



Curcumin in Hepatobiliary Disease: Pharmacotherapeutic Properties and Emerging Potential Clinical Applications

Robert W. Hu,* Elizabeth J. Carey,[†] Keith D. Lindor,^{†,‡} James H. Tabibian^{§,||}

* Department of Biology, College of Arts and Sciences, University of Pennsylvania, Philadelphia, PA, USA.

[†] Division of Gastroenterology and Hepatology, Mayo Clinic, Scottsdale, AZ, USA.

[‡] Executive Vice Provost and Dean, College of Health Solutions, Arizona State University, Phoenix, AZ, USA.

[§] Division of Gastroenterology, Department of Medicine, Hospital of the University of Pennsylvania, Philadelphia, PA, USA.

^{||} Division of Gastroenterology, Department of Medicine, Olive View-UCLA Medical Center, Sylmar, CA, USA.

ABSTRACT

Curcumin, an aromatic phytoextract from the turmeric (*Curcuma longa*) rhizome, has been used for centuries for a variety of purposes, not the least of which is medicinal. A growing body of evidence suggests that curcumin has a broad range of potentially therapeutic pharmacological properties, including anti-inflammatory, anti-fibrotic, and anti-neoplastic effects, among others. Clinical applications of curcumin have been hampered by quality control concerns and limited oral bioavailability, although novel formulations appear to have largely overcome these issues. Recent *in vitro* and *in vivo* studies have found that curcumin's cytoprotective and other biological activities may play a role in an array of benign and malignant hepatobiliary conditions, including but not limited to non-alcoholic fatty liver disease, cholestatic liver disease (e.g. primary sclerosing cholangitis), and cholangiocarcinoma. Here we provide an overview of fundamental principles, recent discoveries, and potential clinical hepatobiliary applications of this pleiotropic phytochemical.

Key words. Phytotherapy. Complementary medicine. Liver diseases. Antioxidants.

INTRODUCTION

Curcumin is a naturally-occurring phytochemical extracted from the turmeric rhizome *Curcuma longa*, a member of the ginger (Zingiberaceae) family. For centuries, it has been used as a dye, culinary spice, ceremonial substance, and traditional medicine in many cultures and countries, particularly in Southern Asia and the Indian subcontinent, where it grows naturally. A primary rationale for its use in traditional (e.g. Ayurvedic) medicine has been its putative anti-inflammatory and consistency properties.¹ More recently, curcumin has been appreciated in modern biomedical studies as having various potentially therapeutic properties, including anti-oxidant, anti-fibrotic, anti-senescent, and anti-neoplastic.²⁻⁸

Despite its long history and well-tolerated nature, curcumin has arguably not yet realized its full therapeutic potential in the prevention and treatment of human disease.

This has in part been due to concerns regarding quality control (e.g. safety, purity, and other chemical attributes), poor oral bioavailability, and ostensibly a degree of skepticism toward traditional (i.e. alternative) medicine. However, several well-characterized, more bioavailable, novel formulations of curcumin have helped facilitate its growing study and emerging (though to date off-label) complementary use in Western medicine. The potential applications thus far include, but are not limited to, prevention and treatment of hepatobiliary, digestive tract, cardiovascular, neurodegenerative, and dermatologic diseases as well as various systemic inflammatory and malignant conditions.^{1,2,8-10}

The goals of this review are to:

- Provide a synopsis of the structural, biochemical, and pharmacological aspects of curcumin.

- Summarize the relevant *in vitro* and *in vivo* experimental data supporting its beneficial properties.
- Present recent findings and perspectives regarding the potential clinical role of curcumin as a pharmacotherapy in the context of hepatobiliary disease and chemoprevention.

PHYSICAL AND CHEMICAL PROPERTIES OF CURCUMIN

Curcumin [1,7-bis (4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione] is a unique biphenolic molecule containing two ferulic acid residues joined by a methylene bridge (Figure 1). It has three key molecular functionalities: an aromatic *o*-methoxy phenolic group, an α , β -unsaturated β -diketone moiety, and a seven carbon linker. Accordingly, there are several important physicochemical features associated with the biological activity and effects of curcumin. For example, the *o*-methoxyphenol group and methylenic hydrogen are responsible for the antioxidant activity of curcumin, and curcumin donates an electron/hydrogen atom to reactive oxygen species (ROS). Curcumin interacts with a number of biomolecules through non-covalent and covalent binding; the hydrogen bonding and hydrophobicity of curcumin, arising from the aromatic functional ends and keto-enol tautomerism of the central portion along, account for the non-covalent interactions. The α , β -unsaturated β -diketone moieties covalently interact with protein thiols through Michael reaction (i.e. Michael nucleophilic addition). The β -diketo group chelates transition metals, thereby decreasing metal-induced toxicity and rendering some metal complexes with antioxidant activity as enzyme mimics. These nuanced molecular characteristics are reviewed in greater detail elsewhere.^{8,11} Notably, new analogues of curcumin have been and continue to be developed with chemical modifications to specific functional groups in order to po-

