,⁶⁴ whereas HLA B14, HLA DR3 and *C4A-Q0* have been incriminated as genetic risk factors in northern Europe.⁶⁵ Recent studies have suggested that the expression of anti-LKM1 may be associated with *DRB1*07* regardless of the autoimmune or viral basis of the liver disease.⁶⁶ Host-specific genetic factors in addition to disease-specific etiologic agents probably contribute to the production of anti-LKM1 and influence its occurrence in different geographical areas and racial groups. Unlike type 1 autoimmune hepatitis, the HLA phenotype of type 2 autoimmune hepatitis has not yet been ascribed a clinical relevance.

Allelic associations

High resolution DNA-based techniques have indicated that the alleles associated with susceptibility, clinical expression and outcome in white northern European and North American patients with type 1 autoimmune hepatitis are *DRB1*0301* and *DRB1*0401*.⁶⁷ These findings implicate the *DRB1* locus as the principal susceptibility re-

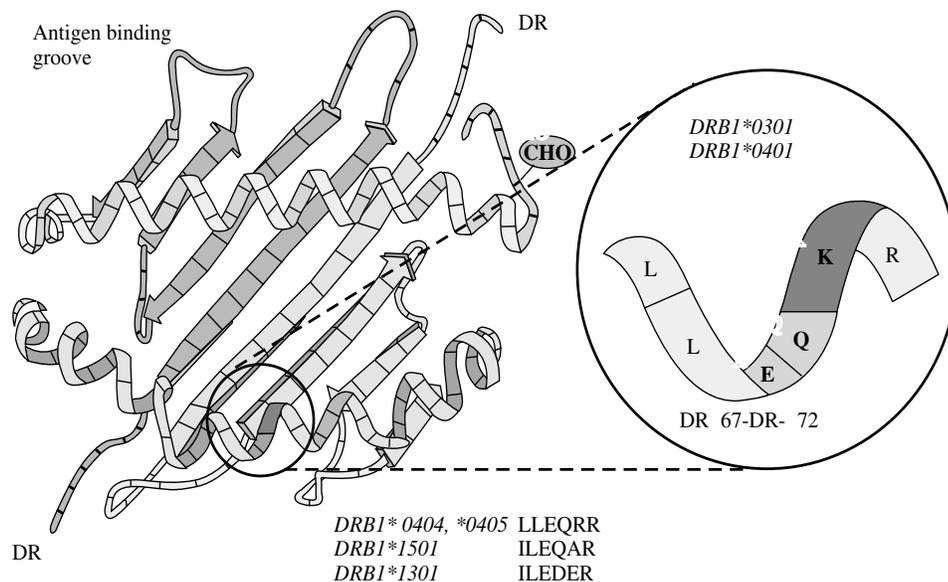


Figure 6. Key susceptibility sequence within the antigen binding groove of the class II MHC molecule. The six amino acid sequence between positions 67 and 72 on the DR β chain of the class II MHC molecule (insert) is the critical sequence encoded by the susceptibility alleles. *DRB1*0301* and *DRB1*0401* encode the identical sequence of LLEQKR (insert) where lysine (K) is the critical determinant at position DR β 71. *DRB1*0404* and *DRB1*0405*, which are the susceptibility alleles in Mexican, Japanese, mainland Chinese and Argentine adults, encode a similar sequence except for an arginine (R) at the DR β 71 position. *DRB1*1501*, which protects against type 1 autoimmune hepatitis in white North Americans and northern Europeans, encodes an isoleucine (I) for leucine (L) at position DR β 67 and an alanine (A) for lysine (K) at position DR β 71. *DRB1*1301*, which is associated with type 1 autoimmune hepatitis in Argentine children and Brazilian patients, encodes ILEDER at positions DR β 67-72 where glutamic acid (E), aspartic acid (D), and glutamic acid (E) are at positions DR β 69, 70 and 71, respectively. The structural and electrostatic properties of the antigen binding groove determine the antigens that can be presented.

gion of the MHC. Patients with *DRB1*0301* are younger than patients with *DRB1*0401*, and they fail corticosteroid therapy more often, die of liver failure or require liver transplantation more commonly, and have a significantly greater frequency of an adverse treatment outcome than patients with *DRB1*0401*.⁶⁸

The risk of type 1 autoimmune hepatitis may relate to amino acid sequences in the antigen binding groove of the class II MHC molecule, and multiple alleles may encode the same or similar sequence^{69,70} (Figure 6). The critical shared motif in white North Americans and northern Europeans with type 1 autoimmune hepatitis is a six-amino-acid sequence represented by the code, LLEQKR.^{67,71} This sequence is located between positions 67 and 72 of the DR β polypeptide chain of the class II MHC molecule, and lysine (K) in position 71 is the critical determinant of susceptibility.

*DRB1*0301* and *DRB1*0401* encode identical amino acid sequences in the DR β 67-72 region, and they affect susceptibility similarly.⁶⁷ *DRB1*0404* and *DRB1*0405* are the susceptibility alleles in Mexican,⁷² Japanese,⁵⁵ mainland Chinese⁵⁶ and Argentine adults,^{58,59} and they encode a similar sequence except for an arginine (R) for lysine (K) at the DR β 71 position. Arginine is a positively charged amino acid that is structurally similar to lysine, and its substitution for lysine would not greatly alter the antigen binding properties of the class II MHC molecule.

In contrast, *DRB1*1501* protects against type 1 autoimmune hepatitis in white North Americans and northern Europeans, and this allele encodes an isoleucine (I) for leucine (L) at position DR β 67 and an alanine (A) for lysine (K) at position DR β 71^{67,71} (Figure 6). Alanine is a neutral, nonpolar amino acid whose substitution for lysine would greatly affect antigen presentation and immunocyte activation. As in other autoimmune diseases such as type 1 diabetes mellitus, the substitution of a single amino acid at a critical location in the antigen binding groove of the class II MHC molecule may affect disease occurrence. By understanding the requirements for optimal autoantigen presentation, it is possible to predict the ideal antigenic peptide. This ideal peptide must have a negatively charged residue, either aspartic acid or glutamic acid, at position P4 from the N terminus to complement the positively charged residue at position DR β 71 (either lysine or arginine).⁷⁰

*DRB1*1301* is associated with type 1 autoimmune hepatitis in Argentine children and Brazilian patients, and it encodes ILEDER at positions DR β 67-72.^{70,73} Glutamic acid (E), aspartic acid (D), and glutamic acid (E) are at positions DR β 69, 70 and 71, respectively, in the class II MHC molecule. These critically located but negatively charged amino acid residues create a different antigen-presenting milieu within the class II MHC molecule than that encoded by the *DRB1*0301* and *DRB1*04* alleles. These findings have generated a “molecular footprint hypothesis” of pathogenesis which holds that susceptibility to type 1 autoimmune hepatitis in different regions and

racial groups relates to indigenous factors or agents favored by certain genetic phenotypes.⁷³

In South America, *DRB1*1301* is associated with protracted hepatitis A virus infection,⁷⁴ and individuals with this allele may be selected from their environment to have prolonged exposure to viral and hepatic antigens that favor the development of autoimmune hepatitis.⁷⁵ The individual susceptibility allele in a geographic region may be a “footprint” by which to track the cause of the disease.

Autoimmune promoters

Genetic autoimmune promoters inside and outside the MHC may also affect the occurrence of autoimmune hepatitis either in synergy with the principal susceptibility alleles (epistasis) or in lieu of them⁷³ (Table III). Polymorphisms of the *tumor necrosis factor- α gene (TNFA*2)*^{76,77} and the *cytotoxic T lymphocyte antigen 4 gene (CTLA-4)*^{78,79} have been associated with increased immune reactivity and disease severity in type 1 autoimmune hepatitis in white North American and northern European patients. These promoters are host-related and not disease-specific. Constellations of them in varying combinations may affect the occurrence, clinical phenotype, and outcome of autoimmune hepatitis. Their occurrence and impact may also vary by geographical region and ethnic group.⁸⁰

Other genetic promoters that have been implicated in the pathogenesis of autoimmune hepatitis include polymorphisms of the *vitamin D receptor (VDR) gene*,⁸¹ point mutation of the *tyrosine phosphatase CD45 gene*,⁸² polymorphisms of the *Fas gene (tumor necrosis factor receptor super family-6 or TNFRSF6)*,⁸³ and polymorphisms of the *MHC class I chain-related A gene (MICA)*⁸⁴ (Table III). Cytokine imbalances perhaps related to genetic polymorphisms that control cytokine production and receptor function are undoubtedly important in affecting the cascade of immune-mediated interactions resulting in hepatocyte injury. In this context, transforming growth factor- β (TGF- β) has recently been implicated as an important protective mechanism against autoimmune hepatitis by suppressing infiltration of the liver with autoreactive T cells.⁸⁵

Gender effects

The female predisposition for autoimmune hepatitis and autoimmune disease in general is unexplained.⁸⁶ HLA DR4-positive women with type 1 autoimmune hepatitis have a greater variety of HLA DR4 alleles associated with their disease than HLA DR4-positive men.⁶² Women may thereby have a greater facility to be sensitized to self or foreign antigens than men. They may also be exposed to unique antigens, and/or they may respond to common antigens against which men do not react.

Triggering factors unique to the female state may include drugs, toxins, infections, and environmental antigens. Pregnancy is a unique state of women and protracted

exposures to fetal cells or antigens in the maternal circulation may stimulate autoreactivity through microchimerism. The diversity and strength of the antigenic stimulation in women may in turn be modulated by the repertoire of HLA DR4 alleles and various hormonal factors.

Estrogen levels do modulate immune reactivity, and they may be contributory to the autoimmune propensity in women.⁸⁷ High estrogen levels favor a type 2 cytokine response which drives activated immunocytes towards antibody production and an anti-inflammatory effect (Figure 5). Normal or low estrogen levels promote a type 1 cytokine response which drives the clonal expansion of tissue infiltrating cytotoxic T cells and causes liver damage (Figure 5).

The female propensity for autoimmune hepatitis is apparent among children and among pre- and post-menopausal adults.^{26,88} The presence or absence of estrogen, therefore, is an insufficient explanation for the risk of disease. Interactions between growth hormone, prolactin, testosterone and estrogen may constitute a changing hormonal milieu that affects immune responsiveness differently at various ages and favors certain antigens during different stages of maturation. The female gender may be the critical determinant affecting the hormonal blend of the interactive network at each age.

