

CHIKUNGUNYA: MOLECULAR ASPECTS, CLINICAL OUTCOMES AND PATHOGENESIS

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ABSTRACT

The alarming worldwide emergence of the chikungunya virus began in the last decade. Since the first autochthonous transmission in Mexico in November of 2014, the virus has spread throughout the country, resulted in multiple outbreaks. This virus produces an acute and self-limiting disease characterized by fever, polyarthralgia, myalgia, exanthema, and general malaise. It is transmitted to humans by the bite of *Aedes aegypti* and *A. albopictus* mosquitoes. The fact that the clinical presentation is similar to that produced by other arboviruses complicates its clinical diagnosis. The chronic stage of the disease can cause severe consequences lasting months or years, from local arthralgia to rheumatoid arthritis. In this review, we emphasize the public health threat posed by this highly disabling emerging disease, the clinical outcomes, and its possible physiopathological process. We outline the diagnosis and the impact that this virus has had in Mexico since its introduction.

Key words: Chikungunya. Pathogenesis. Molecular aspects.

INTRODUCTION

Chikungunya virus (CHIKV) is an emerging arbovirus in the Americas that affects humans, causing chikungunya fever. The virus was initially isolated from human serum and infected mosquitoes during an epidemic in Tanzania in 1953. In 2004, CHIKV reappeared on the coast of Kenya and spread to the Comoro islands, where 5,000 cases were reported¹. In 2005-2006, the virus disseminated to other islands of the Indian Ocean, mainly La Réunion where it is estimated that 300,000

people out of a population of 785,000 were infected, leading to 237 deaths^{2,3}. The epidemic reached the Asian continent in 2006 when large outbreaks in India affected an estimated 1.5 million individuals⁴.

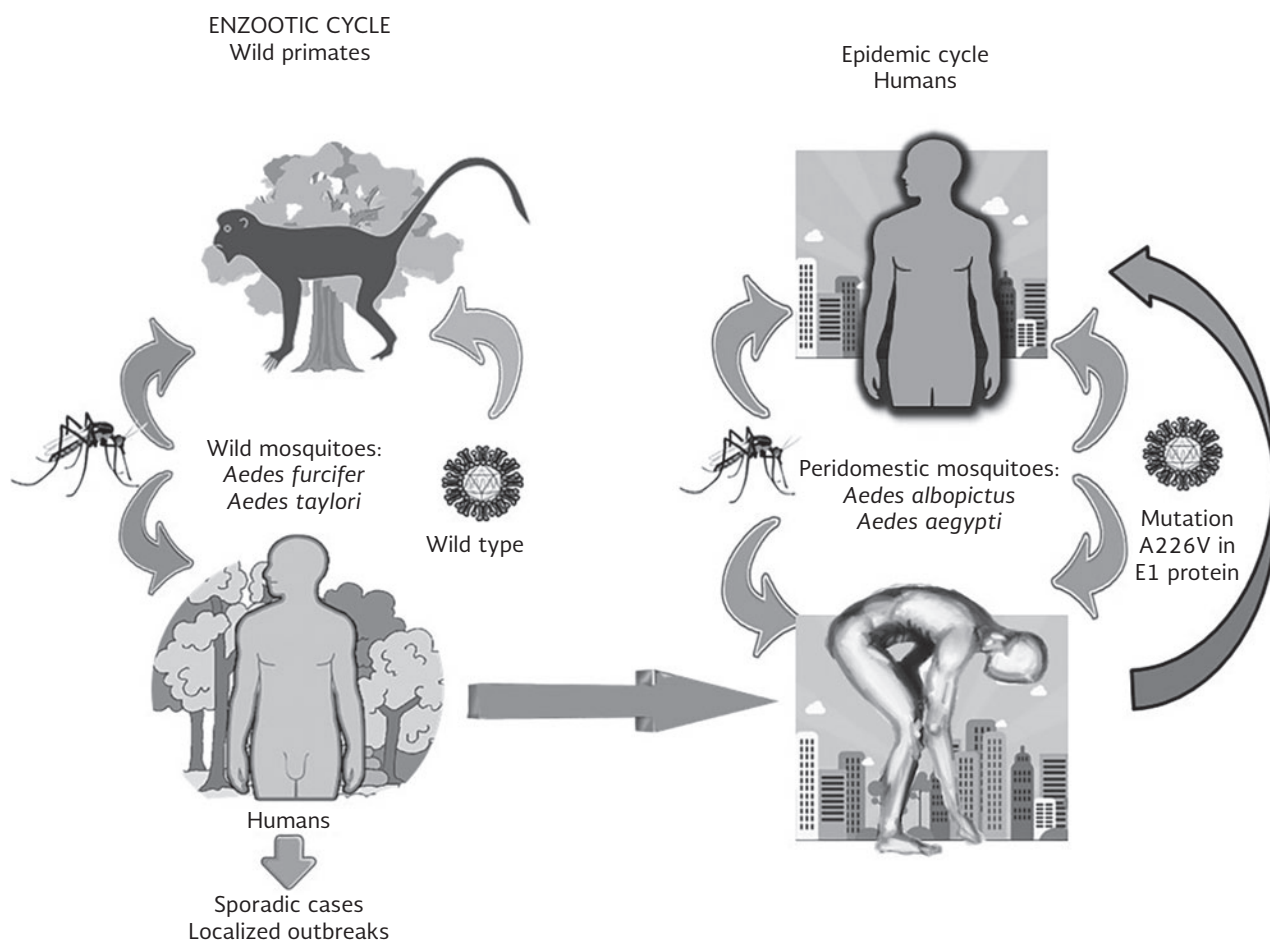
On December 6, 2013, the Pan American Health Organization (PAHO) confirmed the autochthonous transmission of CHIKV in the Americas, detecting cases of the infection in the island of San Martín^{5,6}. The entry of the virus to this continent has led to its spread to different areas of the Americas, mainly countries in the

