

Immunology of Alzheimer's disease

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Abstract

Alzheimer's disease (AD) is a multifactorial neurodegenerative pathology. Neuroinflammation is an early event of the presymptomatic stages in AD and contributes to its progression. We review the participation of astrocytes, microglia and blood-brain barrier cells, the mechanisms of cell death and the inflammatory factors such as chemokines, interferons and Toll-like receptors involved in progression and perpetuation of AD. Some of its prognostic and therapeutic possibilities are also mentioned. Identifying the different actors involved in inflammation and the main mechanisms of damage might allow the development of preventive strategies and treatments to fight against this devastating disease.

Key words: Immunology. Alzheimer's disease. Microglia. Innate immunity. Lymphocytes.

Inmunología de la enfermedad de Alzheimer

Resumen

La enfermedad de Alzheimer (EA) es una patología neurodegenerativa multifactorial. La neuroinflamación es un evento temprano de las etapas presintomáticas en la EA y contribuye a su progresión. En este trabajo revisamos la participación de astrocitos, microglia y células de la barrera hematoencefálica, los mecanismos de muerte celular y los factores inflamatorios como las quimiocinas, los interferones y los receptores tipo Toll que participan en la progresión y la perpetuación de esta enfermedad. También se mencionan algunas de sus posibilidades pronósticas y terapéuticas. La identificación de los diferentes actores involucrados en la inflamación y de los principales mecanismos de daño podrían permitir el desarrollo de estrategias y tratamientos preventivos para combatir esta enfermedad devastadora.

Palabras clave: Inmunología. Enfermedad de Alzheimer. Microglia. Inmunidad innata. Linfocitos.

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Introduction

Alzheimer's disease (AD) is the most common dementia. It is characterized by abnormal protein aggregates of tau protein (tubulin-associated) and β -amyloid ($A\beta$). Tau aggregates acting as “seeds” may propagate pathology by spreading from cell to cell in a “prion-like” manner¹. It occurs in two forms, early onset, that is, genetically determined and late onset that is more frequent and multifactorial². Late-onset AD is genetically complex with 56-79% heritability³. Regardless of the initial trigger that initiates the abnormal aggregation of proteins, there seems to be an inflammatory background that contributes to the perpetuation of neuronal damage². The importance of the immune response in AD has been evidenced with genomic studies in late-onset AD. These studies have shown at least four functional pathways of susceptibility to AD, the immune response, the regulation of endocytosis, the transport of cholesterol, and the ubiquitination of proteins⁴. The aim of this work is to review the most important immunological aspects in AD and its possible therapeutic implications.

Development

The immune system identifies foreign elements (i.e., different from the self) to mount a defense response against them. Natural or innate immunity is not specific and does not require an external challenge to be involved; acquired immunity is specific and keeps the memory of previous challenges. The immune system seems to contribute to the perpetuation of damage in Alzheimer's disease (AD). Some recent research suggests that the manipulation of some of these participants in the immune response such as microglia and cytokines could have beneficial therapeutic effects. Inflammation in AD involves both the innate and the acquired immune system⁵.

Participating cells

Astrocytes

Astrocytes are the most abundant glial cells. They contribute to the support of the neurons, but it is now known that they can perform other functions including providing the biochemical support of the endothelial cells that are part of the blood–brain barrier (BBB); they participate in the maintenance of the extracellular ionic balance and the repair and healing process of the brain

parenchyma. The astrocytes can clean detritus by phagocytosis and support neuronal nutrition; but also, they are mediators of inflammation and are involved in the formation of reactive oxygen species. They have multiple roles in the development of AD: astrocytes degrade $A\beta$ without the need for opsonins or cytokines⁶. They contribute to the clearance of β -amyloid protein ($A\beta$) and limit brain inflammation. If they dysfunction, they can also participate in neurodegeneration, releasing toxins, and altering basic metabolic pathways⁷.

Astrocytes, as well as monocytes/macrophages and endothelial cells, secrete, monocyte chemoattractant protein-1 (MCP-1); this secretion is mediated by the stimulation of $A\beta$ and depends on the physical contact between monocytes and astrocytes. MCP-1 facilitates the formation of $A\beta$ oligomers in microglia. The concentration of MCP-1 in serum and cerebrospinal fluid (CSF) is elevated in patients with AD; the higher plasma concentration of MCP-1, the greater the severity of the disease, and the greater the cognitive deterioration⁸.