The importance of hormonal effects on the pathogenic mechanisms of autoimmune hepatitis is evident during pregnancy. Autoimmune hepatitis commonly improves during pregnancy, possibly because high estrogen levels promote a switch from type 1 cytokine actions which are

cytotoxic to type 2 cytokine actions which are anti-inflammatory. Autoimmune hepatitis may then worsen after delivery, possibly because low estrogen levels promote a switch back to the type 1 cytokine actions which are cytotoxic.⁸⁹ The role of other hormonal interactions during and after pregnancy in modulating this effect is unknown.

Autoantibodies

ANA, SMA, and anti-LKM1 constitute the standard repertoire of autoantibodies that are assessed in autoimmune hepatitis^{1,12,90-92} (Table IV). Antibodies to soluble liver antigen/liver pancreas (anti-SLA/LP), neutrophil cytoplasm (pANCA), endomysium, and tissue transglutaminase are ancillary markers that are useful in evaluating patients who are seronegative for the standard battery.⁹² Other autoantibodies that have been described in autoimmune hepatitis which are either not generally available, investigational in nature, or of limited clinical value are antibodies to asialoglycoprotein receptor (anti-ASGPR),⁹³ actin,⁹⁴ chromatin,⁹⁵ liver cytosol type 1,⁹⁶ double-stranded DNA,⁹⁷ histones,⁹⁸ *Saccharomyces cerevisiae*,^{99,100} and lactoferrin¹⁰¹ (Table IV).

New autoantibodies continue to be characterized in the hope that they will reveal critical pathogenic mechanisms or have prognostic value. Antibodies to SLA/LP are present in 26% of patients with autoimmune hepatitis who are otherwise seronegative,^{44,45} and they are useful in reclassifying cryptogenic chronic hepatitis as autoimmune hepatitis.¹⁰² These autoantibodies also identify individuals

Table IV. Autoantibodies associated with autoimmune hepatitis

Standard repertoire	Target (s)	Clinical value
Antinuclear antibodies	Centromere	Diagnosis of type 1
	Ribonucleoproteins	
Smooth muscle antibodies	Actin, tubulin, vimentin, desmin, skeleton	Diagnosis of type 1
Anti-liver kidney microsome type 1	CYP2D6	Diagnosis of type 2
Supplemental repertoire		
Perinuclear anti-neutrophil cytoplasmic antibodies	Possible nuclear membrane lamina	Diagnosis of type 1 Cryptogenic hepatitis
IgA antibodies to endomysium	Endomysium monkey esophagus	Celiac disease Cryptogenic hepatitis
Nonstandard repertoire		
Antibodies to actin	Microfilaments	Diagnosis of type 1
Antibodies to soluble liver antigen/liver pancreas	Ribonucleoprotein complex	Relapse Cryptogenic hepatitis
Anti-asialoglycoprotein receptor	Asialoglycoprotein receptor	Histologic activity Relapse
Anti-chromatin	Chromatin	Relapse
Anti-liver cytosol type 1	Formiminotransferase cyclodeaminase	Diagnosis of type 2 Treatment failure
Investigational repertoire		
Anti- <i>Saccharomyces cerevisiae</i>	Brewer's or baker's yeast	Crohn disease Celiac disease Autoimmune hepatitis
Anti- lactoferrin	Iron binding protein in granulocytes	Uncertain

who have more severe disease than seronegative counterparts and who invariably relapse after corticosteroid withdrawal.¹⁰³⁻¹⁰⁵ Since anti-SLA/LP are closely associated with HLA DR3, they may be surrogate markers of a genetic propensity for relapse or refractory disease.^{103,105} Analysis of the immunoprecipitated RNAs extracted from HeLa cell extracts is the most powerful, sensitive and specific method to detect anti-tRNP^{(ser)sec}/SLA/LP autoantibodies, but an enzyme-linked immunosorbent assay (ELISA) is available as a commercial kit and performance parameters between the methods are comparable.¹⁰⁶

Perinuclear anti-neutrophil cytoplasmic antibodies (pANCA) and immunoglobulin A (IgA) antibodies to endomysium (EMA) and tissue transglutaminase (tTG) are ancillary markers of autoimmune hepatitis that are available in the clinical laboratory^{91,92} (Table IV). pANCA are found with great frequency (50%-92%) and in high titer in type 1 autoimmune hepatitis, and they can be the sole serological markers of this disease.^{107,108} IgA EMA have a sensitivity of 94% and specificity of 99% for celiac disease,^{109,110} and they are less likely to be falsely positive in chronic hepatitis than IgA antibodies to tTG.^{100,111-113} Serological screening for celiac disease is important in patients with autoimmune hepatitis and in patients with chronic undifferentiated liver disorders since celiac disease can occur coincidentally with autoimmune liver disease^{114,115} or cause liver dysfunction that may improve with gluten restriction.¹¹⁶⁻¹¹⁹

The autoantibodies that are still investigational and that have promise as prognostic indices include anti-ASGPR and anti-actin (Table IV). The presence of anti-ASGPR correlates with histological activity and the propensity to relapse after corticosteroid withdrawal.^{93,120} Continuation of treatment until disappearance of anti-ASGPR has been associated with a sustained remission. Antibodies to actin identify patients with a higher frequency of treatment failure and death from liver failure or requirement for liver transplantation than seronegative patients, but they are restricted to those individuals with SMA.⁹⁴

Conventional treatment schedules

The preferred treatment schedule for all forms of autoimmune hepatitis in adults is prednisone in combination with azathioprine¹²¹⁻¹²³ (Table V). Prednisone alone in higher dose is as effective as the combination regimen, but it is associated with a higher frequency of drug-related side effects (44% versus 10%). The preferred treatment schedule in children is prednisone alone in a dose of 2 mg/kg daily (up to 60 mg daily).¹ Azathioprine or 6-mercaptopurine can also be introduced as a corticosteroid-sparing measure. Tapering schedules or alternate day corticosteroid regimens are commonly used in children to reduce deleterious effects on linear growth, bone development, and physical appearance.

Therapy is continued until remission, treatment failure, incomplete response, or drug toxicity.^{122,123} *Remission* implies the absence of symptoms, resolution of all laboratory indices of liver inflammation, and histological improvement to normal or minimal inflammatory activity. A serum aspartate aminotransferase level of less than twice the upper limit of normal is an acceptable laboratory end point if the histological examination confirms the absence of interface hepatitis. *Treatment failure* connotes deterioration during therapy, and it is characterized by worsening of the serum aspartate aminotransferase or bilirubin level by at least 67% of previous values, progressive histological activity, or onset of ascites or encephalopathy. *Incomplete response* connotes improvement that is insufficient to satisfy remission criteria after 3 years of continuous treatment, and *drug toxicity* implies severe intolerance of the medication. Histological improvement lags behind clinical and laboratory improvement by 3-8 months, and liver tissue examination prior to drug withdrawal is the only means of ensuring inactive disease.^{124,125}

Treatment outcomes

Sixty-five percent of treated patients satisfy remission criteria within 18 months, and 80% achieve this result

Table V. Conventional treatment regimens.

Weeks administered	Combination therapy		Prednisone therapy
	Prednisone (mg daily)	Azathioprine (mg daily)	Prednisone (mg daily)
1	30	50	60
1	20	50	40
2	15	50	30
Maintenance until end point	10	50	20
Relative contraindications	Cytopenia Pregnancy Active malignancy Short course Thiopurine methyltransferase deficiency		Post-menopausal state Osteoporosis Diabetes Hypertension Obesity Emotional lability

within 3 years.^{122,123} The average duration of treatment until remission is 22 months,²⁸ and the 10-year life expectancies of treated patients with and without cirrhosis at accession are 89% and 90%, respectively.¹²⁶ These survivals are comparable to those of an age- and sex-matched normal population from the same geographical region. Patients with cirrhosis respond as well to treatment as patients without cirrhosis, and they should be treated similarly with the same expectation of success.^{126,127} Twenty-one percent of individuals who enter remission sustain this result long term after drug withdrawal (median 7 interval of follow-up, 76 months), and an effort should be made to discontinue initial therapy in all patients with inactive disease.¹²⁸ Thirteen percent develop side effects that justify premature discontinuation of medication (drug toxicity); 9% deteriorate despite compliance with therapy (treatment failure); and 13% improve but not to a degree to satisfy criteria for remission (incomplete response).^{122,123}

Corticosteroid therapy may also reduce hepatic fibrosis.¹²⁹⁻¹³² Fibrosis scores improved in 56% of patients followed for 55±9 months, and they did not progress in 33% of patients followed for 62±14 months.¹³² Histological activity indices decreased concurrently, and patients in whom the histological activity indices improved had a higher frequency of improvement in the fibrosis scores (80% versus 25%, p=0.002).^{132,133} These findings suggest that improvement in hepatic fibrosis occurs in conjunction with reductions in liver inflammation and that corticosteroid therapy facilitates the disappearance of fibrosis by suppressing inflammatory activity. Small case studies have also suggested that cirrhosis can disappear during

treatment, but this possibility must await confirmation by assays more reliably reflective of cirrhosis than conventional needle biopsy of the liver.^{129,132}

Treatment of suboptimal responses

Treatment failure is managed by administering high dose prednisone alone (60 mg daily) or prednisone (30 mg daily) in conjunction with azathioprine (150 mg daily).^{122,123} Doses of prednisone and azathioprine are then reduced by 10 mg and 50 mg, respectively, for each month of laboratory improvement until conventional maintenance levels of drug are achieved. Seventy-five percent of patients treated in this fashion enter clinical and laboratory remission, but only 20% have histological resolution. These patients remain at risk for progressive liver disease and drug toxicity.

Drug toxicity requires premature dose reduction or discontinuation of the offending drug and continued use of the other tolerated medication in adjusted dose.^{122,123} Corticosteroid-related side effects are the most common causes for drug withdrawal, and they include intolerable cosmetic changes or obesity (47%), osteoporosis with vertebral compression (27%), brittle diabetes (20%), and peptic ulceration (6%).¹³⁴ Azathioprine can be administered as a corticosteroid-sparing agent with doses increased to 2 mg/kg daily. The emergence of a cholestatic hepatitis, pancreatitis, rash, progressive cytopenia, or gastrointestinal upset indicates azathioprine toxicity and the need for its withdrawal. The dose of prednisone is then adjusted to suppress disease activity.

Table VI. Evolving Drug Therapies and Site-specific Interventions.

