



## Original Article

# Phytotherapeutic compound YHK exerts an inhibitory effect on early stage of experimentally-induced neoplastic liver lesions

F Marotta;<sup>1,4</sup> M Harada;<sup>2</sup> KL Goh;<sup>3</sup> A Lorenzetti;<sup>1</sup> F Gelosa;<sup>1</sup> E Minelli<sup>4</sup>

## Abstract

**The aim of this study was to investigate the effects of the herbal compound YHK on hepatocarcinogenesis induced by diethylnitrosamine (DEN) in Sprague Dawley rats. Rats were randomly divided into 3 groups and followed up for 15 weeks. Groups 1 was given standard food and represented the healthy control. Liver preneoplastic foci were induced using the DEN method in groups 2 and 3 (20 rats each). However, group 3 was concomitantly given 50mg/kg/day of YHK. For quantitative assessment of liver preneoplastic foci, the placental form of glutathione-S-transferase (GST-P) positive foci were measured using immunohistochemical staining and image analysis. Treatment using DEN caused a significant decrease in body weight and increase in liver weight compared to the control group while concomitant supplementation with YHK prevented body weight loss and liver weight increase. As compared to DEN-only treated rats, the group given YHK showed a significant decrease in the number, size and volume of GST-P-positive foci. Moreover, co-administration of YHK significantly reduced the incidence, number, size and volume of hepatocellular carcinoma. Anti-inflammatory, anti-fibrotic as well as antioxidative properties of this compound are mechanisms which are likely to be advocated for to explain its protective effect. It is concluded that herbal compound YHK by preventing hepatocarcinogenesis in DEN-induced liver preneoplastic lesions in rats has the potential to a large clinical application as a functional food.**

**Key words:** Liver precancerous lesion, YHK, phytotherapy, hepatocarcinogenesis.

## Introduction

Nowadays there is a concern over the possible involvement of xenobiotics in carcinogenesis and this has led to a solid research stream in both humans and rodents. Indeed, DNA can be damaged along the whole process of absorption of carcinogens into the body, distribution to most sensitive tissues, metabolism which gives rise to a further form reacting with DNA, detoxification, and excretion. There are many genotoxic carcinogens occurring naturally in our environment, including the large group of heterocyclic amine mutagens.<sup>1,2</sup> For instance, 2-amino-3,8-dimethylimidazo[4,5-*f*]quinoxaline (MeIQx), one of food-derived agent, is able to induce in rats DNA adduct formation in the liver<sup>3</sup> while an overt hepatocellular carcinomas might develop with treatment at high doses.<sup>4</sup> In clinical practice, it has been shown that the human daily intake of MeIQx is estimated to be 0.2– 2.6 µg/subject<sup>5</sup> and this substance can be recovered and quantified in the urine of healthy volunteers after eating cooked meat.<sup>6-8</sup> However, most important, MeIQx-DNA adducts have been found in kidney and colon tissues in man.<sup>9</sup> Besides the primary importance of the detection and removal of such potential carcinogenetic agents, a protective dietary approach would represent an ideal countermeasure. Indeed, against the constant exposure to xenobiotics, several protective natural compounds have been proposed while only few have proven to possess a validated property in experimental tests. We have previously shown either in vitro and in vivo experimental studies that YHK exerts a potent protective effect on hepatocellular damage and on liver microcirculation in an ischemia-reperfusion model<sup>10,11</sup> as well as exerting potent in vitro protective effect on metal-induced oxidative stress of hepatocytes.<sup>12</sup> Indeed, on the clinical ground, to the contrary of many herbal remedies experimentally tested, the present phytotherapeutic compound has shown to significantly lower within two-three weeks the ALT level in the majority of HCV-related chronic liver disease patients and, moreover, to decrease Maruyama score in an awarded pilot clinical study.<sup>13</sup> Quite recently, a separate Canadian group has

<sup>1</sup> Hepato-Gastroenterology Unit, S. Giuseppe Hospital, Milan, Italy.