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Figure 1. Enzootic and epidemic cycles of chikungunya virus in nature. In the enzootic cycle, the virus maintains itself in nature in wild primates and is transmitted by *Aedes furcifer* and *A. taylori* mosquitoes. In this cycle in the wild, infections with human hosts occur sporadically, are local and in sylvatic environments. In the endemic cycle, the transmitting species are *A. albopictus* and *A. aegypti*. Infections occur in humans in semi-urban environments, with the mosquito-human-mosquito route leading to large epidemics. The mutation of A226V has favored the adaptation of the vector¹⁵.



central region⁷. The A226V mutation in the E1 protein of the virus appears to be directly responsible for the important increase in infectivity of CHIKV in *Aedes albopictus* mosquitoes, an event that led to the most efficient viral dissemination known in humans⁸ (Fig. 1).

In Mexico, in late 2014 during an outbreak of febrile illness in Chiapas, 79% of analyzed samples tested positive for CHIKV⁹ and *A. aegypti* was identified as its primary vector¹⁰. In 2015, 11,577 cases were confirmed, with four deaths, mostly in the south-central states of Mexico (DGE/InDRE, 2016; OPS/OMS, 2016). An extensive sequence analysis of autochthonous Mexican virus isolates showed that these belonged to the Asian genotype¹¹. In Yucatan, Mexico, CHIKV from the Asian lineage was also isolated from febrile individuals, one case in coinfection with DENV-1¹². An extensive discussion on the pattern of transmission

for CHIKV in the Americas and its association with poor control measures has been published¹³.

GENERAL ASPECTS AND VIRUS REPLICATION

The virus belongs to the *Alphavirus* genus in the *Togaviridae* family, and is made up of an icosahedral capsid and a membrane envelope. The size of the virion varies from 60 to 70 nm. Its genome consists of a single-stranded, positive-sense RNA molecule of approximately 12 Kb. The genome consists of two open reading frames (ORF) that codify for nine proteins: five structural proteins (SP), including the capsid (C), the membrane (6k), and three envelope proteins (E1, E2, E3); and four non-structural proteins (NSP), including NS1, NS2, NS3 and NS4¹⁴.

There are three genotypes of CHIKV, named after the geographic region in which they were first detected: the West African genotype, the Asian genotype, and the East/Central/South African genotype (ECSA). To date, no serotypes of CHIKV have been found, meaning that all genotypes are antigenically similar^{15,16}.