Agent	New drug regimens	Possible Uses	Site-specific Interventions	
			Intervention	Mechanism(s)
Cyclosporine ¹⁵⁸⁻¹⁶⁵ (5-6 mg/kg daily)		Treatment failure Steroid intolerance First-line therapy	Blocking synthetic peptides ¹⁸⁰	Blocks 1 st co-stimulatory signal
Mycophenolate mofetil ^{167,168} (2 g daily)		Azathioprine intolerance Steroid withdrawal	Soluble cytotoxic T lymphocyte antigen-4 ¹⁸²	Blocks 2 nd co-stimulatory signal
6-mercaptopurine ¹⁷³ (1.5 mg/kg daily)		Azathioprine intolerance Treatment failure	Oral tolerance regimens ¹⁸³	Dose dependent suppression immune response
Tacrolimus ^{170,171} (4 mg twice daily)		Treatment failure Steroid intolerance	T cell vaccination ¹⁸⁴	Clonal depletion of activated cytotoxic T lymphocytes
Budesonide ¹⁷² (3 mg twice daily)		Front-line therapy Osteopenia	Cytokine manipulations ^{186,187}	Recombinant cytokines Monoclonal antibodies
Ursodeoxycholic acid ¹⁷⁵ (13-15 mg/kg daily)		Mild disease in Japanese	Gene therapy ¹⁸⁸	Limit fibrosis Promote repair Counter cytokines
Methotrexate ¹⁷⁶ Cyclophosphamide ¹⁷⁷		Treatment failure Steroid intolerance		
Deflazacort ¹⁷⁴ (7.5 mg for each 5 mg prednisone)		Steroid sparing		

An *incomplete response* is declared after 3 years of conventional therapy without remission.^{122,123} Patients improve but not to a degree to satisfy remission criteria, and they are at risk for drug-related side effects associated with standard doses of prednisone. Low dose prednisone or long-term maintenance therapy with azathioprine (2 mg/kg daily) is a treatment option.

Relapse after remission and drug withdrawal

Relapse occurs in 20%-86% of patients depending on the criteria for remission prior to drug withdrawal.¹³⁵⁻¹³⁷ Diminished stamina, arthralgias, and increase in the serum aspartate aminotransferase level to more than three-fold normal characterize this occurrence. Re-treatment with the original regimen typically induces another remission, but relapse recurs in 79% within 6 months after drug withdrawal.¹³⁵ With each re-treatment and relapse, the frequency of drug-related side effects increases, and this risk outweighs the low probability of a sustained remission with repeated conventional treatments.¹³⁸ Alternative therapies with either low dose prednisone or azathioprine are warranted after the second relapse.

The low dose prednisone regimen requires induction of clinical and laboratory remission on standard therapy and then reduction in the dose of prednisone by 2.5 mg each month of clinical and laboratory stability.¹³⁹ The lowest dose that prevents symptoms and keeps serum aspartate aminotransferase levels below three-fold normal is maintained. Eighty-seven percent of patients can be managed on prednisone, 10 mg daily or less (median dose, 7.5 mg daily). Side effects associated with earlier conventional treatments improve or disappear in 85%; new side effects do not develop; and survival is unaffected.

Maintenance therapy with azathioprine also requires induction of clinical and laboratory remission by conventional treatments.^{140,141} The corticosteroid component is then withdrawn, and the dose of azathioprine is increased to 2 mg per kg daily and maintained indefinitely. Eighty-seven percent of adult patients managed in this fashion remain in remission during a median observation interval of 67 months. Follow-up liver biopsy assessments disclose inactive or minimal histological disease in 94%; corticosteroid-related side effects improve or disappear in most patients; and the drug is generally well-tolerated. The most common side effects are withdrawal arthralgias (63%), lymphopenia (57%), and myelosuppression (7%). Neoplasms involving diverse cell types occur in 8%.

Relapse does not preclude permanent discontinuation of medication late in the course of the disease.^{128,138} Twenty-eight percent of patients who relapse and are re-treated develop inactive disease and can be withdrawn from medication. The probability of a sustained remission after initial or subsequent therapy is 47% during 10 years of follow-up. Conventional re-treatment schedules are able to induce a sustained remission more commonly than

long-term maintenance schedules (59% *versus* 12%, $p=0.00002$), but all management schedules should be withdrawn periodically to assess this outcome.

Liver transplantation

Liver transplantation is an effective treatment for the decompensated patient. Patient and graft survival after liver transplantation ranges from 83% to 92%, and the actuarial 10-year survival after transplantation is 75%.^{5,142} Recurrence is recognized in at least 17% of patients after 5±1 years, especially in individuals receiving inadequate immunosuppression.¹⁴³ Adjustments in the immunosuppressive regimen are usually able to suppress recurrent disease, but rarely cirrhosis or graft failure occurs.¹⁴⁴ Patients transplanted for autoimmune hepatitis may also have a greater frequency of acute and chronic rejection than patients transplanted for non-autoimmune conditions.^{145,146} These potential consequences have tempered efforts to rapidly withdraw corticosteroids after the procedure.

Autoimmune hepatitis can develop *de novo* in children and adult recipients who undergo transplantation for non-autoimmune liver disease.¹⁴⁷⁻¹⁵² Children seem to have a predilection for the syndrome; immunosuppression with cyclosporine is a common feature; and treatment with prednisone and azathioprine is typically effective. *De novo* autoimmune hepatitis is rare, occurring in 3%-5% of allografts, and it can result in graft loss if not treated with corticosteroids.¹⁵³ In children, *de novo* disease may reflect defective negative selection of autoreactive cells by the thymus, generation of promiscuous lymphocytes by excessive antigenic exposure, and/or impaired apoptosis of autoreactive cells by cyclosporine or tacrolimus. In adult patients, these same mechanisms are pertinent except for thymic dysfunction.^{154,155}

Emerging drug therapies

There is no shortage of new drugs that have been proposed for autoimmune hepatitis^{156,157} (Table VI). Many have emerged from the transplantation arena, but none has been rigorously evaluated or incorporated into a conventional management algorithm. Of the drugs that promise greater blanket immunosuppression than prednisone or azathioprine, cyclosporine and mycophenolate mofetil have shown the most promise. Controlled clinical trials are sorely needed to establish their efficacy.

Cyclosporine has been used successfully as salvage therapy in patients who have failed conventional corticosteroid treatment or been intolerant of the medication.¹⁵⁸⁻¹⁶⁵ It has also been used as first-line therapy in children and adults.¹⁶³⁻¹⁶⁵ The medication binds cyclophilin and inhibits the phosphatase activity of calcineurin. As a calcineurin inhibitor, it impairs transcription of IL-2, cell cycle progression, and expansion of cytotoxic T lymphocytes. Side effects include renal insufficiency, hypertension, and malignancy.

Mycophenolate mofetil is an ester prodrug hydrolyzed by liver esterases to produce the active metabolite, mycophenolic acid, which in turn acts as a non-competitive, reversible inhibitor of inosine monophosphate dehydrogenase.¹⁶⁶ Inosine monophosphate dehydrogenase is the rate-limiting enzyme for *de novo* synthesis of purines, and by inhibiting its action, mycophenolate mofetil selectively prevents the proliferative responses of T and B cells to mitogens or antigens. Mycophenolate mofetil is a purine antagonist like azathioprine, and it has a low frequency of side effects (mainly leukopenia) and independence from the thiopurine methyltransferase pathway of catabolism.

Two small uncontrolled clinical experiences have shown improvement in the laboratory indices of liver inflammation after the administration of mycophenolate mofetil, 1 gram twice daily, to patients unsuccessfully treated with azathioprine or intolerant of this drug.^{167,168} The prospect of replacing corticosteroids with mycophenolate mofetil has also stimulated its empiric use. Other preliminary experiences have failed to demonstrate a potent salvage effect or consistent corticosteroid-sparing action, and the proper role and target population for this drug remain uncertain.¹⁶⁹

Tacrolimus (4 mg twice daily),^{170,171} budesonide (3 mg thrice daily),¹⁷² 6-mercaptopurine (1.5 mg/kg daily),¹⁷³ deflazacort (7.5 mg for each 5 mg prednisone dose daily),¹⁷⁴ ursodeoxycholic acid (13-15 mg/kg daily),¹⁷⁵ methotrexate,¹⁷⁶ and cyclophosphamide¹⁷⁷ have all been used with anecdotal success in treating corticosteroid resistant or intolerant patients. Only ursodeoxycholic acid has been evaluated by randomized controlled clinical trial as a salvage therapy, and it is the one negative experience.¹⁷⁸ Budesonide is currently undergoing clinical trial as front-line therapy after earlier studies had demonstrated its limitations as a salvage treatment.¹⁷⁹

Emerging site-specific interventions

Site-specific interventions are designed to target key steps in the pathogenic pathway and thereby control the disease without inducing blanket immune suppression.¹⁵⁷ These therapies are at theoretical or preliminary stages of development, and they await full clarification of the molecular mechanisms of the disease and the availability of suitable animal models to assess their feasibility. The various intracellular signaling pathways affecting immunocyte activation and proliferation are prime targets for these strategies.

Competitive inhibition of autoantigen presentation has already been applied in rheumatoid arthritis, and it can be considered in autoimmune hepatitis after full characterization of its autoantigens.¹⁸⁰ Synthetic peptides that compete with the autoantigen for presentation by class II MHC molecules can be used to block the first co-stimulatory signal in immunocyte activation. *Soluble cytotoxic T lymphocyte antigen-4 (CTLA-4)* interferes with the sec-

ond co-stimulatory signal and dampens immunocyte activation.¹⁸¹ It has already been used to blunt immune reactivity in mismatched blood marrow recipients, and it is a powerful tool by which to manipulate the immune response.¹⁸² *Oral tolerance regimens* induce systemic non-responsiveness to an autoantigen by oral feedings, and this intervention has already been used in the treatment of multiple sclerosis, rheumatoid arthritis, insulin dependent diabetes mellitus, myasthenia gravis and thyroiditis.¹⁸³ Low dose regimens stimulate cytokine production and suppression of the immune response, whereas high dose regimens cause clonal deletion of immunocytes and anergy. The ingested antigen is delivered directly to the liver by the portal circulation, and the treatment may be especially effective in autoimmune hepatitis once the critical epitope has been defined.¹⁸⁴ *T cell vaccination* can deplete clones of activated cytotoxic T lymphocytes, and it is the only site-specific intervention that has been assessed in experimental murine autoimmune hepatitis.¹⁸⁵ Identification and precise targeting of the critical T cell clones remain challenges for this modality. *Cytokine manipulations* are feasible by administering drugs that disrupt intracellular signaling pathways involved in cytokine transcription or by using recombinant cytokines (such as IL-10) or monoclonal antibodies (such as anti-TNF) to alter the type of cytokine response. Similar interventions are already being evaluated in inflammatory bowel disease¹⁸⁶ and chronic hepatitis C.¹⁸⁷ *Gene therapy* also has promise in the treatment of a polygenic disorder such as autoimmune hepatitis if genes can be delivered that counterbalance the over-production of certain regulatory cytokines, limit fibrosis, or promote regeneration.¹⁸⁸

Variant syndromes

Codification of the clinical criteria for the diagnosis of autoimmune hepatitis has facilitated recognition of variant syndromes.¹⁸⁹⁻¹⁹² These syndromes include patients with autoimmune hepatitis and another type of chronic liver disease (overlap syndrome) or findings suggestive but non-diagnostic of autoimmune hepatitis (outlier syndrome). Overlap syndromes include patients with mixed features of autoimmune hepatitis and PBC or PSC, and outlier syndromes include patients with autoimmune cholangitis (or AMA-negative PBC) and cryptogenic chronic hepatitis. These variant conditions currently lack an established identity, official designation, and treatment strategy. Their occurrences, however, must be recognized, and they should not be assimilated into diagnoses that hide their individuality or imperil the homogeneity of the classical diseases.