<sup>2</sup> MCH Hospital, Tokyo, Japan.

<sup>3</sup> Dept of Gastroenterology and Hepatology, University of Malaya, Kuala Lumpur, Malaysia.

<sup>4</sup> WHO-Cntr. for Biotech & Nat. Med., University of Milan, Italy.

Address for correspondence:

Prof. F. Marotta, MD, PhD

via Pisanello, 4

20146 Milano, Italy

E-mail: fmarchimede@libero.it

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shown its therapeutic effect in patients with non-alcohol steatohepatitis who had been studied either biochemically and histologically.<sup>14</sup> The rat liver represents an ideal model to study the whole sequence of cancer initiation and development since within a few days after administration of various genotoxic hepatocarcinogens single hepatocytes express placental glutathione S-transferase [single placental glutathione S-transferase-positive (GST-P) cells].<sup>15,16</sup> A separate population of such GST-P single cells develop into GST-P foci, which increase further in number and size on treatment with tumour promoters to a final transformation into GST-P tumours. Therefore, single GST-P cells are considered 'initiated' and their number as well as the number and size of GST-P foci can be used as quantitative indicators of subsequent cancer risk<sup>15</sup> although not all positive liver foci may necessarily develop tumours. Nonetheless, there is an over 90% correlation between the two events while the assay also correlates with the incidence of hepatocellular carcinomas in parallel long-term studies.<sup>17</sup> Given these premises, the aim of the present study was to test the potential protective effect of YHK in the early stage of chemical hepatocarcinogenesis.

## Materials & methods

Sprague Dawley rats were housed and maintained in 12-hour light/dark cycles at 23° with a humidity of 60%/10% in an environmentally-controlled vivarium (temperature, ventilation, humidity and light-dark cycle) and with free access to deionized water and non-nutrient fibers ad libitum. The animals were allowed to acclimatize for 15 weeks.

**Experimental protocol** Sixty rats were randomly divided into 3 groups of 20 rats each and treated as follows until the end of the experiment: Group 1 was given regular chow pellet and served as healthy control; Group 2, given standard chow pellet and Group 3, given the standard chow pellet containing YHK (panax pseudo-ginseng, Eucommia Ulmoides, polygonati rhizome, glycyrrhiza licorice, panax ginseng, Kyotsu Jigyo, Tokyo, Japan) calculated as to assure a daily intake of 50mg/kg, represented the hepatocarcinogenesis model. Then, they received a single intraperitoneal injection of diethylnitrosamine (DEN) (200 mg/kg/bw in saline), as described by Solt and Farber<sup>18</sup> with minor modification. The proper mixture between standard food and powdered YHK was prepared each day and the food trays were checked every day, cleared of debris, weighed and filled.

**Histopathological analysis and Glutathione S-transferase placental form (GST-P) staining and counting** At the end of the 15-week study period, rats were sacrificed and macroscopic examination was performed to detect any external pathology. Livers were then excised and weighed. Then, 5 mm-thick slices were cut from each lobe in individual rats and quickly fixed in cold acetone (0-4°C) for 6h. The slices were then embedded in paraffin for subsequent immunohistochemical examination of

GST-P. GST-P foci (defined as lesions of the cells of more than 0.01 mm<sup>2</sup> in area) were assayed by an immunohistochemical method using a streptavidin-biotin-peroxidase complex (ABC) as described by Hsu et al.<sup>19</sup> Briefly, after being deparaffinized with xylene, quenched with hydrogen peroxide and blocked with normal serum, the liver tissue sections were treated sequentially with normal goat serum, anti-rabbit GST-P antibody (1:2000), biotin-labeled goat anti-rabbit IgG (1:400) and finally with ABC. The diaminobenzidine method<sup>20</sup> was used to demonstrate the sites of peroxidase binding. For quantitative assessment of lesions it was considered: the number of GST-positive foci/cm<sup>2</sup>, the percentage of section area occupied by the foci and diameters of GST-P-positive foci and nodules >0.2 mm, as described elsewhere<sup>21,22</sup> by using an image analyzer. Liver lesions were diagnosed according to the criteria described by Squire and Levitt<sup>23</sup> and the descriptions given following the guidelines of the Institute of Laboratory Animal Resources.<sup>24</sup>