The name of the virus and disease comes from the Bantu language of the Makonde people, meaning “that which bends up”, referring to the curved posture of people with the disease, resulting from the painful arthralgia produced by an infection with CHIKV^{17,18}. CHIKV is capable of propagating in an ample variety of the infected cells of hosts, which include humans, monkeys, birds, cattle, and rodents. In humans, the surface glycoprotein E2¹⁹ of the virion interacts with susceptible host cells that present receptors to this protein on their surface.

One of the proposed receptors for this virus is prohibitin-1 (PHB-1). However, not much is known about the participation of this protein in viral pathogenesis²⁰. The PHB-1 is a protein of 32 kDa involved in the regulation of proliferation and apoptosis, among other functions. This protein is normally located in the mitochondria and its expression is affected by high levels of interferon (IFN)- γ and tumor necrosis factor (TNF)- α . It is also known that the exogenous expression of PHB-1 reduces basal autophagy as well as that induced by TNF- α ²¹.

After the interaction between the viral particle and the host-cell receptor, the former is internalized in the cell through endocytosis. Inside the endosome, there is a reduction in pH due to the pumping of H⁺ ions, which leads to a conformational reorganization of the E1-E2 heterodimer. In the glycoprotein E1, domain II is exposed and fusion is carried out between the membrane of the endosome and that of the viral particle, thus releasing the nucleocapsid of the viral genome¹³. The interaction of the largest RNA subunit 60S with the proteins of the nucleocapsid signals its decoupling, thus releasing the genome. This is a highly conserved mechanism among *Alphavirus*¹⁴.

Once the viral genome (49S) is released in the host cell, the translation of the first ORF is carried out, this codifying for the synthesis of non-structural proteins (NSP1, NSP2, NSP3, NSP4) by means of the production of a polyprotein named P1234. The latter is processed proteolytically, generating the complex of viral replication (Fig. 2). NSP1 participates in the synthesis

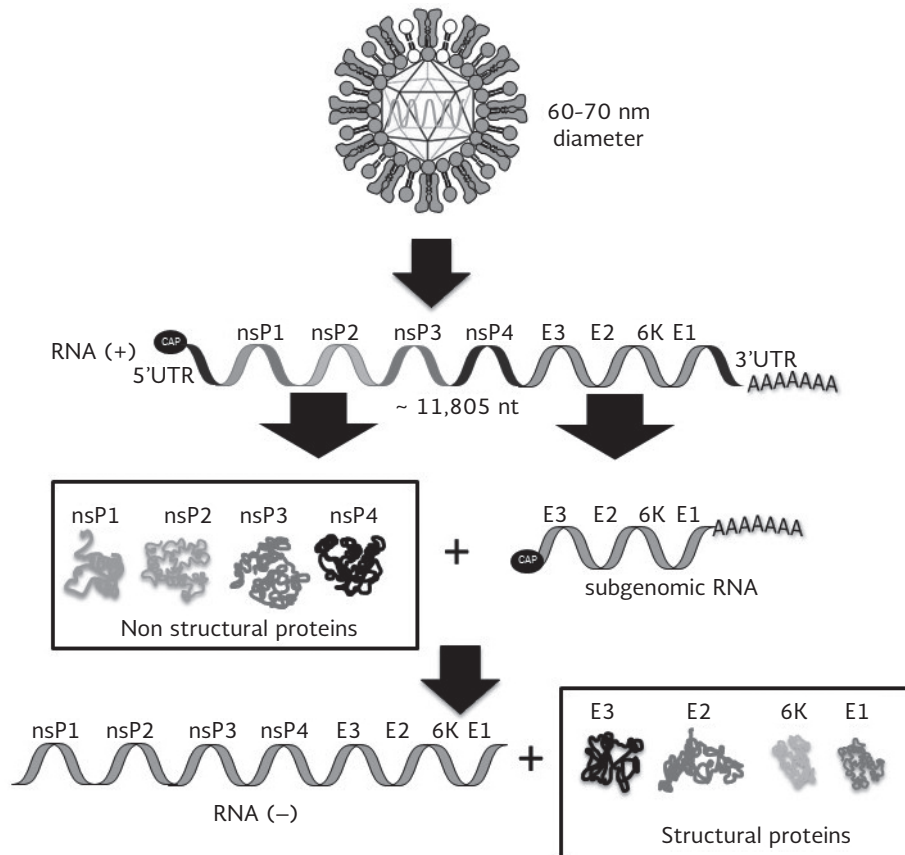
of the complementary chain of viral RNA as well as in the methylation and binding of CAP to the 5' extreme of the viral genome. NSP2 presents activity of RNA helicase, RNA triphosphate and proteinase, and inhibits the transcription of cellular mRNAs. NSP3 participates in the production of the negative chain of viral RNA. NSP4 is the polymerase RNA dependent on DNA².