The variant syndromes are important to recognize not only because they are common (occurring in 18% of patients with autoimmune liver disease), but they respond variably to corticosteroid treatment.^{193,194} The principal determinant of corticosteroid response is the degree of

cholestasis at presentation, and this is most easily assessed by determining the magnitude of the serum alkaline phosphatase level above the upper limit of normal. Patients with variant syndromes and serum alkaline phosphatase levels greater than twofold normal are unlikely to respond to this treatment.¹⁹³⁻¹⁹⁶

Management of the variant syndromes is empiric and based on the predominant manifestations of the disease. Patients with autoimmune hepatitis and features of PBC who have serum alkaline phosphatase levels less than twofold normal can be treated with corticosteroids.¹⁹³⁻¹⁹⁵ Patients with higher serum alkaline phosphatase levels and those with florid duct lesions on histological examination are candidates for treatment with corticosteroids and ursodeoxycholic acid.¹⁹⁶ Patients with autoimmune hepatitis and PSC lack an effective treatment, but they are candidates for a trial of therapy with prednisone and high dose ursodeoxycholic acid (15-20 mg/kg daily).¹⁹³ Patients with autoimmune cholangitis can be treated with prednisone, ursodeoxycholic acid, or both depending on the serum alkaline phosphatase level.^{193,197} Multicenter, collaborative studies are needed to codify diagnostic criteria and establish treatment algorithms.¹¹

Summary

Autoimmune hepatitis should be considered in all patients with acute or chronic hepatitis of undetermined cause, including patients with fulminant presentations and those who have been transplanted. Recurrent autoimmune hepatitis is possible after liver transplantation, and the disease can develop *de novo* in children and adults transplanted for non-autoimmune diseases. The histological spectrum of autoimmune hepatitis includes centrilobular or perivenular (Rappaport zone 3) necrosis, and background histological features of bile duct injury do not dissuade the diagnosis or alter therapy. Prednisone in combination with azathioprine is the preferred treatment in adults, and it may reduce fibrosis by suppressing inflammatory activity. New autoantibodies have promise as diagnostic and prognostic tools, and several are now available as commercial kits. Genetic factors influence disease expression and behavior, and they may be clues to region-specific etiologic agents. New treatments are evolving that promise better blanket immunosuppression and site-specific intervention. Variant syndromes are common, and they should be sought in all patients, especially in those who are refractory to corticosteroid therapy.

References

- Czaja AJ, Freese DK. Diagnosis and treatment of autoimmune hepatitis. *Hepatology* 2002; 36: 479-497.
- Czaja AJ, Carpenter HA. Sensitivity, specificity and predictability of biopsy interpretations in chronic hepatitis. *Gastroenterology* 1993; 105: 1824-1832.
- Boberg KM, Aadland E, Jahnsen J, Raknerud N, Stiris M, Bell H. Incidence and prevalence of primary biliary cirrhosis, primary sclerosing cholangitis, and autoimmune hepatitis in a Norwegian population. *Scand J Gastroenterol* 1998; 33: 99-103.
- Boberg KM. Prevalence and epidemiology of autoimmune hepatitis. *Clin Liver Dis* 2002; 6: 635-647.
- Seaberg EC, Belle SH, Beringer KC, Schivins JL, Detre KM. Liver transplantation in the United States from 1987-1998: updated results from the Pitt-UNOS liver transplant registry. In: Cecka JM, Terasaki PI, eds. *Clinical Transplants 1998*. Los Angeles: UCLA Tissue Typing Laboratories 1999: 17-37.
- Hurlburt KJ, McMahon BJ, Deubner H, Hsu-Trawinski B, Williams JL, Kowdley KV. Prevalence of autoimmune hepatitis in Alaska natives. *Am J Gastroenterol* 2002; 97: 2402-2407.
- Lim KN, Casanova RL, Boyer TD, Bruno CJ. Autoimmune hepatitis in African Americans: presenting features and responses to therapy. *Am J Gastroenterol* 2001; 96: 3390-3394.
- Nakamura K, Yoneda M, Yokohama S, Tamori K, Sato Y, Aso K, Aoshima M, Hasegawa T, Makino I. Efficacy of ursodeoxycholic acid in Japanese patients with type 1 autoimmune hepatitis. *J Gastroenterol Hepatol* 1998; 13: 490-495.
- Czaja AJ, Souto EO, Bittencourt PL, Cancado ELR, Porto G, Goldberg AC, Donaldson PT. Clinical distinctions and pathogenic implications of type 1 autoimmune hepatitis in Brazil and the United States. *J Hepatol* 2002; 37: 302-308.
- Zolfino T, Heneghan MA, Norris S, Harrison PM, Portmann BC, McFarlane IG. Characteristics of autoimmune hepatitis in patients who are not of European/Caucasoid ethnic origin. *Gut* 2002; 50: 713-717.
- Czaja AJ, Bianchi FB, Carpenter HA, Krawitt EL, Lohse AW, Manns MP, McFarlane IG, Mieli-Vergani G, Toda G, Vergani D, Vierling J, Zeniya M. Treatment challenges and investigational opportunities in autoimmune hepatitis. *Hepatology* 2005; 41: 207-215.
- Alvarez F, Berg PA, Bianchi FB, Bianchi L, Burroughs AK, Cancado EL, Chapman RW, Cooksley WGE, Czaja AJ, Desmet VJ, Donaldson PT, Eddleston ALWF, Fainboim L, Heathcote J, Homberg J-C, Hoofnagle JH, Kakumu S, Krawitt EL, Mackay IR, MacSween RNM, Maddrey WC, Manns MP, McFarlane IG, Meyer zum Buschenfelde K-H, Mieli-Vergani G, Nakanuma Y, Nishioka M, Penner E, Porta G, Portmann BC, Reed WD, Rodes J, Schalm SW, Scheuer PJ, Schrupf E, Seki T, Toda G, Tsuji T, Tygstrup N, Vergani D, Zeniya M. International Autoimmune Hepatitis Group report: review of criteria for diagnosis of autoimmune hepatitis. *J Hepatol* 1999; 31: 929-938.
- Amontree JS, Stuart TD, Bredfeldt JE. Autoimmune chronic active hepatitis masquerading as acute hepatitis. *J Clin Gastroenterol* 1989; 11: 303-307.
- Porta G, Da Costa Gayotto LC, Alvarez F. Anti-liver-kidney microsome antibody-positive autoimmune hepatitis presenting as fulminant liver failure. *J Pediatric Gastroenterol Nutr* 1990; 11: 138-140.
- Nikias GA, Batts KP, Czaja AJ. The nature and prognostic implications of autoimmune hepatitis with an acute presentation. *J Hepatol* 1994; 21: 866-871.
- Kessler WR, Cummings OW, Eckert G, Chalasani N, Lumeng L, Kwo PY. Fulminant hepatic liver failure as the initial presentation of acute autoimmune hepatitis. *Clin Gastroenterol Hepatol* 2004; 2: 625-631.
- Burgart LJ, Batts KP, Ludwig J, Czaja AJ. Recent onset autoimmune hepatitis: biopsy findings and clinical correlations. *Am J Surg Pathol* 1995; 19: 699-708.
- Singh R, Nair S, Farr G, Mason A, Perrillo R. Acute autoimmune hepatitis presenting with centrilobular liver disease: case report and review of the literature. *Am J Gastroenterol* 2002; 97: 2670-2673.
- Okano N, Yamamoto K, Sakaguchi K, Miyake Y, Shimada N, Hakoda T, Terada R, Baba S, Suzuki T, Tsuji T. Clinicopathological features of acute-onset autoimmune hepatitis. *Hepatol Res* 2003; 25: 263-270.
- Davis GL, Czaja AJ, Baggenstoss AH, Taswell HF. Prognostic and therapeutic implications of extreme serum aminotransferase elevation in chronic active hepatitis. *Mayo Clin Proc* 1982; 57: 303-309.
- Gough A, Chapman S, Wagstaff K, Emery P, Elias E. Minocycline induced autoimmune hepatitis and systemic lupus erythematosus-like syndrome. *BMJ* 1996; 312: 169-172.