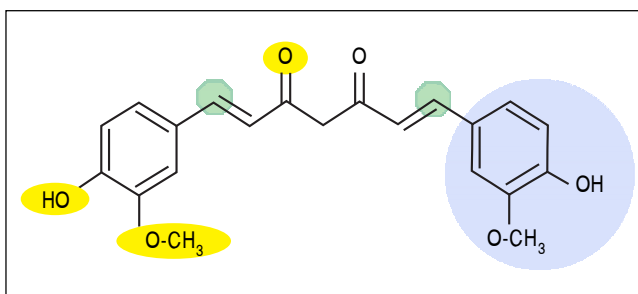


Figure 1. Molecular structure of curcumin emphasizing key functional groups and chemical properties. Yellow ovals depict hydrogen bonding sites (not shown on right side of molecule), green octagons depict Michael acceptor sites, and light blue circle depicts hydrophobic moieties (not shown on left side of molecule). In addition, keto-enol tautomerism occurs between the central portion of the molecule (di-keto tautomer shown).

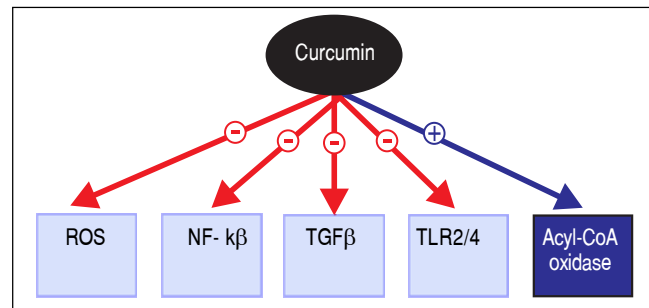


Figure 2. Proposed mechanisms of pharmacotherapeutic action of curcumin. ROS: Reactive oxygen species. NF- κ B: Nuclear factor kappa B. TGF β : Transforming growth factor beta. TLR: Toll-like receptor. Acyl-CoA: Acyl-coenzyme A.

tentiate or increase specificity of particular molecular interactions and downstream biological effects.¹¹

FORMULATIONS OF CURCUMIN

The clinical application of native curcumin has been limited due to its poor bioavailability (largely due to low solubility), physicochemical instability, and pharmacokinetics (in particular rapid metabolism), as mentioned earlier;¹² however, these issues can be largely mitigated by utilization an efficient delivery system. Therefore, various formulations of curcumin have been developed, including nanoparticles, liposomal encapsulation, and emulsions, in addition to more conventional delivery systems such as tablets and powders. These novel formulations have been found to improve curcumin's pharmacokinetics, especially its oral bioavailability, as well as its pharmacodynamics, with resultant enhanced biological activity.¹³⁻¹⁵

PHARMACOTHERAPEUTIC EFFECTS AND GENERAL APPLICATIONS

The various potentially therapeutic pharmacological properties of curcumin have shown promise in effectuating several important cytological and biological effects; these include induction of cell apoptosis, inhibition of abnormal cell proliferation, and anti-angiogenic and antimicrobial activities, among others.¹⁶ The molecular mechanisms implicated are pleiotropic, including regulation of ROS production, LPS-mediated signaling, cell survival signaling pathways by nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) and Akt, and cell survival-associated proteins such as Bcl-2 and Bcl-XL, as illustrated in figure 2,¹⁷⁻²² and it is therefore not surprising that the use of curcumin has been investigated for a variety of indications.^{8,23}

Although this review focuses on hepatobiliary applications (Figure 3), it is worth noting that curcumin has been examined in *in vitro* and animal models as well as in clinical

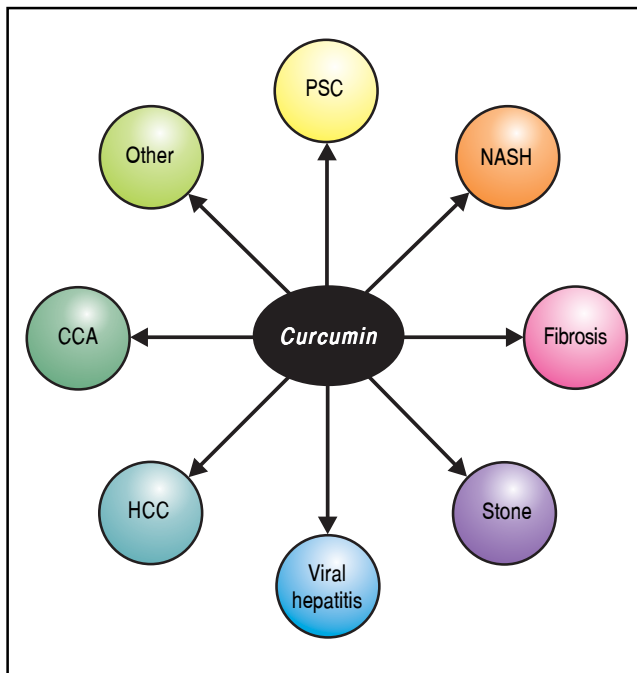


Figure 3. Potential applications of curcumin in hepatobiliary disease. CCA: Cholangiocarcinoma. HCC: Hepatocellular carcinoma. NASH: Nonalcoholic steatohepatitis. PSC: Primary sclerosing cholangitis.