After the first ORF is translated, the complementary chain of viral RNA (negative chain) is produced. This serves as a template for the transcription of the second ORF known as subgenomic RNA 26S. The translation of this ORF gives rise to the generation of the structural proteins, C, E3, E2, 6k and E1. The C and 6k proteins are accumulated in the cellular cytoplasm to form new nucleocapsids, which are assembled with a copy of the viral genome by means of proteolytic cuts.

Proteins E3, E2, and E1 undergo post-translation processing, being glycosylated in the endoplasmic reticulum. They are then sent to the Golgi apparatus to be later transported in vesicles to the cell membrane. When the nucleocapsid interacts with the accumulated glycoproteins in the cell membrane, the viral particles undergo a maturation process by acquiring the membrane envelope, and then finally are released by means of exocytosis¹⁷. It is important to mention that whereas the synthesis and translation of subgenomic 26CC RNS remains constant, the synthesis of the NSPs decreases and the transcription of genomic RNA 49S increases to the extent that the infection progresses inside the cell^{14,19}.

Once a mosquito infected with CHIKV bites a healthy individual, the virus first multiplies in the fibroblasts located in the epithelium, and then, through infected macrophages, it disseminates to the lymph nodes. Additionally, the virus disseminates to the liver and joints through the blood flow that transports the virions produced¹⁷. The virus continues to propagate itself, evidenced by a viral load of up to 10⁸ PFU/ml (viremia phase) that has been detected at these sites²². Later, it disseminates to the entire organism, thus initiating the signs and symptoms of the disease. The incubation period varies from 2-12 days¹⁸. The active infection only occurs in cells permissive to CHIKV. In mice, these cells primarily consist of muscle, joints, and skin fibroblasts. However, they have also been found in epithelial and endothelial cells of different organs, such as the liver, spleen, and brain¹⁷.

Figure 2. General structure of chikungunya virus as well as genomic configuration and proteins expressed. The particles of this *Alphavirus* are 60-70 nm in diameter, are wrapped in an envelope, and contain a genome of approximately 11,805 nts. Initially, non-structural proteins are expressed and subgenomic RNA is generated. This gives rise in a second phase of transcription to structural viral proteins and the RNA template for copying the genomes of the viral progeny.



CHIKV induces a self-limiting disease, as it is eventually eliminated from the organism. The mechanisms of the innate immune response, mainly the production of IFN- α/β ¹⁷, and later those of adaptive immunity, the IgM and IgG antibodies, circumscribe the infection and finally control and eliminate it from the organism^{23,24}.

TRANSMISSION OF CHIKV

CHIKV is transmitted to humans through a bite from a female *Aedes* mosquito, especially two species¹⁷, *A. aegypti* and *A. albopictus*. The former is considered the primary vector in the transmission of the disease since it is commonly associated with human habitat. However, during the outbreak of this virus on La Réunion Island in 2006, as well as in Africa and India, *A. albopictus* was also identified as an efficient vector, particularly from the ECSA genotype. This event was attributed to adaptive mutations in the genome of

CHIKV within the coding genes for two SPs, E1 and E2¹⁶, mainly involving the mutation of A226V in the E1 protein. This change favors replication within the *A. albopictus* vector and the efficiency of the interaction of the virus with the host cell receptor¹⁵.

The virus circulates in nature through two cycles of transmission: the sylvatic (enzootic) cycle and the urban (epidemic) cycle, which are found in distinct geographic areas²⁴. In Africa, CHIKV is maintained through the sylvatic cycle, in which non-human primates are the primary reservoirs. This situation leads to frequent outbreaks in urban areas¹⁵. Typically, these outbreaks coincide with the season of intense rain and the corresponding increase in mosquito population density. The species of mosquitoes that transmit CHIKV in Africa have been identified as *A. furcifer*, *A. taylori*, *A. vittatus*, *A. fulgens*, *A. luteocephalus*, *A. dalzieli*, *A. vigilax*, *A. camptorhynchites* and *A. africanus*. *Culex annulirostris* and *Mansonia uniformis* are also considered competent vectors²⁵.