22. Johnson PJ, McFarlane IG, Alvarez F, Bianchi FB, Bianchi L, Burroughs A, Chapman RW, Czaja AJ, Desmet V, Edleston ALWF, Gerber MA, Hoofnagle JH, Kakumu S, MacSween RNM, Maddrey WC, Manns MP, Meyer zum Buschenfelde K-H, Mieli-Vergani G, Portmann BC, Reed WD, Schalm SW, Scheuer PJ, Toda G, Tsuji T, Tygstrup N, Vergani D, Zeniya M. Meeting Report. International Autoimmune Hepatitis Group. *Hepatology* 1993; 18: 998-1005.
23. Czaja AJ, Carpenter HA. Validation of a scoring system for the diagnosis of autoimmune hepatitis. *Dig Dis Sci* 1996; 41: 305-314.
24. Czaja AJ, Manns MP. The validity and importance of subtypes of autoimmune hepatitis: a point of view. *Am J Gastroenterol* 1995; 90: 1206-1211.
25. McFarlane IG. The relationship between autoimmune markers and different clinical syndromes in autoimmune hepatitis. *Gut* 1998; 42: 599-602.
26. Wang KK, Czaja AJ. Prognosis of corticosteroid-treated hepatitis B surface antigen-negative chronic active hepatitis in postmenopausal women: a retrospective analysis. *Gastroenterology* 1989; 97: 1288-1293.
27. Schramm C, Kanzler S, Meyer zum Buschenfelde K-H, Galle PR, Lohse AW. Autoimmune hepatitis in the elderly. *Am J Gastroenterol* 2001; 96: 1587-1591.
28. Czaja AJ, Davis GL, Ludwig J, Baggenstoss AH, Taswell HF. Autoimmune features as determinants of prognosis in steroid-treated chronic active hepatitis of uncertain etiology. *Gastroenterology* 1983; 85: 713-717.
29. Perdigoto R, Carpenter HA, Czaja AJ. Frequency and significance of chronic ulcerative colitis in severe corticosteroid-treated autoimmune hepatitis. *J Hepatol* 1992; 14: 325-331.
30. Kogan J, Safadi R, Ashur Y, Shouval D, Ilan Y. Prognosis of symptomatic versus asymptomatic autoimmune hepatitis. A study of 68 patients. *J Clin Gastroenterol* 2002; 35: 75-81.
31. Homberg J-C, Abuaf N, Bernard O, Islam S, Alvarez F, Khalil SH, Poupon R, Darnis F, Levy V-G, Gripon P, Opolon P, Bernuau J, Benhamou J-P, Alagille D. Chronic active hepatitis associated with antiliver/kidney microsome antibody type 1: a second type of "autoimmune" hepatitis. *Hepatology* 1987; 7: 1333-1339.
32. Gueguen M, Meunier-Rotival M, Bernard O, Alvarez F. Anti-liver kidney microsome antibody recognizes a cytochrome P450 from the IID subfamily. *J Exp Med* 1988; 168: 801-806.
33. Zanger UM, Hauri H-P, Loeper J, Homberg J-C, Meyer UA. Antibodies against human cytochrome P-450db1 in autoimmune hepatitis type II. *Proc Natl Acad Sci USA* 1988; 85: 8256-8260.
34. Manns MP, Griffin KJ, Sullivan KF, Johnson EF. LKM-1 autoantibodies recognize a short linear sequence in P450IID6, a cytochrome P-450 monooxygenase. *J Clin Invest* 1991; 88: 1370-1378.
35. Kerkar N, Choudhuri K, Ma Y, Mahmoud A, Bogdanos DP, Muratori L, Bianchi F, Williams R, Mieli-Vergani G, Vergani D. Cytochrome P450D6₁₉₃₋₂₁₂: a new immunodominant epitope and target of virus/self cross-reactivity in liver kidney microsomal autoantibody type I-positive liver disease. *J Immunol* 2003; 170: 1481-1489.
36. Yamamoto AM, Cresteil D, Homberg JC, Alvarez F. Characterization of the anti-liver-kidney microsome antibody (anti-LKM1) from hepatitis C virus-positive and -negative sera. *Gastroenterology* 1993; 104: 1762-1767.
37. Klein R, Zanger UM, Berg T, Hopf U, Berg PA. Overlapping but distinct specificities of anti-liver-kidney microsome antibodies in autoimmune hepatitis type II and hepatitis C revealed by recombinant native CYP2D6 and novel peptide epitopes. *Clin Exp Immunol* 1999; 118: 290-297.
38. Czaja AJ, Manns MP, Homburger HA. Frequency and significance of antibodies to liver/kidney microsome type 1 in adults with chronic active hepatitis. *Gastroenterology* 1992; 103: 1290-1295.
39. Reddy KR, Krawitt EL, Homberg JC, Jeffers LJ, de Medina M, Chastenay B, Poupon R, Opolon P, Beaugrand M, Abuaf N, Johanet C, Gergeois J, Schiff ER. Absence of anti-LKM1 in hepatitis C viral infection in the United States. *J Viral Hepat* 1995; 2: 175-179.
40. Durazzo M, Philipp T, van Pelt FNAM, Luttig B, Borghesio E, Michel G, Schmidt E, Loges S, Rizzetto M, Manns MP. Heterogeneity of microsomal autoantibodies (LKM) in chronic hepatitis C and D virus infection. *Gastroenterology* 1995; 108: 455-462.
41. Gerotto M, Pontisso P, Giostra F, Francesconi R, Muratori L, Ballardini G, Lenzi M, Tsiminetzky S, Bianchi FB, Baralle FB, Alberti A. Analysis of the hepatitis C virus genome in patients with anti-LKM-1 autoantibodies. *J Hepatol* 1994; 21: 273-276.
42. Clemente MG, Meloni A, Obermayer-Staub P, Frau F, Manns MP, De Virgiliis S. Two cytochromes P450 are major hepatocellular autoantigens in autoimmune polyglandular syndrome type 1. *Gastroenterology* 1998; 114: 324-328.
43. Aaltonen J, Borses P, Sandkuijl L, Perheentupa J, Peltonen L. An autosomal locus causing autoimmune disease: autoimmune polyglandular disease type 1 assigned to chromosome 21. *Nature Genetics* 1994; 8: 83-87.
44. Manns M, Gerken G, Kyriatsoulis A, Staritz M, Meyer zum Buschenfelde KH. Characterization of a new subgroup of autoimmune chronic active hepatitis by autoantibodies against a soluble liver antigen. *Lancet* 1987; 1: 292-294.
45. Stechemesser E, Klein R, Berg PA. Characterization and clinical relevance of liver-pancreas antibodies in autoimmune hepatitis. *Hepatology* 1993; 18: 1-9.
46. Wies I, Brunner S, Henninger J, Herkel J, Meyer zum Buschenfelde KH, Lohse AW. Identification of target antigen for SLA/LP autoantibodies in autoimmune hepatitis. *Lancet* 2000; 355: 1510-1515.
47. Costa M, Rodrigues-Sanchez JL, Czaja AJ, Gelpi C. Isolation and characterization of cDNA encoding the antigenic protein of the human tRNA^{Ser}^{Ser} complex recognized by autoantibodies from patients with type 1 autoimmune hepatitis. *Clin Exp Immunol* 2000; 121: 364-374.
48. Czaja AJ, Carpenter HA, Manns MP. Antibodies to soluble liver antigen, P450IID6, and mitochondrial complexes in chronic hepatitis. *Gastroenterology* 1993; 105: 1522-1528.
49. Kanzler S, Weidemann C, Gerken G, Iohr HF, Galle PR, Meyer zum Buschenfelde KH, Lohse AW. Clinical significance of autoantibodies to soluble liver antigen in autoimmune hepatitis. *J Hepatol* 1999; 31: 635-640.
50. Czaja AJ. Understanding the pathogenesis of autoimmune hepatitis. *Am J Gastroenterol* 2001; 96: 1224-1231.
51. Vergani D, Choudhuri K, Bogdanos DP, Mieli-Vergani G. Pathogenesis of autoimmune hepatitis. *Clin Liver Dis* 2002; 6: 727-737.
52. Czaja AJ, Sievers C, Zein NN. Nature and behavior of serum cytokines in type 1 autoimmune hepatitis. *Dig Dis Sci* 2000; 45: 1028-1035.
53. Lapierre P, Djilali-Saiah I, Vitozzi S, Alvarez F. A murine model of type 2 autoimmune hepatitis: xenoinmunization with human antigens. *Hepatology* 2004; 39: 1066-1074.
54. Donaldson PT, Doherty DG, Hayllar KM, McFarlane IG, Johnson PJ, Williams R. Susceptibility to autoimmune chronic active hepatitis: human leukocyte antigens DR4 and A1-B8-DR3 are independent risk factors. *Hepatology* 1991; 13: 701-706.
55. Seki T, Ota M, Furuta S, Fukushima H, Kondo T, Hino K, Mizuki N, Ando A, Tsuji K, Inoko H, Kiyosawa K. HLA class II molecules and autoimmune hepatitis susceptibility in Japanese patients. *Gastroenterology* 1992; 103: 1041-1047.
56. Qiu D-K, Ma X. Relationship between human leukocyte antigen-DRB1 and autoimmune hepatitis type I in Chinese patients. *J Gastroenterol Hepatol* 2003; 18: 63-67.
57. Muratori P, Czaja AJ, Muratori L, Pappas G, Maccariello S, Cassani F, Granito A, Ferrari R, Mantovani V, Lenzi M, Bianchi FB. Genetic predispositions to autoimmune hepatitis in Italian and North American patients. *World J Gastroenterol* (in press).
58. Fainboim L, Marcos Y, Pando M, Capucchio M, Reyes GB, Galoppo C, Badia I, Remondino G, Ciocca M, Ramonet M, Fainboim H, Satz ML. Chronic active autoimmune hepatitis in children. Strong association with a particular HLA DR6 (DRB1*1301) haplotype. *Hum Immunol* 1994; 41: 146-150.
59. Pando M, Larriba J, Fernandez GC, Fainboim H, Ciocca M, Ramonet M, Badia I, Daruich J, Findor J, Tanno H, Canero-Velasco C, Fainboim L. Pediatric and adult forms of type 1 autoimmune hepatitis in Argentina: evidence for differential genetic predisposition. *Hepatology* 1999; 30: 1374-1380.
60. Goldberg AC, Bittencourt PL, Mouglin B, Cancado ELR, Porta G, Carrilho F, Kalil J. Analysis of HLA haplotypes in autoimmune hepa-