studies of a wide array of human conditions, including benign or malignant neurodegenerative, cardiovascular, renal, and metabolic diseases as well as their associated complications.¹¹ Two phase I clinical trials concluded that curcumin is safe and not toxic to humans, even at very high dosage.^{5,24,25} In a minority of patients, side-effects such as nausea, diarrhea, headache, somnolence, and contact dermatitis (with topical use) have been reported; these have typically been mild and self-limited. Considering the relatively low cost and favorable safety profile of curcumin, a therapeutic impact on these diseases with this agent, even if requiring long-term treatment or as an adjuvant, represents an exciting prospect.

CURCUMIN IN NON-MALIGNANT CHRONIC HEPATOBIILIARY DISEASE

Preclinical data

A large body of preclinical literature has suggested pharmacotherapeutic properties of curcumin relevant to non-malignant (or pre-malignant) hepatobiliary disease. For example, *in vitro* studies using different cell lines such as human HSC-16 and Hep2G cells have shown significant antifibrotic effects through transforming growth factor-beta (TGF β) signaling.^{22,26,27} In addition, it has also been shown *in vitro* that curcumin has potent antioxidant

effects through inducing the activity of glutathione, glutathione peroxidase, superoxide dismutase, and catalase as well as dose-dependent induction of HO-1. Moreover, curcumin can inhibit NF- κ B activation, thereby preventing secretion of pro-inflammatory cytokines such as tumor necrosis factor alpha (TNF- α), interleukin 1 β), and interleukin 6.^{22,28}

With respect to animal model data, there is a considerable body of evidence to support potential applications of curcumin in chronic, non-malignant liver disease. For example, using a model such as CCl₄ or bile duct ligation-induced hepatic fibrosis, curcumin has been shown to significantly attenuate fibrogenesis.^{22,29,30} Similar effects were seen in the *mdr2* knockout murine model of sclerosing cholangitis with 3% curcumin chow.³¹ Extending the *mdr2* knockout model findings, our group recently found curcumin to inhibit cellular senescence and the senescence-associated secretory phenotype (unpublished data), which may be fundamentally linked to the pathogenesis of primary sclerosing cholangitis (PSC), in cultured human cholangiocyte models.^{32,33} Second, curcumin supplementation appears to reduce hepatocyte lipid accumulation attenuate nonalcoholic steatohepatitis (NASH) in rats,^{34,36} with similar findings in hamster³⁵ and rabbit³⁷ models. These benefits of curcumin appear to be mediated through mitochondrial protection and inhibition of apoptosis and are further supported by studies in a methionine and choline deficiency diet murine model demonstrating a protective role of curcumin against oxidative stress through reduced CYP2E1 and Prx1 expression and upregulated Prx6 expression.³⁸ Collectively, these and other studies^{39,40} suggest that curcumin can decrease hepatic cholesterol, triglyceride, and free fatty acid accumulation and resultant hepatic injury and thus may have a promising role in treating or preventing NASH. Third, hepato-protective effects of curcumin have been evaluated and shown in non-NASH animal models. For example, curcumin appears to inhibit CYP2E1 activity,⁴¹ thus abrogating ROS production, and activate NF-E2-related factor 2 (Nrf2) translocation to the nucleus, thereby increasing expression of antioxidant enzymes.^{42,43} Fourth, the recently identified effect of inhibiting hepatitis C virus entry in a hepatoma cell line and in primary human hepatocytes⁴⁴ as well as anti-hepatitis B virus activity in the HepG2 cell line⁴⁵ suggest that curcumin may have preventive and/or therapeutic effects in viral hepatitis. Lastly, several studies have demonstrated significant anti-lithogenic effects of curcumin in murine experimental cholesterol cholelithiasis models, thus supporting a role in gallstone disease.^{46,47}

These *in vitro* and *in vivo* data suggest that curcumin has therapeutic potential in benign chronic liver disease meriting further investigation and improved mechanistic understanding.

Clinical applications

A small but growing number of clinical trials have examined the effects of curcumin on chronic liver diseases, including antituberculosis treatment-induced hepatotoxicity,⁴⁸ gallstone disease (e.g. gallbladder hypocontractility),⁴⁹ and biliary dyskinesia.⁵⁰ To date, however, curcumin has not become a mainstream or approved therapy for these indications. Of note, we recently received approval of an investigational new drug application to conduct a clinical trial of curcumin in patients with PSC based on promising pre-clinical findings; study enrollment is planned for later this year.