## Statistical analysis

Results are expressed as mean±s.d. Statistical analysis was performed using an SPSS programme for Windows XP. The differences between groups were evaluated using one way analysis of variance, followed by Dunnette's test for pair-wise comparison and Tukey's family error rate. In all cases,  $p<0.05$  was considered as the minimum level of statistical significance.

## Results

### Body and liver weight

All the rats survived in good condition until the scheduled sacrifices. Treatment with DEN significantly decreased the body weight and increased the liver weight compared to the control group ( $p<0.05$ ) (*Table I*). Oral supplementation YHK proved to prevent DEN-induced body weight loss and liver weight increase ( $p<0.05$ ).

### Assessment of GST-positive hepatocellular foci.

The results of quantitative analysis of the frequency of GST-P-positive foci are summarized in *table II*. The two-dimensional assessment showed that GST-P-positive lesions were significantly lesser in rats administered YHK (group 3) than in group 2 (*Figure 1*). The same results appeared when the statistical analysis was applied to volumetric assessment too such as number of lesions per cm<sup>3</sup>, mean volume and the volume when expressed as a percentage of parenchyma of GST-P-positive lesions.

**Tumor incidence.** No liver tumors was detected in untreated rats while tumor hepatocellular origin were observed in DEN-treated rats (groups 2 and 3). The inci-

**Table I.** Body and liver weight changes during den-induced hepatocarcinogenesis: Effect of concomitant supplementation with YHK

Group	Treatment	Body weight		Liver weight grams
		start of the study	end of the study	
1	Standard food only	127 ± 1	353 ± 9	14.7 ± 0.5
2	DEN	125 ± 2	321 ± 5 *	16.9 ± 0.5 *
3	DEN + YHK 50mg/kg/day	126 ± 1	349 ± 8 §	15.0 ± 0.6 §

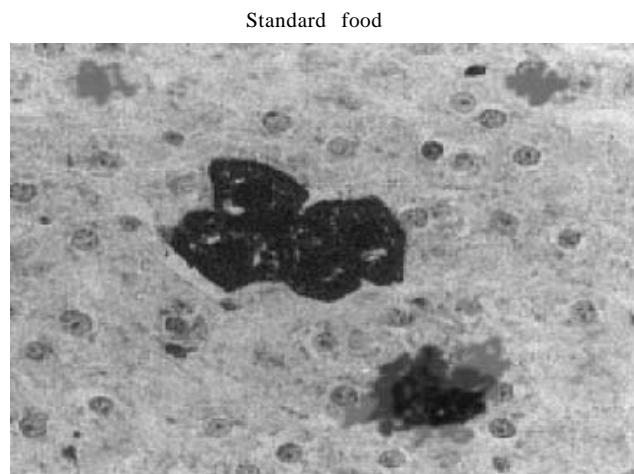
\* p&lt;0.05 vs YHK-supplemented rats and vs healthy control rats

§ p&lt;0.05 vs DEN-only treated rats

**Table II.** Number and size of GST-P-positive hepatic lesions in DEN-Induced hepatocarcinogenesis: Effect of concomitant supplementation with YHK