- titis type 1: identifying the major susceptibility locus. *Hum Immunol* 2001; 62: 165-169.
61. Czaja AJ, Carpenter HA, Santrach PJ, Moore SB. Significance of HLA DR4 in type 1 autoimmune hepatitis. *Gastroenterology* 1993; 105: 1502-1507.
 62. Czaja AJ, Donaldson PT. Gender effects and synergisms with histocompatibility leukocyte antigens in type 1 autoimmune hepatitis. *Am J Gastroenterol* 2002; 97: 2051-2057.
 63. Czaja AJ, Kruger M, Santrach PJ, Moore SB, Manns MP. Genetic distinctions between types 1 and 2 autoimmune hepatitis. *Am J Gastroenterol* 1997; 92: 2197-2200.
 64. Bittencourt PL, Goldberg AC, Cancado ELR, Porta G, Carrilho FJ, Farias AQ, Palacios SA, Chiarella JM, Abrantes-Lemos CP, Baggio VL, Laudanna AA, Kalil J. Genetic heterogeneity in susceptibility to autoimmune hepatitis types 1 and 2. *Am J Gastroenterol* 1999; 94: 1906-1913.
 65. Manns MP, Kruger M. Immunogenetics of chronic liver diseases. *Gastroenterology* 1994; 106: 1676-1697.
 66. Muratori P, Czaja AJ, Muratori L, Granito A, Pappas G, Cassani F, Mantovani W, Lenzi M, Bianchi FB. Evidence for a genetic basis in the different geographical occurrence of liver/kidney microsomal antibody type 1 in hepatitis C. *J Hepatol* (submitted).
 67. Strettell MDJ, Donaldson PT, Thomson LJ, Santrach PJ, Moore SB, Czaja AJ, Williams R. Allelic basis for HLA-encoded susceptibility to type 1 autoimmune hepatitis. *Gastroenterology* 1997; 112: 2028-2035.
 68. Czaja AJ, Strettell MDJ, Thomson LJ, Santrach PJ, Moore SB, Donaldson PT, Williams R. Associations between alleles of the major histocompatibility complex and type 1 autoimmune hepatitis. *Hepatology* 1997; 25: 317-323.
 69. Czaja AJ, Donaldson PT. Genetic susceptibilities for immune expression and liver cell injury in autoimmune hepatitis. *Immunol Rev* 2000; 174: 250-259.
 70. Czaja AJ, Doherty DG, Donaldson PT. Genetic bases of autoimmune hepatitis. *Dig Dis Sci* 2002; 47: 2139-2150.
 71. Doherty DG, Donaldson PT, Underhill JA, Farrant JM, Duthie A, Mieli-Vergani G, McFarlane IG, Johnson PJ, Eddleston ALWF, Mowat AP, Williams R. Allelic sequence variation in the HLA class II genes and proteins in patients with autoimmune hepatitis. *Hepatology* 1994; 19: 609-615.
 72. Vazquez-Garcia MN, Alaez C, Olivo A, Debaz H, Perez-Luque E, Burguete A, Cano S, de la Rosa G, Bautista N, Hernandez A, Bandera J, Torres LF, Kersenobich D, Alvarez F, Gorodezky C. MHC class II sequences of susceptibility and protection in Mexicans with autoimmune hepatitis. *J Hepatol* 1998; 28: 985-990.
 73. Donaldson PT, Czaja AJ. Genetic effects on susceptibility, clinical expression, and treatment outcome of type 1 autoimmune hepatitis. *Clin Liver Dis* 2002; 6: 707-725.
 74. Fainboim L, Velasco VCC, Marcos CY, Ciocca M, Roy A, Theiler G, Capucchio M, Nuncifora S, Sala L, Zelazko M. Protracted, but not acute, hepatitis A virus infection is strongly associated with HLA-DRB1*1301, a marker for pediatric autoimmune hepatitis. *Hepatology* 2001; 33: 1512-1517.
 75. Vento S, Garofano T, Di Perri G, Dolci L, Concia E, Bassetti D. Identification of hepatitis A virus as a trigger for autoimmune chronic hepatitis type 1 in susceptible individuals. *Lancet* 1991; 337: 1183-1187.
 76. Cookson S, Constantini PK, Clare M, Underhill JA, Bernal W, Czaja AJ, Donaldson PT. Frequency and nature of cytokine gene polymorphisms in type 1 autoimmune hepatitis. *Hepatology* 1999; 30: 851-856.
 77. Czaja AJ, Cookson S, Constantini PK, Clare M, Underhill JA, Donaldson PT. Cytokine polymorphisms associated with clinical features and treatment outcome in type 1 autoimmune hepatitis. *Gastroenterology* 1999; 117: 645-652.
 78. Agarwal K, Czaja AJ, Jones DEJ, Donaldson PT. CTLA-4 gene polymorphism and susceptibility to type 1 autoimmune hepatitis. *Hepatology* 2000; 31: 49-53.
 79. Djilali-Saiah I, Ouellette P, Caillat-Zucman S, Debray D, Kohn JI, Alvarez F. CTLA-4/CD28 region polymorphisms in children from families with autoimmune hepatitis. *Human Immunol* 2001; 62: 1356-1362.
 80. Bittencourt PL, Palacios SA, Cancado ELR, Porta G, Carrilho FJ, Laudanna AA, Kalil J, Goldberg AC. Cytotoxic T lymphocyte antigen-4 gene polymorphisms do not confer susceptibility to autoimmune hepatitis types 1 and 2 in Brazil. *Am J Gastroenterol* 2003; 98: 1616-1620.
 81. Vogel A, Strassburg CP, Manns MP. Genetic association of vitamin D receptor polymorphisms with primary biliary cirrhosis and autoimmune hepatitis. *Hepatology* 2002; 35: 126-131.
 82. Vogel A, Strassburg CP, Manns MP. 77 C/G mutation in the tyrosine phosphatase CD45 gene and autoimmune hepatitis: evidence of a genetic link. *Genes Immun* 2003; 4: 79-81.
 83. Seki S, Kitada T, Iwai S, Kadoya H, Yamada T, Kawada N, Sakaguchi H, Wakasa K. Immunohistochemical detection of Fas and apoptosis in type-1 autoimmune hepatitis. *Hepatogastroenterology* 2003; 50: 1274-1277.
 84. McKiernan S, Norris S, Vaughan R, Czaja AJ, Donaldson PT. MICA polymorphisms and susceptibility to autoimmune hepatitis: further evidence that susceptibility maps to the HLA class II region on chromosome 6p21.3 (abstract). *Hepatology* 2002; 36: 503A.
 85. Schramm C, Protschka M, Kohler HH, Podlech J, Reddehase MJ, Schirmacher P, Galle PR, Lohse AW, Blessing M. Impairment of TGF- β signaling in T cells increases susceptibility to experimental autoimmune hepatitis in mice. *Am J Physiol Gastrointest Liver Physiol* 2003; 284: G525-G535.
 86. McFarlane IG, Heneghan MA. Autoimmunity and the female liver. *Hepatol Res* 2004; 28: 171-176.
 87. Whitacre CC, Reingold SC, O'Looney PA. A gender gap in autoimmunity. *Science* 1999; 283: 1277-1278.
 88. Gregorio GV, Portmann B, Reid F, Donaldson PT, Doherty DG, McCartney M, Mowat AP, Vergani D, Mieli-Vergani G. Autoimmune hepatitis in childhood. A 20 year survey. *Hepatology* 1997; 25: 541-547.
 89. Buchel E, Van Steenberg W, Nevens F, Fevery J. Improvement of autoimmune hepatitis during pregnancy followed by flare-up after delivery. *Am J Gastroenterol* 2002; 97: 3160-3165.
 90. Czaja AJ. Autoantibodies. *Bailliere's Clin Gastroenterol* 1995; 9: 723-744.
 91. Czaja AJ, Homburger HA. Autoantibodies in liver disease. *Gastroenterology* 2001; 120: 239-249.
 92. Czaja AJ, Norman GL. Autoantibodies in the diagnosis and management of liver disease. *J Clin Gastroenterol* 2003; 37: 315-329.
 93. Czaja AJ, Pfeifer KD, Decker RH, Vallari AS. Frequency and significance of antibodies to asialoglycoprotein receptor in type 1 autoimmune hepatitis. *Dig Dis Sci* 1996; 41: 1733-1740.
 94. Czaja AJ, Cassani F, Cataleta M, Valentini P, Bianchi FB. Frequency and significance of antibodies to actin in type 1 autoimmune hepatitis. *Hepatology* 1996; 24: 1068-1073.
 95. Czaja AJ, Shums Z, Binder WL, Lewis SJ, Nelson VJ, Norman GL. Frequency and significance of antibodies to chromatin in autoimmune hepatitis. *Dig Dis Sci* 2003; 48: 1658-1664.
 96. Czaja AJ, Shums Z, Norman GL. Nonstandard antibodies as prognostic markers in autoimmune hepatitis. *Autoimmunity* 2004; 37: 195-201.
 97. Czaja AJ, Morshed SA, Parveen S, Nishioka M. Antibodies to single-stranded and double-stranded DNA in antinuclear antibody-positive type 1-autoimmune hepatitis. *Hepatology* 1997; 26: 567-572.
 98. Czaja AJ, Ming C, Shirai M, Nishioka M. Frequency and significance of antibodies to histones in autoimmune hepatitis. *J Hepatol* 1995; 23: 32-38.
 99. Muratori P, Muratori L, Guidi M, Maccariello S, Pappas G, Ferrari R, Gionchetti P, Campieri M, Bianchi FB. Anti-*Saccharomyces cerevisiae* antibodies (ASCA) and autoimmune liver diseases. *Clin Exp Immunol* 2003; 132: 473-476.
 100. Czaja AJ, Shums Z, Donaldson PT, Norman GL. Frequency and significance of antibodies to *Saccharomyces cerevisiae* in autoimmune hepatitis. *Dig Dis Sci* 2004; 49: 611-618.
 101. Ohana M, Okazaki K, Hajiro K, Uchida K. Antilactoferrin antibodies in autoimmune liver diseases. *Am J Gastroenterol* 1998; 93: 1334-1339.
 102. Baeres M, Herkel J, Czaja AJ, Wies I, Kanzler S, Cancado ELR, Porta G, Nishioka M, Simon T, Daehrich C, Schlumberger W, Galle PR, Lohse AW. Establishment of standardized SLA/LP immunoassays: specificity for autoimmune hepatitis, worldwide occurrence, and clinical characteristics. *Gut* 2002; 51: 259-264.