In cell cultures of astrocytes, it has been shown that it is possible to inhibit inflammation induced by $A\beta$ by pyrrolidine dithiocarbamate acid, indicating that nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) is probably involved in the inflammatory process⁹. Resveratrol reduces inflammation in rat astrocytes probably by inhibiting NF- κ B⁹.

Interleukin 8 (IL-8) was the first chemokine identified in the brain. Astrocytes, neurons, and microglia are capable of producing IL-8 *in vitro*, whereas IL-8 receptor or CXCR2 is located in the neuritic portion of the plaques around $A\beta$ deposits in tissues of patients with AD. Type 2 diabetes *mellitus* is a risk factor for dementia. It has been shown that hyperglycemia increases the expression of IL-6 mRNA and the astrocytic secretion of IL-6 and IL-8, contributing to astrocyte-mediated neuroinflammation¹⁰.

Microglia

Microglia are resident myeloid cells of the brain capable of recognizing endogenous and exogenous insults and initiating an immune response. They promote phagocytic cleaning and provide trophic support to ensure tissue repair and maintain homeostasis. In addition, they participate actively in the remodeling of synapses with the release of brain-derived neurotrophic factor (BDNF) that contributes to the formation of memory circuits.¹¹ Microglial cells originate from the yolk sac during primitive hematopoiesis¹². Their differentiation need the PU.1 transcription factor and the

regulatory factor of interferon 8 or IRF8; to survive, they also require the signaling of the colony stimulating factor 1 receptor (CSF1R)¹³. The release of IL-10 by the microglia causes an increase in the number of dendritic spines. AD is associated with apolipoprotein E ϵ 4 (APOE4) while APOE2 has a protective nature. Both astrocytes and microglia express APOE under the control of nuclear hormone receptors. In AD, the inflammatory activity of the microglia is increased. The microglial cells mount an immune response against the misfolded A β protein. The deposit of A β induces inflammatory changes in the brain parenchyma, and activation of the microglia has been demonstrated with an increase in the levels of pro-inflammatory cytokines in the regions characteristically most affected in the disease¹⁴. If the problem is not solved and the stimulus continues, the activity of the microglia deviates from its physiological and beneficial functions. To recognize aggressions, the microglia have different types of receptors, some designed to identify molecular patterns associated with pathogens (PAMPS) and damage, known as PAMPS/DAMPS. It also presents the class 1-scavenger receptor, CD36 (Cluster designation/differentiation), CD14, integrin α 6 β 1, CD47 (integrin-associated protein), and Toll-like receptors (TLR2, TLR4, TLR6, and TLR9)¹⁴. The amyloid precursor protein (APP) is an integral membrane protein expressed in many tissues and concentrated in the neuronal synapse where it probably has a trophic and regulatory function with both extracellular and intracellular interaction with different signal transduction pathways¹⁵. It seems to participate in the development of the neural stem cell, in neuronal survival, and the growth and repair of neurites. APP is cut by some enzymes, giving rise to peptides, some of which are released outside the cell¹⁶. Two of these fragments, or peptides that leave the cell are:

- Soluble sAPP that promotes the growth of nerve cells and can play an important role in the formation of neurons, both before and after birth.
- A β , a peptide of 36-43 amino acids that activates kinases. Oligomeric A β 42 enhances Ras (contraction of Rat Sarcoma)/ERK signaling cascade and glycogen synthase kinase-3 (GSK-3) activation. Both ERK and GSK-3 induce hyperphosphorylation of tau and APP at Thr668¹⁷ and could be related to cholesterol metabolism. In fact, inhibition of cholesterol biosynthesis reduces γ -secretase activity and A β generation¹⁸. It is also believed that it may have some antimicrobial activity (it intervenes in the inflammatory action)¹⁶.

APP undergoes two consecutive cuts by two membrane-bound proteases. It is initially cut by BACE1 (β -site APP-cleaving enzyme 1). Subsequently, a second cut is made by the secretase γ (gamma) complex within the transmembrane region of the APP resulting in fragments of 37-42 amino acids at the C-terminal end called A β . In particular, the A β of 42 amino acids (A β ₁₋₄₂) has a tendency to form soluble oligomers and fibrils¹⁶.

The binding of A β to CD36, TLR4, and TLR6 activates microglia and with it, the production of pro-inflammatory cytokines and chemokines. Microglial cells ingest A β fibrils that enter the endosomal/lysosomal pathway. However, these fibrils are resistant to enzymatic degradation unlike soluble A β that is degraded by extracellular proteases. Preliminary studies have demonstrated that CNP520, a BACE-1 inhibitor, reduced the amount of A β in CSF and in the brain parenchyma of healthy rats and dogs, and the deposit of A β in the plaques of transgenic-APP mice. In adults over 60, it was well tolerated and it reduced A β concentration in the CSF. These studies are still in progress¹⁹.