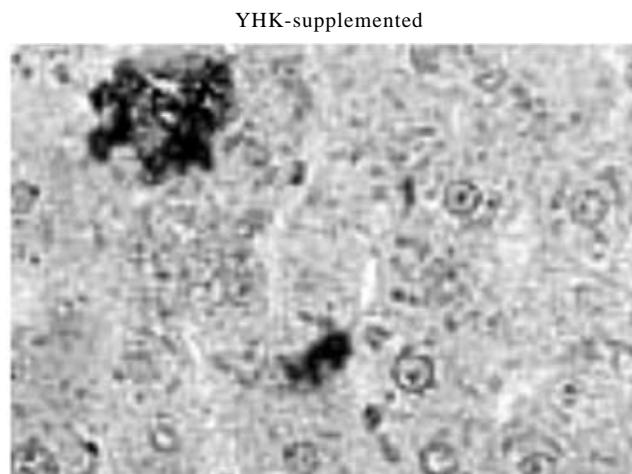
Group	2 DEN	3 DEN + YHK 50mg/kg/day
No./cm <sup>2</sup>	12 ± 4	6 ± 3 *
Mean area (mm <sup>2</sup> )	0.32 ± 0.04	0.25 ± 0.03 *
No./cm <sup>3</sup>	2,012 ± 133	1,545 ± 109 *
Mean volume (mm <sup>3</sup> )	0.17 ± 0.03	0.14 ± 0.02 *
Foci/parenchyma %	28.2 ± 2.5	21.7 ± 2.1 *

\* p&lt;0.05 vs DEN-only treated rats

**Table III.** Incidence, number, size and volume of DEN-Induced hepatocellular carcinoma: Effect of concomitant supplementation with YHK

Group	2 DEN	3 DEN + YHK 50mg/kg/day
No. of rats with HCC (%)	96 ± 4	71 ± 4
Mean area (mm <sup>2</sup> )	1.40 ± 0.47	0.17 ± 0.09 *
No./cm <sup>3</sup>	1.3 ± 0.3	0.8 ± 0.2 *
Mean volume (mm <sup>3</sup> )	0.79 ± 0.28	0.02 ± 0.01 *
HCC/parenchyma %	0.7 ± 0.2	0.2 ± 0.1 *

\* p&lt;0.01 vs DEN-only treated rats

**Figure 1.** GST-P positive staining in DEN-induced liver lesions (x400). Standard food-fed rats showed a greater number of positive foci with clustering appearance. YHK-supplemented rats showed a markedly lesser number of scattered foci and with milder staining intensity.

dences of carcinoma of group 3 were significantly lower than those of group 2 (p<0.01). The multiplicities of carcinoma and total tumors of groups 3 were significantly smaller than the corresponding values of group 2, when also assessed by volumetric calculation (p<0.05).

## Discussion

Hepatocellular carcinoma (HCC) is a devastating and increasingly common disease and progress in the management of this cancer has been slow while a high rate of recurrence is still a limiting factor in the success of surgical resection.<sup>25</sup> While hepatitis C and B and aflatoxin in some areas are the main cause of HCC, there is an increasing concern over the wider involvement of xenobiotics in car-

cinogenesis. Indeed, there are many genotoxic carcinogens occurring naturally in our environment, such as the large group of heterocyclic amine mutagens.<sup>1,2</sup> A number of chemicals agents are currently employed to experimentally mimick such condition since genotoxic carcinogens can induce irreversible DNA damage in primary cells which then proliferate in the presence of promoter substances until they acquire autonomic growth capability. Classically, chemical hepatocarcinogenesis is regarded as a multistep process with at least three stages, i.e. initiation, promotion and progression, and each of these steps involves host biochemical, endocrinological, immunological, and microenvironmental regulatory systems. On a practical ground, DNA can be damaged along the whole process of absorption of carcinogens into the body, dis-