103. Czaja AJ, Donaldson PT, Lohse AW. Antibodies to soluble liver antigen/liver pancreas and HLA risk factors in type 1 autoimmune hepatitis. *Am J Gastroenterol* 2002; 97: 413-419.
104. Czaja AJ, Shums Z, Norman GL. Frequency and significance of antibodies to soluble liver antigen/liver pancreas in variant autoimmune hepatitis. *Autoimmunity* 2002; 35: 475-483.
105. Ma Y, Okamoto M, Thomas MG, Bogdanos DP, Lopes AR, Portmann B, Underhill J, Durr R, Mieli-Vergani G, Vergani D. Antibodies to conformational epitopes of soluble liver antigen define a severe form of autoimmune liver disease. *Hepatology* 2002; 35: 658-664.
106. Torres-Collado AX, Czaja AJ, Gelpi C. Anti-tRNP^{ser}/SLA/LP autoantibodies: comparative study using in-house ELISA with recombinant 48.8 kDa protein, immunoblot, and analysis of immunoprecipitated RNAs. *Liver Int* (in press).
107. Targan SR, Landers C, Vidrich A, Czaja AJ. High-titer antineutrophil cytoplasmic antibodies in type 1 autoimmune hepatitis. *Gastroenterology* 1995; 108: 1159-1166.
108. Zauli D, Ghetti S, Grassi A, Descovich C, Cassani F, Ballardini G, Muratori L, Bianchi FB. Anti-neutrophil cytoplasmic antibodies in type 1 and type 2 autoimmune hepatitis. *Hepatology* 1997; 25: 1105-1107.
109. Biagi F, Pezzimenti D, Campanella J, Vadacca GB, Corazzo GR. Endomysial and tissue transglutaminase antibodies in coeliac sera: a comparison not influenced by previous serological testing. *Scand J Gastroenterol* 2001; 35: 955-958.
110. Gomez JC, Selvaggio G, Pizarro B, Viola MJ, Motta GL, Smecuol E, Castelletto R, Echeverria R, Vazquez H, Mazure R, Crivelli A, Sugai E, Maurino E, Bai JC. Value of a screening algorithm for celiac disease using tissue transglutaminase antibodies as first level in a population-based study. *Am J Gastroenterol* 2002; 97: 2785-2790.
111. Leon F, Camarero C, Pena R, Eiras P, Sanchez L, Baragano M, Lombardia M, Bootello A, Roy G. Anti-transglutaminase IgA ELISA: clinical potential and drawbacks in celiac disease diagnosis. *Scand J Gastroenterol* 2001; 36: 849-853.
112. Peracchi M, Trovato C, Longhi M, Gasparin M, Conte D, Tarantino C, Prati D, Bardella MT. Tissue transglutaminase antibodies in patients with end-stage heart failure. *Am J Gastroenterol* 2002; 97: 2850-2854.
113. Vecchi M, Folli C, Donato MF, Formenti S, Arosio E, De Franchis R. High rate of positive anti-tissue transglutaminase antibodies in chronic liver disease. Role of liver decompensation and of the antigen source. *Scand J Gastroenterol* 2003; 38: 50-54.
114. Volta U, De Franceschi L, Molinaro N, Cassani F, Muratori L, Lenzi M, Bianchi FB, Czaja AJ. Frequency and significance of anti-gliadin and anti-endomysial antibodies in autoimmune hepatitis. *Dig Dis Sci* 1998; 43: 2190-2195.
115. Abdo A, Meddings J, Swain M. Liver abnormalities in celiac disease. *Clin Gastroenterol Hepatol* 2004; 2: 107-112.
116. Bardella MT, Fraquelli M, Quatrini M, Molteni N, Bianchi P, Conti D. Prevalence of hypertransaminasemia in adult celiac patients and effect of gluten-free diet. *Hepatology* 1995; 22: 833-836.
117. Kaukinen K, Halme L, Collin P, Farkkila M, Maki M, Vehmanen P, Partanen J, Hockerstedt K. Celiac disease in patients with severe liver disease: gluten-free diet may reverse hepatic failure. *Gastroenterology* 2002; 122: 881-888.
118. Volta U, Rodrigo L, Granito A, Petrolinin N, Muratori P, Muratori L, Linares A, Veronesi L, Fuentes D, Zauli D, Bianchi FB. Celiac disease in autoimmune cholestatic liver disorders. *Am J Gastroenterol* 2002; 97: 2609-2613.
119. Sedlack RE, Smyrk TC, Czaja AJ, Talwalkar JA. Celiac-disease-associated autoimmune cholangitis. *Am J Gastroenterol* 2002; 97: 3196-3198.
120. McFarlane IG, Hegarty JE, McSorley CG, McFarlane BM, Williams R. Antibodies to liver-specific protein predict outcome of treatment withdrawal in autoimmune chronic active hepatitis. *Lancet* 1984; 2: 954-956.
121. Summerskill WHJ, Korman MG, Ammon HV, Baggenstoss AH. Prednisone for chronic active liver disease: dose titration, standard dose, and combination with azathioprine compared. *Gut* 1975; 16: 876-883.
122. Czaja AJ. Treatment strategies in autoimmune hepatitis. *Clin Liver Dis* 2002; 6: 799-824.
123. Czaja AJ. Treatment of autoimmune hepatitis. *Semin Liver Dis* 2002; 22: 365-377.
124. Czaja AJ, Wolf AM, Baggenstoss AH. Laboratory assessment of severe chronic active liver disease (CALD): correlation of serum transaminase and gamma globulin levels with histologic features. *Gastroenterology* 1981; 80: 687-692.
125. Czaja AJ, Carpenter HA. Histological features associated with relapse after corticosteroid withdrawal in type 1 autoimmune hepatitis. *Liver International* 2003; 23: 116-123.
126. Roberts SK, Therneau T, Czaja AJ. Prognosis of histological cirrhosis in type 1 autoimmune hepatitis. *Gastroenterology* 1996; 110: 848-857.
127. Davis GL, Czaja AJ, Ludwig J. Development and prognosis of histologic cirrhosis in corticosteroid-treated HBsAg-negative chronic active hepatitis. *Gastroenterology* 1984; 87: 1222-1227.
128. Czaja AJ, Menon KVN, Carpenter HA. Sustained remission after corticosteroid therapy for type 1 autoimmune hepatitis: a retrospective analysis. *Hepatology* 2002; 35: 890-897.
129. Cotler SJ, Jakate S, Jensen DM. Resolution of cirrhosis in autoimmune hepatitis with corticosteroid therapy. *J Clin Gastroenterol* 2001; 32: 428-430.
130. Schvarcz R, Glaumann H, Weiland O. Survival and histological resolution of fibrosis in patients with autoimmune chronic active hepatitis. *J Hepatol* 1993; 18: 15-23.
131. Dufour J-F, DeLellis R, Kaplan MM. Reversibility of hepatic fibrosis in autoimmune hepatitis. *Ann Intern Med* 1997; 127: 981-985.
132. Czaja AJ, Carpenter HA. Decreased fibrosis during corticosteroid therapy of autoimmune hepatitis. *J Hepatol* 2004; 40: 644-650.
133. Czaja AJ, Carpenter HA. Progressive fibrosis during corticosteroid therapy of autoimmune hepatitis. *Hepatology* 2004; 39: 1631-1638.
134. Czaja AJ, Davis GL, Ludwig J, Taswell HF. Complete resolution of inflammatory activity following corticosteroid treatment of HBsAg-negative chronic active hepatitis. *Hepatology* 1984; 4: 622-627.
135. Czaja AJ, Ammon HV, Summerskill WHJ. Clinical features and prognosis of severe chronic active liver disease (CALD) after corticosteroid-induced remission. *Gastroenterology* 1980; 78: 518-523.
136. Czaja AJ, Ludwig J, Baggenstoss AH, Wolf A. Corticosteroid-treated chronic active hepatitis in remission. Uncertain prognosis of chronic persistent hepatitis. *N Engl J Med* 1981; 304: 5-9.
137. Hegarty JE, Nouri-Aria KT, Portmann B, Eddleston ALWF, Williams R. Relapse following treatment withdrawal in patients with autoimmune chronic active hepatitis. *Hepatology* 1983; 3: 685-689.
138. Czaja AJ, Beaver SJ, Shiels MT. Sustained remission following corticosteroid therapy of severe HBsAg-negative chronic active hepatitis. *Gastroenterology* 1987; 92: 215-219.
139. Czaja AJ. Low dose corticosteroid therapy after multiple relapses of severe HBsAg-negative chronic active hepatitis. *Hepatology* 1990; 11: 1044-1049.
140. Stellon AJ, Keating JJ, Johnson PJ, MacFarlane IG, Williams R. Maintenance of remission in autoimmune chronic active hepatitis with azathioprine after corticosteroid withdrawal. *Hepatology* 1988; 8: 781-784.
141. Johnson PJ, McFarlane IG, Williams R. Azathioprine for long-term maintenance of remission in autoimmune hepatitis. *N Engl J Med* 1995; 333: 958-963.
142. Sanchez-Urdazpal L, Czaja AJ, van Hoek B, Krom RAF, Wiesner RH. Prognostic features and role of liver transplantation in severe corticosteroid-treated autoimmune chronic active hepatitis. *Hepatology* 1992; 15: 215-221.
143. González-Koch A, Czaja AJ, Carpenter HA, Roberts SK, Charlton MR, Porayko MK, Rosen CB, Wiesner RH. Recurrent autoimmune hepatitis after orthotopic liver transplantation. *Liver Transplantation* 2001; 4: 302-310.
144. Ratziu V, Samuel D, Sebah M, Farges O, Saliba F, Ichai P, Farahmand H, Gigou M, Feray C, Reynes M, Bismuth H. Long-term follow-up after liver transplantation for autoimmune hepatitis: evidence of recurrence of primary disease. *J Hepatol* 1999; 30: 131-141.