CURCUMIN IN MALIGNANT HEPATOBILIARY DISEASE

Preclinical data

Preclinical data, both *in vitro* and *in vivo*, suggest that curcumin has anti-neoplastic effects with regard to hepatocellular carcinoma (HCC). Several *in vitro* studies using murine hepatoma⁵¹ or HepG2 cancer cell lines⁵² have demonstrated that its effect are likely through activation of ER stress and apoptosis or mediating MMP turnover⁵³ as well as through NF- κ B signaling.⁵⁴ *In vivo*, curcumin has similarly exhibited anti-properties in several animal models of hepatocellular carcinoma, as demonstrated in the N-bis-(2-hydroxypropyl) nitrosamine induced liver adenoma model,³⁸ HepG2 xenografts,⁵⁵⁻⁵⁷ orthotopic implantation model of HCC CBO140C12 cells.⁵⁸ Curcumin also has been shown to mitigate oxidative tissue damage during chemically induced hepatocarcinogenesis in N-nitrosodimethylamine -initiated and phenobarbital-promoted hepatocarcinogenesis in Wistar rats.⁵⁹⁻⁶¹ Moreover, a recent breakthrough study revealed that curcumin not only inhibited the proliferation and invasion of HCC cell lines *in vitro*, but also drastically suppressed primary tumor growth and lung metastases *in vivo*. Furthermore, in combination with sorafenib, curcumin induced HCC cell apoptosis and cell cycle arrest by synergistically down-regulating the expression of MMP9 via NF- κ B/P65 signaling pathway. This highlights the potential neo-adjuvant application of curcumin in the treatment of HCC.⁶²

Curcumin may also have promise in the treatment of cholangiocarcinoma (CCA), a lethal malignancy for which effective pharmacotherapy remains lacking. Several *in vitro* studies using different CCA cell lines have shown anti-tumorigenic effects on cholangiocarcinoma, which appear to be via suppressing proliferation and inducing apoptosis in malignant cells through modulating multiple cell signaling pathways.^{63,64} Similar effects were also observed *in vivo* hamster⁶⁵ and nude mouse xenograft models.⁶⁶

Collectively, these data suggest that curcumin may have potential therapeutic implications in hepatobiliary malignancies.

Clinical applications

Although there are no published clinical trials to date on malignant hepatobiliary disease with curcumin, the *in vivo* and *in vitro* data thus far may inform clinical studies of curcumin in treating such conditions.

CURCUMIN AS A HEPATOPROTECTIVE AGENT

Preclinical data

Numerous *in vivo* studies have found that curcumin has potent antioxidant and anti-inflammatory properties, which may account for its hepatoprotective effect. These properties may be mediated by inhibiting NF- κ B signaling and by inhibiting production of nitric oxide and TNF- α by activated Kupffer cells, among other mechanisms, and as mentioned earlier.^{8,67,68} This has been shown in various models of hepatobiliary injury, including D-galactosamine-, thioacetamide-, and CCl₄-, iron-, and ethanol- induced acute liver injury.^{8,69,70} It is interesting to note that experimental liver steatosis induced by TNF- α injection in mice also showed significant attenuation after curcumin treatment as indicated by decreased oxidative stress and neutrophil infiltration and improved hepatic histopathological features.⁷¹ Curcumin also has been shown to decrease hepatocyte lipid storage in various animal models.^{34,35,39,40,72}

Clinical applications

There are only limited data, much of which is related to pharmacokinetic and toxicity evaluation, with regard to clinical studies assessing the hepatoprotective (i.e. primary preventive) properties of curcumin.⁷³ Given the progress made to pharmacologically improve the bioavailability of curcumin and its demonstrable safety profile, preliminary clinical application has been thus far successful in healthy human volunteers using a nanoparticle formulation for anti-cancer (preventive or therapeutic) purposes.⁷⁴ The findings appear promising but require further study.

FUTURE DIRECTIONS

With the development of novel drug delivery systems and structural modifications to improve the oral bioavailability of curcumin, it is likely that pre-clinical research investigating its mechanisms of action in prevention and

treatment of disease and potential applications to clinical medicine will continue to expand. Given its low negligible toxicity and relatively low cost, it would reason to believe that its role in human disease, be it preventive or for treatment of benign or malignant disorders, will draw increasing medical and pharmaceutical attention. Further study with well-designed clinical trials are needed to ascertain appropriate indications, dispel potential positive publication bias, and determine optimal formulations, doses, and other regimen-related factors. In addition, epidemiological data from regions of high dietary curcumin intake (e.g. India) may shed additional light on chemopreventive properties; for example, it is suggested that may in part be related to high curcumin consumption the low incidence of colorectal and small intestinal cancer in India,⁷⁵ but data regarding hepatobiliary disease are lacking.

CONCLUSIONS

Curcumin is a phytochemical which has been used as a vital traditional medicine for centuries. Pharmacological advances, including significantly improved oral bioavailability, and curcumin's favorable adverse effect profile and low cost have helped garner growing interest in its molecular properties and biological effects as well as its possible applications in the prevention and treatment of hepatobiliary and other (e.g. pancreatobiliary) diseases. The data thus far regarding its potential role as an adjunctive or primary pharmacologic agent appears promising. Future research is expected and eagerly awaited in this regard and to better and gauge clinical efficacy and utility elucidate mechanisms of action.