tribution to most sensitive tissues, metabolism which gives rise to a further form reacting with DNA, detoxification, and excretion. In this instance, a protective dietary approach would represent an ideal strategy when considering that there is an established evidence that diet plays a major role in the prevention of many diseases, including cancer.<sup>26</sup> Thus, nowadays there is an increasing literature supporting the benefit of specific nutrients which, back in the early 1980s, had been termed in Japan as "functional foods". In the present study we employed YHK, a controlled herbal remedy, which has been shown to exert potent hepatoprotective properties in several experimental models of liver injury.<sup>10-12</sup> However, unlike other natural remedies which are either unapplicable to human trials or have shown to occasionally trigger severe side effects,<sup>27,28</sup> being regarded as unsafe in long-term treatment,<sup>29</sup> this compound can be safely integrated in normal diet and long-term studies have proved to significantly exert a transaminases-lowering effect in HCV-related cirrhotic patients<sup>13</sup> and, most recently, in non-alcohol steatohepatitis.<sup>14</sup> This study showed that this compound, when orally ingested together with an established chemical hepatotoxin, could counteract the early phase of carcinogenesis, as expressed by the study of GST-P. This represents a stable marker for persistent preneoplastic and neoplastic cells not only at the protein but also at the mRNA level throughout hepatocarcinogenesis in rats.<sup>30,31</sup> Overall, the GSTs are a family of dimeric proteins (labelled as Alpha, Mu, Pi, Theta, Sigma, Kappa, and Zeta) that play important roles in both the intracellular transport of hydrophobic molecule and the metabolism of toxic compounds.<sup>32</sup> GST-P protein is hardly detectable in normal rat liver but becomes expressed and detectable in hyperplastic nodules and hepatocellular carcinomas, irrespective of the kind of carcinogen used. GST-positive cells are typically characterized by an elevated DNA replication and the growth of GST-P-positive single cells and GST-P-positive liver foci is believed to be results between such replication and the counterbalance determined by death of cells.<sup>33,34</sup> However, as stated above, a number of chemical and/or dietary toxins may act as tumor promoters by triggering a progressive cellular damage. In this context, are of worthwhile interest the data of Matsui et al. showing the beneficial effect of a functional food approach in improving the prognosis of postoperative HCC<sup>35</sup> which has generated a thoughtful editorial stressing the need of well-planned studies tackling such complex issue.<sup>36</sup> In particular, our study showed that the number, size and volume of either GST-P-positive foci and of overt HCC were significantly reduced by co-administration of YHK, the latter event being at an higher significance level. In general, a number of mechanisms underlie the effects of chemopreventive agents, the suppression of lipid peroxidation or DNA adduct formation and the modulation of phase I or II enzymes being among them. More recently,

we have shown in that this compound could offer a significant protection to hepatocytes and lysosomal fraction against metal-induced oxidative stress<sup>12</sup> as well as modulating phase I and phase II carcinogens-metabolizing enzymes (Marotta et al. personal data, manuscript in preparation). The mechanism by which YHK provides significant protection against hepatocarcinogenesis is not clear as yet. However, taken overall it is conceivable that some YHK components endowed by potent antioxidant/antiinflammatory property such as eucommia ulmoides and panax pseudo-ginseng<sup>37,38</sup> and anti-proliferative or pro-apoptotic effect such as gallic acid and glycyrrhiza,<sup>39,40</sup> are advocated for to explain its prevention of preneoplastic lesion formation. On the other hand, its safety make it a potential functional food of large clinical application in the quest to achieve a better control of HCC transformation in chronic liver disease.<sup>41</sup> This holds of particular interest when considering that a number of "would-be" protective natural compound have failed to do the same<sup>42,43</sup> if not even worsen the carcinogenesis process.<sup>44,45</sup>