145. Hayashi M, Keefe EB, Krams SM, Martinez OM, Ojogho ON, So SKS, Garcia G, Imperial JC, Esquivel CO. Allograft rejection after liver transplantation for autoimmune liver disease. *Liver Transplant Surg* 1998; 4: 208-214.
146. Neuberger J. Transplantation for autoimmune hepatitis. *Semin Liver Dis* 2002; 22: 379-385.
147. Kerkar N, Hadzic N, Davies ET, Portmann B, Donaldson PT, Rela M, Heaton ND, Vergani D, Mieli-Vergani G. De-novo autoimmune hepatitis after liver transplantation. *Lancet* 1998; 353: 409-413.
148. Spada M, Bertani A, Sonzogni A, Petz W, Riva S, Torre G, Melzi ML, Alberti D, Colledan M, Segalin A, Lucianetti A, Gridelli B. A cause of late graft dysfunction after liver transplantation in children: de-novo autoimmune hepatitis. *Transplantation Proc* 2001; 33: 1747-1748.
149. Tan CK, Ho JMS. Concurrent de novo autoimmune hepatitis and recurrence of primary biliary cirrhosis post-liver transplantation. *Liver Transplantation* 2001; 7: 461-465.
150. Gupta P, Hart J, Millis JM, Cronin D, Brady L. De novo hepatitis with autoimmune antibodies and atypical histology. A rare cause of late graft dysfunction after pediatric liver transplantation. *Transplantation* 2001; 71: 664-668.
151. Aguilera I, Wichmann I, Sousa JM, Bernardos A, Franco E, Garcia-Lozano R, Nunez-Roldan A. Antibodies against glutathione S-transferase T1 (GSTT1) in patients with *de novo* immune hepatitis following liver transplantation. *Clin Exp Immunol* 2001; 126: 535-539.
152. Heneghan MA, Portmann BC, Norris SM, Williams R, Muiesan P, Rela M, Heaton ND, O'Grady JG. Graft dysfunction mimicking autoimmune hepatitis following liver transplantation in adults. *Hepatology* 2001; 34: 464-470.
153. Salcedo M, Vaquero J, Banares R, Rodriguez-Mahou M, Alvarez E, Vicario JL, Hernandez-Albujar A, Tiscar JLR, Rincon D, Alonso S, De Diego A, Clemente G. Response to steroids in de novo autoimmune hepatitis after transplantation. *Hepatology* 2002; 35: 349-356.
154. Hess AD, Fischer AC, Horwitz LR, Laulis MK. Cyclosporine-induced autoimmunity: critical role of autoregulation in the prevention of major histocompatibility class II-dependent autoaggression. *Transplant Proc* 1993; 25: 2811-2813.
155. Czaja AJ. Autoimmune hepatitis after liver transplantation and other lessons of self-intolerance. *Liver Transplantation* 2002; 8: 505-513.
156. Vierling JM, Flores PA. Evolving new therapies of autoimmune hepatitis. *Clin Liver Dis* 2002; 6: 825-850.
157. Czaja AJ. Emerging treatments for autoimmune hepatitis. *Current Drug Targets-Inflammation & Allergy* 2002; 1: 317-326.
158. Mistilis SP, Vickers CR, Darroch MH, McCarthy SW. Cyclosporin, a new treatment for autoimmune chronic active hepatitis. *Med J Austral* 1985; 143: 463-465.
159. Hyams JS, Ballou M, Leichtner AM. Cyclosporine treatment of autoimmune chronic active hepatitis. *Gastroenterology* 1987; 93: 890-893.
160. Person JL, McHutchison JG, Fong TL, Redeker AG. A case of cyclosporine-sensitive, steroid resistant, autoimmune chronic active hepatitis. *J Clin Gastroenterol* 1993; 17: 317-320.
161. Jackson LD, Song E. Cyclosporin in the treatment of corticosteroid resistant autoimmune chronic active hepatitis. *Gut* 1995; 36: 459-461.
162. Fernandes NF, Redeker AG, Vierling JM, Villamil FG, Fong TL. Cyclosporine therapy in patients with steroid resistant autoimmune hepatitis. *Am J Gastroenterol* 1999; 94: 241-248.
163. Debray D, Maggiore G, Girardet JP, Mallet E, Bernard O. Efficacy of cyclosporin A in children with type 2 autoimmune hepatitis. *J Pediatrics* 1999; 135: 111-114.
164. Alvarez F, Ciocca M, Canero-Velasco C, Ramonet M, de Davila MT, Cuarterolo M, Gonzalez T, Jara-Vega P, Camarena C, Brochu P, Drut R, Alvarez E. Short-term cyclosporine induces a remission of autoimmune hepatitis in children. *J Hepatol* 1999; 30: 222-227.
165. Malekzadeh R, Nasser-Moghaddam S, Kaviani M-J, Taheri H, Kamalian N, Sotoudeh M. Cyclosporin-A is a promising alternative to corticosteroids in autoimmune hepatitis. *Dig Dis Sci* 2001; 46: 1321-1327.
166. Allison AC, Eugui EM. Mycophenolate mofetil and its mechanisms of action. *Immunopharmacology* 2000; 47: 85-118.
167. Richardson PD, James PD, Ryder SD. Mycophenolate mofetil for maintenance of remission in autoimmune hepatitis patients resistant to or intolerant of azathioprine. *J Hepatol* 2000; 33: 371-375.
168. Devlin SM, Swain MG, Urbanski SJ, Burak KW. Mycophenolate mofetil for the treatment of autoimmune hepatitis in patients refractory to standard therapy. *Can J Gastroenterol* 2004; 18: 321-326.
169. Czaja AJ, Carpenter HA. Empiric therapy of autoimmune hepatitis with mycophenolate mofetil: a retrospective comparison of treatment outcomes. *J Clin Gastroenterol* (submitted).
170. Thomson AW, Carroll PB, McCauley J, Woo J, Abu-Elmagd K, Starzl TE, Van Thiel DH. FK 506: a novel immunosuppressant for treatment of autoimmune disease. Rationale and preliminary clinical experience at the University of Pittsburgh. *Springer Semin Immunopathol* 1993; 14: 323-344.
171. Van Thiel DH, Wright H, Carroll P, Abu-Elmagd K, Rodriguez-Rilo H, McMichael J, Irish W, Starzl TE. Tacrolimus: a potential new treatment for autoimmune chronic active hepatitis: results of an open-label preliminary trial. *Am J Gastroenterol* 1995; 90: 771-776.
172. Danielsson A, Prytz H. Oral budesonide for treatment of autoimmune chronic active hepatitis. *Aliment Pharmacol Ther* 1994; 8: 585-590.
173. Pratt DS, Flavin DP, Kaplan MM. The successful treatment of autoimmune hepatitis with 6-mercaptopurine after failure with azathioprine. *Gastroenterology* 1996; 110: 271-274.
174. Rebollo Bernardez J, Cifuentes Mimoso C, Pinar Moreno A, Caunedo Alvarez A, Salas Herrero E, Jimenez-Saenz M, Herrerias Gutierrez J. Deflazacort for long-term maintenance of remission in type 1 autoimmune hepatitis. *Rev Esp Enferm Dig* 1999; 91: 630-638.
175. Nakamura K, Yoneda M, Yokohama S, Tamori K, Sato Y, Aso K, Aoshima M, Hasegawa T, Makino I. Efficacy of ursodeoxycholic acid in Japanese patients with type 1 autoimmune hepatitis. *J Gastroenterol Hepatol* 1998; 13: 490-495.
176. Burak KW, Urbanski SJ, Swain MG. Successful treatment of refractory type 1 autoimmune hepatitis with methotrexate. *J Hepatol* 1998; 29: 990-993.
177. Kanzler S, Gerken G, Dienes HP, Meyer zum Buschenfelde KH, Lohse AW. Cyclophosphamide as alternative immunosuppressive therapy for autoimmune hepatitis - report of three cases. *Z Gastroenterol* 1997; 35: 571-578.
178. Czaja AJ, Carpenter HA, Lindor KD. Ursodeoxycholic acid as adjunctive therapy for problematic type 1 autoimmune hepatitis: a randomized placebo-controlled treatment trial. *Hepatology* 1999; 30: 1381-1386.
179. Czaja AJ, Lindor KD. Failure of budesonide in a pilot study of treatment-dependent autoimmune hepatitis. *Gastroenterology* 2000; 119: 1312-1316.
180. Fridkis-Hareli M, Rosloniec EF, Fugger L, Strominger JL. Synthetic peptides that inhibit binding of the collagen type II 261-273 epitope to rheumatoid arthritis-associated HLA-DR1 and -DR4 molecules and collagen-specific T-cell responses. *Hum Immunol* 2000; 61: 640-650.
181. Schwartz RS. The new immunology - the end of immunosuppressive drug therapy? (editorial). *N Engl J Med* 1999; 340: 1754-1756.
182. Guinan EC, Boussiotis VA, Neuberger D, Brennan LL, Hirano N, Nadler LM, Gribben JG. Transplantation of anergic histoincompatible bone marrow allografts. *N Eng J Med* 1999; 340: 1704-1714.
183. Wardrop RM, Whitacre CC. Oral tolerance in the treatment of inflammatory autoimmune diseases. *Inflamm Res* 1999; 48: 106-119.
184. Nagler A, Pines M, Abadi U, Pappo O, Zeira M, Rabbani E, Engelhardt D, Ohana M, Chowdhury NR, Chowdhury JR, Ilan Y. Oral tolerization ameliorates liver disorders associated with chronic graft versus host disease in mice. *Hepatology* 2000; 31: 641-648.
185. Lohse AW, Dienes HP, Meyer zum Buschenfelde K-H. Suppression of murine experimental autoimmune hepatitis by T-cell vaccination or immunosuppression. *Hepatology* 1998; 27: 1536-1543.

186. Rutgeerts P, D'Haens G, Targan S, Vasiliauskas E, Hanauer SB, Present DH, Mayer L, Van Hogezaand RA, Braakman T, DeWoody KL, Schaible TF, Van Deventer SJ. Efficacy and safety of retreatment with anti-tumor necrosis factor antibody (infliximab) to maintain remission in Crohn's disease. *Gastroenterology* 1999; 117: 761-769.
187. Nelson DR, Lauwers GY, Lau JY, Davis GL. Interleukin 10 treatment reduces fibrosis in patients with chronic hepatitis C: a pilot study of interferon nonresponders. *Gastroenterology* 2000; 118: 655-660.
188. Touhy VK, Mathisen PM. *Gene therapy for autoimmune diseases*. In: Manns MP, Paumgartner G, Leuschner U, eds, *Immunology and Liver*, Falk Symposium 114, Kluwer Academic Publishers, BV, Dordrecht, 2000, pp 376-385.
189. Czaja AJ. Chronic active hepatitis: the challenge for a new nomenclature. *Ann Intern Med* 1993; 119: 510-517.
190. Czaja AJ. The variant forms of autoimmune hepatitis. *Ann Intern Med* 1996; 125: 588-598.
191. Ben-Ari Z, Czaja AJ. Autoimmune hepatitis and its variant syndromes. *Gut* 2001; 49: 589-594.
192. Carpenter HA, Czaja AJ. The role of histologic evaluation in the diagnosis and management of autoimmune hepatitis and its variants. *Clin Liver Dis* 2002; 6: 685-705.
193. Czaja AJ. Frequency and nature of the variant syndromes of autoimmune liver disease. *Hepatology* 1998; 28: 360-365.
194. Czaja AJ, Carpenter HA. Autoimmune hepatitis with incidental histologic features of bile duct injury. *Hepatology* 2001; 34: 659-665.
195. Lohse AW, Meyer zum Buschenfelde KH, Kanzler FB, Gerken G, Dienes HP. Characterization of the overlap syndrome of primary biliary cirrhosis (PBC) and autoimmune hepatitis: evidence for it being a hepatic form of PBC in genetically susceptible individuals. *Hepatology* 1999; 29: 1078-1084.
196. Chazouilleres O, Wendum D, Serfaty L, Montebault S, Rosmorduc O, Poupon R. Primary biliary cirrhosis-autoimmune hepatitis overlap syndrome: clinical features and response to therapy. *Hepatology* 1998; 28: 296-301.
197. Czaja AJ, Carpenter HA, Santrach PJ, Moore SB. Autoimmune cholangitis within the spectrum of autoimmune liver disease. *Hepatology* 2000; 31: 1231-1238.