ABBREVIATIONS

- **CCA:** cholangiocarcinoma.
- **HCC:** hepatocellular carcinoma.
- **NASH:** nonalcoholic steatohepatitis.
- **NF- κ B:** nuclear factor kappa-light-chain-enhancer of activated B cells.
- **PSC:** primary sclerosing cholangitis.
- **ROS:** reactive oxygen species.
- **TNF- α** tumor necrosis factor alpha.
- **TGF β :** transforming growth factor-beta.

CONFLICTS OF INTEREST, DISCLOSURES

None.

REFERENCES

1. Gupta SC, Patchva S, Koh W, Aggarwal BB. Discovery of curcumin, a component of golden spice, and its miraculous biological activities. *Clin Exp Pharmacol Physiol* 2012; 39: 283-99.
2. Aggarwal BB, Sundaram C, Malani N, Ichikawa H. Curcumin: the Indian solid gold. *Adv Exp Med Biol* 2007; 595: 1-7.
3. Chandran B, Goel A. A randomized, pilot study to assess the efficacy and safety of curcumin in patients with active rheumatoid arthritis. *Phytother Res* 2012; 26: 1719-25.
4. Goel A, Boland CR, Chauhan DP. Specific inhibition of cyclooxygenase-2 (COX-2) expression by dietary curcumin in HT-29 human colon cancer cells. *Cancer Lett* 2001; 172: 111-8.
5. Cheng AL, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, Ko JY, et al. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res* 2001; 21: 2895-900.
6. Singh S, Aggarwal BB. Activation of transcription factor NF-kappa B is suppressed by curcumin (diferuloylmethane) [corrected]. *J Biol Chem* 1995; 270: 24995-5000.
7. Maiti K, Mukherjee K, Gantait A, Saha BP, Mukherjee PK. Curcumin-phospholipid complex: Preparation, therapeutic evaluation and pharmacokinetic study in rats. *Int J Pharm* 2007; 330: 155-63.
8. Pulido-Moran M, Moreno-Fernández J, Ramírez-Tortosa C, Ramírez-Tortosa M. Curcumin and Health. *Molecules* 2016; 21: 264.
9. Lam P, Cheung F, Tan HY, Wang N, Yuen MF, Feng Y. Hepatoprotective Effects of Chinese Medicinal Herbs: A Focus on Anti-Inflammatory and Anti-Oxidative Activities. *Int J Mol Sci* 2016; 17:465.
10. Vera-Ramírez L, Pérez-López P, Varela-López A, Ramírez-Tortosa M, Battino M, Quiles JL. Curcumin and liver disease. *Biofactors* 2013; 39: 88-100.
11. Priyadarsini KI. Chemical and structural features influencing the biological activity of curcumin. *Curr Pharm Des* 2013; 19: 2093-100.
12. Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: problems and promises. *Mol Pharm* 2007; 4: 807-18.
13. Gupta SC, Patchva S, Aggarwal B. Therapeutic roles of Curcumin: lessons learned from clinical trials. *The AAPS Journal* 2013; 15: 195-218.
14. Yallapu MM, Nagesh PK, Jaggi M, Chauhan SC. Therapeutic Applications of Curcumin Nanoformulations. *AAPS J* 2015; 17: 1341-56.
15. Prasad S, Tyagi AK, Aggarwal BB. Recent developments in delivery, bioavailability, absorption and metabolism of curcumin: the golden pigment from golden spice. *Cancer Res Treat* 2014; 46: 2-18.
16. Fan X, Zhang X, DB Liu, Yan J, Liang HP. The clinical applications of Curcumin: Current state and the future. *Curr Pharm Des* 2013; 19: 2011-5.
17. Shishodia S, Sethi G, Aggarwal BB. Curcumin: getting back to the roots. *Ann N Y Acad Sci* 2005; 1056: 206-17.
18. Singh S, Aggarwal BB. Activation of transcription factor NF-kB is suppressed by curcumin (diferuloylmethane). *J Biol Chem* 1995; 270: 24995-5000.
19. Shih SC, Claffey KP. Role of AP-1 and HIF-1 transcription factors in TGF- β activation of VEGF expression. *Growth Factors* 2001; 19: 19-34.
20. Bachmeier BE, Nerlich AG, Iancu CM, Cilli M, Schleicher E, Vene R, Dell'Eva R, et al. The chemopreventive polyphenol Curcumin prevents hematogenous breast cancer metastases in immunodeficient mice. *Cell Physiol Biochem* 2007; 19: 137-52.
21. El-Azab F, Hishe H, Moustafa Y, El-Awady E. Anti-angiogenic effect of resveratrol and curcumin in ehrlich ascites carcinoma-bearing mice. *Eur J Pharmacol* 2011; 652: 7-14.