## References

1. Sugimura T. *History, present and future, of heterocyclic amines, cooked food mutagens*. In "Heterocyclic Amines in Cooked Foods: Possible Human Carcinogens," ed. R. H. Adamson, J.-Å. Gustafsson, N. Ito, M. Nagao, T. Sugimura, K. Wakabayashi and Y. Yamazoe, 1995: 214-231. Princeton Scientific Publishing Co., Inc., Princeton, NJ.
2. Sugimura T. Nutrition and dietary carcinogens. *Carcinogenesis* 2000; 21: 387-395.
3. Yamashita K, Adachi, M, Kato S, Nakagama H, Ochiai M, Wakabayashi K, Sato S, Nagao M, Sugimura T. DNA adducts formed by 2-amino-3,8-dimethylimidazo[4,5-f]quinoxaline in rat liver: dose-response on chronic administration. *Jpn J Cancer Res* 1990; 81:470-476.
4. Kato T, Ohgak, H, Hasegawa H, Sato S, Takayama S, Sugimura T. Carcinogenicity in rats of a mutagenic compound, 2-amino-3,8-dimethylimidazo[4,5-f]quinoxaline. *Carcinogenesis* 1988; 9: 71-73.
5. IARC Working Group on the Evaluation of Carcinogenic Risk to Humans. MeIQx (2-amino-3,8-dimethylimidazo [4,5-f]quinoxaline. In "IARC Monographs on the Evaluation of Carcinogenic Risks to Humans-Some Naturally Occurring Substances: Food Items and Constituents, Heterocyclic Aromatic Amines and Mycotoxins," 1993; 56: 211-228. IARC, Lyon.
6. Murray S, Gooderham NJ, Boobis AR, Davis DS. Detection and measurement of MeIQx in human urine after ingestion of a cooked meat meal. *Carcinogenesis* 1989; 10: 763-765.
7. Tannenbaum SR, Stillwell WG, Ji H, Skipper PL, Yu MC, Ross RK, Henderson BE, Turesky RJ, Gross GA. MeIQx (2-amino-3,8-dimethylimidazo [4,5-f]quinoxaline): metabolism in humans and urinary metabolites in human populations. In: "Heterocyclic Amines in Cooked Foods: Possible Human Carcinogens," ed. RH Adamson JÅ Gustafsson, N Ito, M Nagao, T. Sugimura K. Wakabayashi and Y Yamazoe, 1993: 197-206. Princeton Scientific Publishing Co., Inc., Princeton, NJ.
8. Ushiyama H, Wakabayashi K, Hirose M, Itoh H, Sugimura T, Nagao M. Presence of carcinogenic heterocyclic amines in urine of healthy volunteers eating normal diet, but not in patients receiving parenteral alimentation. *Carcinogenesis* 1991; 12: 1417-1422.
9. Totsuka Y, Fukutome K, Takahashi M, Takahashi S, Tada A, Sugimura T, Wakabayashi K. Presence of N2-(deoxyguanosin-8-yl)-2-amino-3,8-dimethylimidazo[4,5-f]quinoxaline (dG-C8-MeIQx) in human tissues. *Carcinogenesis* 1996; 17: 1029-1034.