As a consequence of population movements from endemic zones of Africa (e.g. Kenya and Comoro islands) to territory where the vector but not the virus previously existed, CHIKV has become an emerging virus. During the years 2005 and 2006, there were multiple outbreaks in India and the islands of the Indian Ocean, including La Réunion Island, in urban cycles of viral transmission^{1,2,4}. In these cycles, humans in urban zones constitute the main host and the virus disperses successfully by means of human-mosquito-human transmission. The species that participate in this transmission are *A. aegypti* and *A. albopictus*, as previously mentioned, leading to massive outbreaks²⁴.

Once the mosquito bites an infected host, the virus arrives to the intestinal tract of the insect, multiplies in the vector's cells, and disseminates to the salivary glands. Then, the mosquito can transmit the virus in each feeding for the rest of its lifetime. To date, there has been no consistent evidence of transovarial transmission in the vectors²⁶.

CLINICAL PRESENTATIONS

With a mosquito bite, viral particles are deposited in the skin, from where they eventually reach the lymph nodes and then the blood flow. They are then distributed to target organs that include joints, muscle, and skin (Fig. 3). With less frequency, the virus can affect the liver or cause encephalopathy, encephalitis, myocarditis, and heart blockage²⁷. Although 5-15% of individuals suffer from asymptomatic infection²⁶, the rest generally develop symptoms that progress from the acute to the chronic phase²⁵.

Acute phase

The acute phase lasts 3-10 days and is characterized by the abrupt presentation of fever ($\geq 39^\circ\text{C}$). Other symptoms follow during the next few days²⁶. Joint pain is reported in 87-98% of the cases. These two symptoms are the most characteristic of this disease.

Joint pain exists in more than one joint and is usually bilateral (symmetrical), occurring mainly in the peripheral joints (knees, ankles, hands, wrists)²⁸. There is myalgia in 46-72% of the cases, affecting arms, thighs, and calves²⁵. This clinical picture is called chikungunya fever (CHIKF) and tends to be limiting and

even disabling in relation to the normal physical activity of individuals^{24,29}.

The exanthema that appears in 40-50% of cases is of the petechial or maculopapular type, mainly affecting the limbs, trunk, and face, and can cause pruritus. The lesions are transitory and generally appear 2-5 days after disease onset²⁹. Other symptoms that are less common are diarrhea, vomiting, edema on limbs, bleeding, otitis, and ocular disease (especially anterior uveitis)³⁰⁻³². Some of the more severe manifestations, which are infrequently found, include neurological diseases such as meningoencephalitis, the Guillain-Barré syndrome, myocarditis, and multiorgan failure. The latter can be fatal, particularly in neonates and older adults with comorbidity^{33,34}. Laboratory findings from persons in the acute stage of the disease have evidenced lymphopenia (500-1000 cells/mm³) and moderate thrombocytopenia (100,000-150,000 cells/mm³). Other uncommon abnormalities include leukopenia, elevation of hepatic enzyme levels, anemia, elevated creatinine level, and hypocalcemia¹⁸.