22. Bassiouny AR, Zaky A, Fawky F, Kandeel K. Alteration of AP-endonuclease1 expression in curcumin-treated fibrotic rats. *Ann Hepatol* 2011; 10: 516-30.
23. Lang A, Salomon N, Wu JC, Kopylov U, Lahat A, Har-Nov O, Ching JY, et al. Curcumin in Combination With Mesalamine Induces Remission in Patients With Mild-to-Moderate Ulcerative Colitis in a Randomized Controlled Trial. *Clin Gastroenterol Hepatol* 2015; 13: 1444-9.
24. Sharma RA, Euden SA, Platton SL, Cooke DN, Shafayat A, Hewitt HR, Marczylo TH, et al. Phase I clinical trial of oral curcumin: biomarkers of systemic activity and compliance. *Clin Cancer Res* 2004; 10: 6847-54.
25. Epstein J, Sanderson IR, Macdonald TT. Curcumin as a therapeutic agent: the evidence from in vitro, animal and human studies. *Br J Nutr* 2010; 103: 1545-57.
26. Lin YL, Lin CY, Chi CW, Huang YT. Study on antifibrotic effects of curcumin in rat hepatic stellate cells. *Phytother Res* 2009; 23: 927-32.
27. Nakayama N, Nakamura T, Okada H, Iwaki S, Sobel BE, Fujii S. Modulators of induction of plasminogen activator inhibitor type-1 in HepG2 cells by transforming growth factor- β . *Coronary Artery Dis* 2011; 22: 468-78.
28. Ghosh N, Ghosh R, Mandal V, Mandal SC. Recent advances in herbal medicine for treatment of liver diseases. *Pharm Biol* 2011; 49: 970-88.
29. Reyes-Gordillo K, Segovia J, Shibayama M, Tsutsumi V, Vergara P, Moreno MG, Muriel P. Curcumin prevents and reverses cirrhosis induced by bile duct obstruction or CCl4 in rats: role of TGF- β modulation and oxidative stress. *Fundam Clin Pharm* 2008; 22: 417-27.
30. El Swefy S, Hasan RA, Ibrahim A, Mahmoud MF. Curcumin and hemopressin treatment attenuates cholestasis-induced liver fibrosis in rats: role of CB1 receptors. *Naunyn Schmiedeberg's Arch Pharmacol* 2016; 389: 103-16.
31. Baghdasaryan A, Claudel T, Kosters A, Gumhold J, Silbert D, Thuringer A, Leski K, et al. Curcumin improves sclerosing cholangitis in Mdr2 $^{-/-}$ mice by inhibition of cholangiocyte inflammatory response and portal myofibroblast proliferation. *Gut* 2010; 59: 521-30.
32. Tabibian JH, Trussoni CE, O'Hara SP, Splinter PL, Heimbach JK, LaRusso NF. Characterization of cultured cholangiocytes isolated from livers of patients with primary sclerosing cholangitis. *Lab Invest* 2014; 94: 1126-33.
33. Tabibian JH, O'Hara SP, Splinter PL, Trussoni CE, LaRusso NF. Cholangiocyte senescence by way of N-ras activation is a characteristic of primary sclerosing cholangitis. *Hepatology* 2014; 59: 2263-75.
34. Rao DS, Sekhara NC, Satyanarayana MN, Srinivasan M. Effect of curcumin on serum and liver cholesterol levels in the rat. *J Nutr* 1970; 100: 1307-15.
35. Jang EM, Choi MS, Jung UJ, Kim MJ, Kim HJ, Jeon SM, Shin SK, et al. Beneficial effects of curcumin on hyperlipidemia and insulin resistance in high-fat-fed hamsters. *Metabolism* 2008; 57: 1576-83.
36. Wang L, Lv Y, Yao H, Yin L, Shang J. Curcumin prevents the non-alcoholic fatty hepatitis via mitochondria protection and apoptosis reduction. *Int J Clin Exp Pathol* 2015; 8: 11503-9.
37. Ramirez-Tortosa MC, Ramirez-Tortosa CL, Mesa MD, Grados S, Gil A, Quiles JL. Curcumin ameliorates rabbit's steatohepatitis via respiratory chain, oxidative stress, and TNF- α . *Free Radic Biol Med* 2009; 47: 924-31.
38. Lee SJ, Kang JH, Iqbal W, Kwon OS. Proteomic analysis of mice fed methionine and choline deficient diet reveals marker proteins associated with steatohepatitis. *PLoS One* 2015; 7: 10.