10. Marotta F, Rouge A, Harada M, Anzulovic H, Idéo GM, Yanaihara N, Princess G, Idéo G. Beneficial effect of a controlled Chinese herbal remedy, K-17-22, in CCl4-induced liver toxicity: an in vivo and in vitro study. *Biomed Res* 2001; 22:167-174.
11. Marotta F, Bertuccelli J, Albergati F, Harada M, Safran P, Yanaihara N, Idéo G. Ischemia-reperfusion liver injury: effect of a nutritional approach with K-17.22 on hepatic antioxidant defense system. *Biomed Res* 2001; 22: 221-227.
12. Marotta F, Lecroix P, Harada M, Masulair K, Safran P, Lorenzetti A, Ono-Nita SK, Marandola P. Liver exposure to xenobiotics: the aging factor and potentials for functional foods. *Rejuven Res* 2006; 10:67-408-13.
13. Harada M, Marotta F, Sha SH, Minelli E. YHK, a novel herbal remedy with effective antifibrotic action, in chronic liver disease: a pilot clinical study aiming to a successful integrative medicine development. First JSH Single Topic Conference "Therapy of viral hepatitis and prevention of hepatocellular carcinoma" November 2002: 14-15, Yamanashi, Japan.
14. Chande N, Laidlaw M, Adams P, Marotta P. Yo Jyo Hen Shi Ko (YHK) improves transaminases in nonalcoholic steatohepatitis (NASH): a randomized pilot study. *Dig Dis Sci* 2006; 51: 1183-119.
15. Satoh K, Kitahara A, Satoh K, Ishikawa T, Tatematsu M, Ito N. The placental form of glutathione S-transferase as a new marker protein for preneoplasia in rat chemical hepatocarcinogenesis. *Gann* 1984; 75: 199-202.
16. Satoh K, Hatayama I, Tateoka N, Tamai K, Shimizu T, Tatematsu M, Ito N, Satoh K. Transient induction of single GST-P positive hepatocytes by DEN. *Carcinogenesis* 1989; 10: 2107-2111.
17. Hasegawa R, Ito N. *Hepatocarcinogenesis in the rat*. In: Waalkes MP, Ward JM. Carcinogenesis. New York: Raven Press; 1994: 39-64.
18. Solt D, Farber E. New principle for the analysis of chemical carcinogenesis. *Nature* 1976; 263: 701-703.
19. Hsu SM, Raine L, Fanger H. Use of avidin-biotin-peroxidase complex (ABC) in immuno-peroxidase techniques: a comparison between ABC and unlabeled antibody (PAP) procedures. *J Histochem Cytochem* 1981; 29: 577-580.
20. Peacock CS, Thompson IW, Van Noorden S. Silver enhancement of polymerised diaminobenzidine: increased sensitivity for immunoperoxidase staining. *J Clin Pathol* 1991; 44: 756-758.
21. Pugh TD, King JH, Koen H, Nychka D, Chover J, Wahba G, Hey Y, Goldfarb S. Reliable stereological method for estimating the number of microscopic hepatocellular foci from their transection. *Cancer Res* 1983; 43:1261-1268.
22. Campbell HA, Pitot HC, Potter VR, Laishes BA. Application of quantitative stereology to the evaluation of enzyme-altered foci in rat liver. *Cancer Res* 1982; 42: 465-472.
23. Squire RA, Levitt MH. Report of a workshop on classification of specific hepatocellular lesions in rats. *Cancer Res* 1975; 35: 3214-23.
24. Phillips DF. Conference explores ethics of animal research with critical thinking and balanced argument. *JAMA* 1996; 276: 87-8.
25. Bruix J, Llovet JM. Prognostic prediction and treatment strategy in hepatocellular carcinoma. *Hepatology* 2002; 35: 519-524.
26. Hasler CM. The changing face of functional foods. *J Am Coll Nutr* 2000; 19(suppl. 5): 499-506.
27. Ishizaki T, Sasaki F, Ameshima S, Shiozaki K, Takahashi H, Abe Y, Ito S, Kuriyama M, Nakai T, Kitagawa M. Pneumonitis during interferon and/or herbal drug therapy in patients with chronic active hepatitis. *Eur Respir J* 1996; 9: 2691-2696.
28. Yoshida Y. A non-cardiogenic type of pulmonary edema after administration of Chinese herbal medicine (shosaikoto)-a case report. *Nihon Kokyuki Gakkai Zasshi* 2003; 41: 300-303. in Japanese.