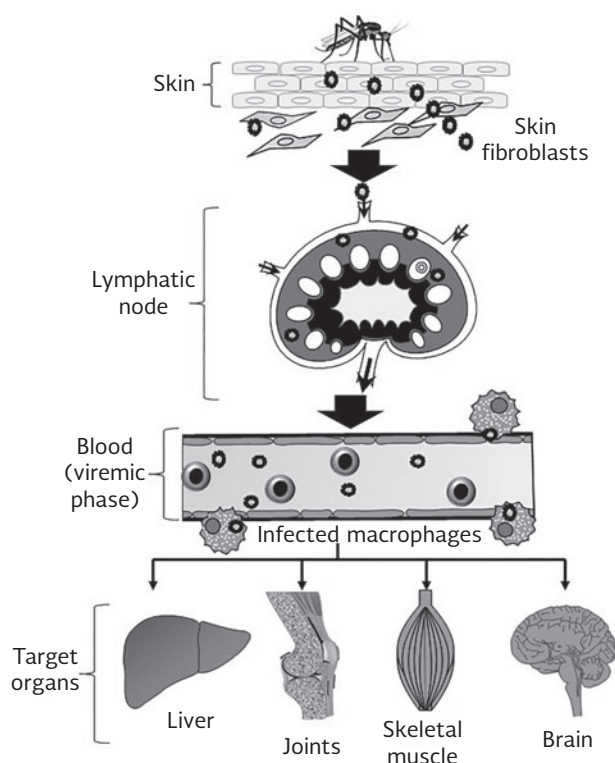
In neonates, CHIKF can be accompanied by convulsions, peripheral cyanosis, podalic edema, and epithelial vesicular lesions that eventually dry and scale^{35,36}. This disease is usually considered benign in children³⁷. However, neurological manifestations have been reported that include febrile convulsions, meningeal syndrome, acute encephalopathy, diplopia, aphasia, acute disseminated encephalomyelitis, and encephalitis^{35,38-40}. The development of chronic arthralgia and exacerbation of underlying medical conditions is unusual²⁶.

Chronic phase

After the diverse outbreaks and epidemics of chikungunya occurred in different regions of Africa, Asia and Europe, follow-up studies generated sufficient evidence to indicate that the infection with this virus can induce chronic rheumatic diseases. In adults, these conditions can last months or even years after the infection, while they are less common in children^{19,25,37}.

The musculoskeletal disorders include arthralgia, inflammatory arthritis, polyarthralgia, tenosynovitis, enthesitis, and exacerbation of existing rheumatic disease^{29,41-46}. Other less common symptoms include neuropathy, cerebral disorder, neurosensory deficiency, burning mouth syndrome, paresthesia, cubital

Figure 3. Cells and target organs of chikungunya virus in humans. The virus is deposited on the skin during a mosquito bite, where it has been reported that cells like fibroblasts capture it. Later, the virus reaches the peripheral lymph nodes and arrives to the blood flow, causing viremia. The macrophages are often found infected by the virus and the main target organs are joints, skeletal muscle, liver and brain¹⁷.



tunnel syndrome, gastrointestinal disorders, exanthema, pruritus, bursitis, and synovitis^{28,36,44,47-49}.

Infection with CHIKV has been associated with destructive rheumatoid arthritis, which is similar to rheumatoid arthritis and induces a similar inflammatory response, although the laboratory tests are negative for the rheumatoid factor and the antibodies for cyclic citrullinated peptides (CCP)^{26,44,50}. Radiographic studies of some of these individuals show bone erosion, swollen joints and synovitis^{44,51}. The symptoms can be recurrent or chronic and can affect multiple joints (mainly those previously injured), which significantly diminishes the quality of life of the patient both physically (regarding function, pain and general health) and psychological^{18,52,53}.

The development of arthralgia may be due to continuous inflammation in joints in response to viral antigens, evidenced by the fact that viral RNA has been detected in perivascular macrophages⁵⁴. Risk factors for the development of chronic arthralgia identified to date

include advanced age, long duration of the acute phase of CHIKV⁵³ with more than six joints manifesting pain¹⁹, existing joint disease³⁹, and a delayed adaptive immune response (mainly IgG neutralizing antibodies)⁵⁵.

Congenital infection

Pregnant women with detectable viremia a few days before childbirth can transmit CHIKV to the newborn, which can result in a severe neonatal disease, generally encephalopathy followed by neurodisability^{3,56}. Other signs and symptoms observed in newborns are convulsions, thrombocytopenia, hypotension, ventricular dysfunction, pericarditis, hyperechoic coronary arteries, parenchymal hemorrhage, and cytotoxic edema. It has been demonstrated that maternal-fetal transmission occurs due to contact of the product with infected maternal blood during childbirth⁵⁷.

There is no specific antiviral drug treatment against CHIKV. Nonetheless, symptomatic treatment includes acetaminophen to relieve the fever and nonsteroid anti-inflammatory drugs (e.g. paracetamol) for polyarthralgia¹⁸.