39. Rukkumani R, Sri Balasubashini M, Vishwanathan P, Menon VP. Comparative effects of curcumin and photo-irradiated curcumin on alcohol- and polyunsaturated fatty acid-induced hyperlipidemia. *Pharmacol Res* 2002; 46: 257-64.
40. Rukkumani R, Aruna K, Varma PS, Viswanathan P, Rajasekaran KN, Menon VP. Protective role of a novel curcuminoid on alcohol and PUFA-Induced hyperlipidemia. *Toxicol Mech Methods* 2005; 15: 227-34.
41. Guangwei X, Rongzhu L, Wenrong X, Suhua W, Xiaowu Z, Shizhong W, Ye Z, et al. Curcumin pretreatment protects against acute acrylonitrile-induced oxidative damage in rats. *Toxicology* 2010; 267: 140-6.
42. Charoensuk L, Pinlaor P, Prakobwong S, Hiraku Y, Laothong U, Ruangjirachuporn W, Yongvanit P, et al. Curcumin induces a nuclear factor-erythroid 2-related factor 2-driven response against oxidative and nitrate stress after praziquantel treatment in liver fluke-infected hamsters. *Int J Parasitol* 2011; 41: 615-26.
43. Farombi EO, Shrotriya S, Na HK, Kim SH, Surh YJ. Curcumin attenuates dimethylnitrosamine-induced liver injury in rats through Nrf2-mediated induction of heme oxygenase-1. *Food Chem Toxicol* 2008; 46: 1279-87.
44. Anggakusuma, Colpitts CC, Schang LM, Rachmawati H, Frentzen A, Pfaender S, Behrendt P, et al. Turmeric curcumin inhibits entry of all hepatitis C virus genotypes into human liver cells. *Gut* 2014; 63: 1137-49.
45. Waiyaput W, Payungporn S, Issara-Amphorn J, Panjaworayan NT. Inhibitory effects of crude extracts from some edible Thai plants against replication of hepatitis B virus and human liver cancer cells. *BMC Complement Altern Med* 2012; 12: 246.
46. Shubha MC, Reddy RR, Srinivasan K. Antilithogenic influence of dietary capsaicin and curcumin during experimental induction of cholesterol gallstone in mice. *Appl Physiol Nutr Metab* 2011; 36: 201-9.
47. Li Y, Li M, Wu S, Tian Y. Combination of curcumin and piperine prevents formation of gallstones in C57BL6 mice fed on lithogenic diet: whether NPC1L1/SREBP2 participates in this process? *Lipids Health Dis* 2015; 14: 100.
48. Adhvaryu MR, Reddy N, Vakharia BC. Prevention of hepatotoxicity due to anti-tuberculosis treatment: a novel integrative approach. *World J Gastroenterol* 2008; 14: 4753-62.
49. Rasyid A, Lelo A. The effect of curcumin and placebo on human gall-bladder function: an ultrasound study. *Aliment Pharmacol Ther* 1999; 13: 245-9.
50. Niederau C, Gopfert E. The effect of chelidonium- and turmeric root extract on upper abdominal pain due to functional disorders of the biliary system. Results from a placebo-controlled double-blind study. *Med Klin (Munich)* 1999; 94: 425-30.
51. Chuang SE, Kuo ML, Hsu CH, Chen CR, Lin JK, Lai GM, Hsieh CY, et al. Curcumin-containing diet inhibits diethylnitrosamine-induced murine hepatocarcinogenesis. *Carcinogenesis* 2000; 21: 331-5.
52. Xiao J, Chu Y, Hu K, Wan J, Huang Y, Jiang C, Liang G, et al. Synthesis and biological analysis of a new Curcumin analogue for enhanced anti-tumor activity in HepG 2 cells. *Oncol Rep* 2010; 23: 1435-41.
53. Jia L, Wang H, Qu S, Miao X, Zhang J. CD147 regulates vascular endothelial growth factor- α expression, tumorigenicity, and chemosensitivity to curcumin in hepatocellular carcinoma. *IUBMB Life* 2008; 60: 57-63.
54. Notarbartolo M, Poma P, Perri D, Dusonchet L, Cervello M, D'Alessandro N. Antitumor effects of curcumin, alone or in combination with cisplatin or doxorubicin, on human hepatic cancer cells. Analysis of their possible relationship to changes in NF- κ B activation levels and in IAP gene expression. *Cancer Lett* 2005; 224: 53-65.