29. Nose M, Tamura M, Ryu N, Mizukami H, Ogihara Y. Sho-saiko-to and Saiko-keisei-to, the traditional Chinese and Japanese herbal medicines, altered hepatic drug-metabolizing enzymes in mice and rats when administered orally for a long time. *J Pharm Pharmacol* 2003; 55: 1419-1426.
30. Ito N, Tsuda H, Tatematsu M, Inoue T, Tagawa Y, Aoki T, Uwagawa S, Kagawa M, Ogiso T, Masui T, Imaida K, Fukushima S, Asamoto M. Enhancing effect of various hepatocarcinogens on induction of preneoplastic glutathione S-transferase placental form positive foci in rat-an approach for a new medium-term bioassay system. *Carcinogenesis* 1988; 9: 387-394.
31. Sato K, Kitahara A, Satoh K, Ishikawa T, Tatematsu M, Ito N. The placental form of glutathione S-transferaseas a new marker protein for preneoplasia in rat chemical hepatocarcinogenesis. *Gann* 1984; 75: 199-202.
32. Eaton DL, Bannmer TK. Concise review of the glutathione S-transferase and their significance to toxicology. *Toxicol Sci* 1999; 49: 156-164.
33. Bursch W, Lauer B, Timmermann-Trosiener I, Barthel G, Schuppler J, Schulte-Hermann R. Controlled death (apoptosis) of normal and putative preneoplastic cells in rat liver following withdrawal of tumor promoters. *Carcinogenesis* 1984; 5: 453-458.
34. Schulte-Hermann R, Timmermann-Trosiener I, Barthel G, Bursch W. DNA synthesis, apoptosis and phenotypic expression as determinants of growth of altered foci in rat liver during phenobarbital promotion. *Cancer Res* 1990; 50: 5127-5135.
35. Matsui Y, Uhara J, Satois S, Kaibori M, Yamada H, Kitade H. Improved prognosis of hepatocellular carcinoma patients when treated with functional food: a prospective cohort study. *J Hepatol* 2002; 37: 78-86.
36. Bass N. It could have been something they ate—functional food and the treatment of liver cancer. *J Hepatol* 2002; 37: 147-150.
37. Hsieh CL, Yen GC. Antioxidant actions of Du-Zhong (Eucommia ulmoides oliv.) towards oxidative damage in biomolecules. *Life Sci* 2000; 66: 1387-1400.
38. Zhang Y, Ye QF, Lu L, Xu XL, Ming YZ, Xiao JS. Panax notoginseng saponins preconditioning protects rat liver grafts from ischemia/reperfusion injury via an antiapoptotic pathway. *Hepatobiliary Pancreat Dis Int* 2005; 4: 207-212.
39. Yanez J, Vicente V, Alcaraz M, Castillo J, Benavente-Garcia O, Canteras M, Teruel JA. Cytotoxicity and antiproliferative activities of several phenolic compounds against three melanocytes cell lines: relationship between structure and activity. *Nutr Cancer* 2004; 49(2): 191-9.
40. Induction of apoptosis in HL-60 cells treated with medicinal herbs. Nishida S, Kikuchi S, Yoshioka S, Tsubaki M, Fujii Y, Matsuda H, Kubo M, Irimajiri K. *Am J Chin Med* 2003; 31(4): 551-62.
41. Okamoto T, Kajino K, Hino O. Hepatoprotective drugs for the treatment of virus-induced chronic hepatitis: from hypercarcinogenic state to hypocarcinogenesis state. *Jpn J Pharmacol* 2001; 98:177-180.
42. Barbisan LF, Spinardi-Barbisan AL, Moreira EL, Salvadori DM, Ribeiro LR, da Eira AF, de Camargo JL. Agaricus blazer (Himematsutake) does not alter the development of rat diethylnitrosamine-initiated hepatic preneoplastic foci. *Cancer Sci* 2003; 94: 188-192.
43. Low-Baselli A, Huber WW, Kafer M, Bukokska K, Shulte-Hermann R, Grasl-Kraupp B. Failure to demonstrate chemoprevention by the monoterpane perillyl alcohol during early rat hepatocarcinogenesis: a cautionary note. *Carcinogenesis* 2000; 21: 1869-1877.
44. Shukla Y, Aora A. Enhancing effects of mustard oil on preneoplastic hepatic foci development in Wistar rats. *Hum Exp Toxicol* 2003; 22:51-55.
45. Breinholt VM, Molck AM, Svendsen GW, Daneshvar B, Vinggaard AM, Poulsen M, Dragsted LO. Effects of dietary antioxidants and 2-amino-3-methylimidazo[4,5-f]- quinoline (IQ) on preneoplastic lesions and on oxidative damage, hormonal status, and detoxification capacity in the rat. *Food Chem Toxicol* 2003; 41: 1315-23.