PATHOGENESIS OF CHIKUNGUNYA VIRUS DISEASE

Alphavirus, including CHIKV, cause severe forms of diseases such as chronic and highly disabling arthralgia/arthritis. Chronic symptomatology is observed with greater frequency in persons over 60 years of age that have very high viremia ($> 10^{10}$ PFU/ml) during the acute phase of the disease²⁷. The infection is initially very rapid and the virus is typically eliminated 5-7 days after onset of the fever. During the infection there is a robust and rapid activation of dendritic, NK/CD4+ and CD8+ cells. Persistently high levels of IL-12 are mainly found in individuals with chronic symptoms, mostly in persons with rheumatoid arthritis, as inflammation provoked by CHIKV infection can cause bone erosion and severe arthralgia.

The activity of osteoclasts increases, thus promoting bone absorption⁵⁸. Additionally, the activity of osteoblasts is inhibited during infection, which leads to the inhibition of bone formation⁵⁹. The differentiation of osteoclasts from their precursors requires participation of the inductor cytokine RANKL (receptor activator for

the nuclear factor κ B ligand)⁶⁰ and the interaction with its RANK receptor⁶¹. The activity of RANKL is inhibited by osteoprotegerin (OPG), a natural receptor that functions as a decoy for RANKL and blocks its interaction with RANK⁶² (Fig. 4). The concentrations of OPG and RANKL are maintained in a proportion that regulates the process of osteoclastogenesis⁶³. When the concentration of RANKL rises, the production of osteoclasts increases, which leads to greater bone absorption.

Infections by *Alphavirus* share some common characteristics with other diseases such as rheumatoid arthritis⁶⁴. Replication of CHIKV inside cells in biopsies of synovial tissues has been clearly demonstrated. Pulkia, et al. showed the direct infection of primary synoviocytes with CHIKV. These infected cells attract monocytes/macrophages and induce the formation of osteoclasts⁵⁶. Primary cultures of osteoblasts have great susceptibility to infection with CHIKV, an infection that promotes the formation of osteoclasts and the loss of bone tissue⁶⁵.

The innate immune system acts as the front line against an invasion by *Alphavirus*. There is increasing evidence of an immunopathogenesis due to the over-activation of the immune system by these viruses⁶⁶. Apparently, the infiltration of macrophages to the joints determines the severity and persistence of the infection. The persistent infection of perivascular synovial macrophages by CHIKV and the constitutive infiltrate of CD14+ monocytes to the interior of the synovial cavity have been observed 18 months after the onset of symptoms in individuals infected with CHIKV⁵⁴. During such an infection, a large number of cytokines, chemokines, and growth factors are stimulated in the plasma of these persons. Accordingly, a variety of interleukins have been detected: IL-1 β , IL-5, IL-6, IL-7, IL-10, IL-15 and IL-17. Ligands of chemokines found include CCL-3, CCL-4, CCL-2 (MCP-1) and CCL-5 (RANTES). Finally, granulocyte-macrophage colony-stimulating factor (GM-CSF) has been found^{67,68}.

In synovial liquid of infected individuals, there are important levels of CCL-2, IL-16 and IL-8⁶³, as well as elevated concentrations of RANKL, which could directly lead to bone loss⁶⁹. These observations correlate with findings in animal models, such as mice and primates, in which it has been demonstrated that IL-6 promotes the formation and liberation of RANKL. Hence, a probable strategy for controlling the infection is the use of anti-IL-6 antibodies⁷⁰.

In the infection by *Alphavirus*, an increase has been found in the activation of T CD4+ lymphocytes^{54,71}, which differentiate to Th17 and infiltrate the synovial space, possibly promoting local inflammation⁷².