55. Yoysungnoen P, Wirachwong P, Bhattarakosol P, Niimi H, Patumraj S. Antiangiogenic activity of curcumin in hepatocellular carcinoma cells implanted nude mice. *Clin Hemorheol Microcirc* 2005; 33: 127-35.
56. Yoysungnoen P, Wirachwong P, Bhattarakosol P, Niimi H, Patumraj S. Effects of curcumin on tumor angiogenesis and biomarkers, COX-2 and VEGF, in hepatocellular carcinoma cell-implanted nude mice. *Clin Hemorheol Microcirc* 2006; 34: 109-15.
57. Yoysungnoen P, Wirachwong P, Changtam C, Suksamrarn A, Patumraj S. Anti-cancer and anti-angiogenic effects of Curcumin and tetrahydrocurcumin on implanted hepatocellular carcinoma in nude mice. *World J Gastroenterol* 2008; 14: 2003-9.
58. Ohashi Y, Tsuchiya Y, Koizumi K, Sakurai H, Saiki I. Prevention of intrahepatic metastasis by curcumin in an orthotopic implantation model. *Oncology* 2003; 65: 250-8.
59. Piper JT, Singhal SS, Salameh MS, Torman RT, Awasthi YC, Awasthi S. Mechanisms of anticarcinogenic properties of curcumin: the effect of curcumin on glutathione linked detoxification enzymes in rat liver. *Int J Biochem Cell Biol* 1998; 30: 445-56.
60. Sreepriya M, Bali G. Chemopreventive effects of embelin and curcumin against N-nitrosodiethylamine/phenobarbital induced hepatocarcinogenesis in Wistar rats. *Fitoterapia* 2005; 76: 549-55.
61. Sreepriya M, Bali G. Effects of administration of Embelin and Curcumin on lipid peroxidation, hepatic glutathione Antioxidant defense and hematopoietic system during N-nitrosodiethylamine/Phenobarbital-induced hepatocarcinogenesis in Wistar rats. *Mol Cell Biochem* 2006; 284: 49-55.
62. Hu B, Sun D, Sun C, Sun YF, Sun HX, Zhu QF, Yang XR, et al. A polymeric nanoparticle formulation of curcumin in combination with sorafenib synergistically inhibits tumor growth and metastasis in an orthotopic model of human hepatocellular carcinoma. *Biochem Biophys Res Commun* 2015; 468: 525-32.
63. Prakobwong S, Gupta SC, Kim JH, Sung B, Pinlaor P, Hiraku Y, Wongkham S, et al. Curcumin suppresses proliferation and induces apoptosis in human biliary cancer cells through modulation of multiple cell signaling pathways. *Carcinogenesis* 2011; 32: 1372-80.
64. Suphim B, Prawan A, Kukongviriyapan U, Kongpetch S, Buranrat B, Kukongviriyapan V. Redox modulation and human bile duct cancer inhibition by curcumin. *Food Chem Toxicol* 2010; 48: 2265-72.
65. Prakobwong S, Khoontawad J, Yongvanit P, Pairojkul C, Hiraku Y, Sithithaworn P, Pinlaor P, et al. Curcumin decreases cholangiocarcinogenesis in hamsters by suppressing inflammation-mediated molecular events related to multistep carcinogenesis. *Int J Cancer* 2011; 129: 88-100.
66. Plengsuriyakarn T, Viyanant V, Eursitthichai V, Picha P, Kuproadinun P, Itharat A, Na-Bangchang K. Anticancer activities against cholangiocarcinoma, toxicity and pharmacological activities of Thai medicinal plants in animal models. *BMC Complement Altern Med* 2012; 12: 23.
67. Lukita-Atmadja W, Ito Y, Baker GL, McCuskey RS. Effect of curcuminoids as anti-inflammatory agents on the hepatic microvascular response to endotoxin. *Shock* 2002; 17: 399-403.
68. Yun SS, Kim SP, Kang MY, Nam SH. Inhibitory effect of curcumin on liver injury in a murine model of endotoxemic shock. *Biotechnol Lett* 2010; 32: 209-14.
69. Shapiro H, Ashkenazi M, Weizman N, Shahmurov M, Aeed H, Bruck R. Curcumin ameliorates acute thioacetamide-induced hepatotoxicity. *J Gastroenterol Hepatol* 2006; 21: 358-66.
70. Bisht S, Khan MA, Bekhit M, Bai H, Cornish T, Mizuma M, Rudek MA, et al. A polymeric nanoparticle formulation of curcumin (NanoCurcTM) ameliorates CCl4-induced hepatic injury and fibrosis through reduction of pro-inflammatory cytokines and stellate cell activation. *Lab Invest* 2011; 91: 1383-95.
71. Mouzaoui S, Rahim I, and Djerdjouri B. Aminoguanidine and curcuminattenuated tumor necrosis factor (TNF)- α -induced oxidative stress, colitis and hepatotoxicity in mice. *Int Immunopharmacol* 2012; 12: 302-11.
72. Asai A, Miyazawa T. Dietary curcuminoids prevent high-fat diet-induced lipid accumulation in rat liver and epididymal adipose tissue. *J Nutr* 2001; 131: 2932-5.
73. He SM, Chan E, Zhou SF. ADME properties of herbal medicines in humans: evidence, challenges and strategies. *Curr Pharm Des* 2011; 17: 357-407.
74. Kanai M, Imaizumi A, Otsuka Y, Sasaki H, Hashiguchi M, Tsujiko K, Matsumoto S, et al. (2012) Dose-escalation and pharmacokinetic study of nanoparticle curcumin, a potential anticancer agent with improved bioavailability, in healthy human volunteers. *Cancer Chemother Pharmacol* 2012; 69: 65-70.
75. Mohandas KM, Desai DC. Epidemiology of digestive tract cancers in India. V. Large and small bowel. *Indian J Gastroenterol* 1999; 18(3): 118-21.

Correspondence and reprint request:

James H. Tabibian, M.D., Ph.D.
Olive View-UCLA Medical Center
14445 Olive View Dr., 2B-182, Sylmar, CA 91342, USA
Tel.: (747) 210-4627
E-mail: jtabibian@dhs.lacounty.gov