FINAL REMARKS

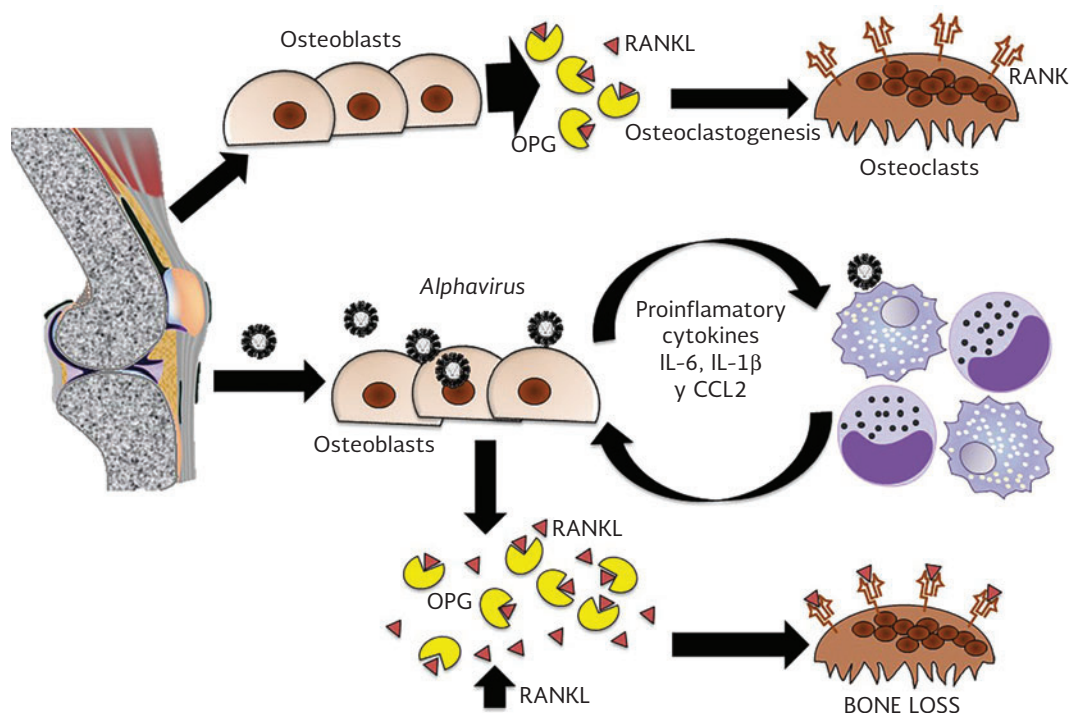
It is important to consider that diverse diseases have a clinical presentation similar to that observed with a CHIKV infection. There are a great number of clinical conditions that must be ruled out to confirm the diagnosis and these include: leptospirosis, malaria, infections with other alphaviruses, and other viral entities that cause exanthema.

Even though CHIKF is characterized by fever with polyarthralgia, these indicators are unspecific. Moreover, the infection can take place in a subclinical form or coexist with other infections like dengue⁶⁵. Therefore, diseases that should be considered in a differential diagnosis vary in relation to the relevant epidemiological characteristics, such as place of residence, travel to endemic zones, and exposure to mosquito bites⁶.

In the differential diagnosis of CHIKV in Mexico, it is of great importance to consider DENV, due to a high similarity between the clinical manifestations of these two infections, as well as the high incidence of the latter agent in this country. Symptoms like myalgia, arthralgia, and exanthema are more associated with CHIKV, while thrombocytopenia may be more indicative of dengue^{40,67}. Despite the broad similarity between both diseases, CHIKF has a more acute onset and lasts a shorter time; additionally, maculopapular exanthema is more frequent and joint pain is more intense/localized and highly incapacitating.

In the absence of a vaccine or antiviral drug on the market, vector control is the only currently available strategy for controlling CHIKV. The previous experience with DENV control efforts is not encouraging. The virus spread throughout the Americas was imminent due to inefficient vector control and the emergence of resistance to insecticides in mosquitoes. Nevertheless, it is essential to continue efforts to improve epidemiological surveillance of CHIKV and other viruses by strengthening the health services and facilities at different levels, mainly for primary care, to detect new cases of the virus in a timely manner. Moreover,

Figure 4. Infection with *Alphavirus* and its effect on osteoclastogenesis through RANKL. The osteoclasts differentiate from their precursors due to the effect of RANKL and the interaction with its receptor RANK. The binding of RANKL to RANK is inhibited by the decoy protein osteoprotegerin, which allows the maturing of osteoclasts. The changes that occur during *Alphavirus* infections induce an increase in RANKL that is not captured by osteoprotegerin, interfering with osteoclastogenesis⁶⁵. CCL: chemokine ligand; IL: interleukin; OPG: osteoprotegerin; RANKL: receptor activator for the nuclear factor κ B ligand.



efforts to develop novel treatment strategies to fight this severe viral infection should continue.

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