Microglia are able to release the insulin-degrading enzyme (IDE) that degrades insulin, amylin, and A β . IDE seems to be the main regulator of A β levels in microglia and neurons. In animal models, the homozygous deletion of the IDE gene results in a 50% decrease in the degradation of A β and in a similar deficit in the breakdown of insulin in the liver with an accumulation of A β in the brain. A meta-analysis demonstrated recently that AD patients have lower protein levels of IDE in comparison with controls (mRNA levels were not systematically lower)²⁰.

It has also been shown that IDE degrades the APP intracellular domain (AICD). IDE regulates the levels of non-phosphorylated AICD. It seems that phosphorylation protects AICD from its breakdown by IDE. The IDE could be a therapeutic target in AD²¹.

ApoE gene, which codes for ApoE, the most important genetic risk for AD seems to have an immunomodulatory function. This function is related to the activation of triggering receptor expressed on myeloid cells2 (TREM2) expressed by microglia²².

Mutations in the extracellular domain of TREM2 confer an elevated risk of developing late-onset AD. A risk allele (R47H) of TREM2 had an effect similar to ApoE4 (odds ratio 2.90-5.05) in a Colombian population²³. TREM2 is a receptor expressed in macrophages including microglia in the brain. TREM2 participates in the survival and proliferation of microglia, chemotaxis, and phagocytosis²³. In murine models of AD, the loss of TREM2 causes increased deposits of A β in the

hippocampus due to a dysfunctional response of the microglia to A β suggesting that TREM2 facilitates the clearance of A β by these cells²⁴.

The microglial response mediated by TREM2/DAP12 (DNA activation protein of 12kDa) limits the diffusion and toxicity of the amyloid plaques forming a protective barrier. TREM2 propagates its signal through the adapter protein DAP12, which, in turn, activates several signaling pathways including Spleen tyrosine kinase, phosphoinositide 3-kinase, and the mitogen-activated protein kinase (MAPK) which culminates in increased phagocytosis and an anti-inflammatory profile in the microglia²⁴. On the other hand, dendritic cells deficient in TREM2 secrete more tumor necrosis factor- α (TNF- α), IL6 and IL-12 compared to wild type, especially when activated by lipopolysaccharides; this suggests that there may be a shift toward a more inflammatory profile in the absence of TREM2²⁵.

The loss of TREM2 in microglia confers an increased risk of developing late-onset AD and is associated with loss of endothelial homeostasis²⁵.

In AD, there is over-regulation of TREM2 that seems to serve as a compensatory response to A β_{1-42} and protects against the progression of the disease by modulating the functions of microglia. TREM2 promotes the survival of microglia by activating the Wnt/ β -catenin signaling pathway. The manipulation of TREM2/Wnt/ β -catenin may be a therapeutic target in AD²⁶.

It is known that all isoforms of apoE are an agonist of TREM2²⁷. APOE3 maintains lipid homeostasis and has a protective cardiovascular effect. APOE2 is associated with dysbetalipoproteinemia and APOE4 is a risk factor for AD²⁷. Microglial cells are able to produce nitric oxide (NO), TNF- α , and IL-1 β and promote the generation of antibodies against A β , stimulating the clearance of amyloid plaques. The soluble A β oligomers and the A β fibrils have the ability to bind to receptors expressed in the microglia, such as CD14, CD36, CD47 integrin α 6 β 1, the Class A eliminador receptor, the receptor for advanced glycation end products (RAGE), and TLR. In macrophages and microglia, classical M1 activation is characterized by a pro-inflammatory profile of cytokines including TNF- α , interleukins 1 (IL-1), 6 (IL-6), 12, and 18, and it is accompanied by a deficient phagocytic capacity, while the M2 profile is characterized by secretion of anti-inflammatory cytokines IL-4, IL-10, and IL-13 and transforming growth factor-beta (TGF- β) and by a high phagocytic capacity without NO production. A third phenotype is a deactivated state associated with corticosteroids or with TGF- β . *In vitro*,

bipolar/rod-shaped microglia are highly proliferative, express various M1/M2 markers and are quickly transformed into amoeboid microglia within 30 min of lipopolysaccharide treatment, leading to the upregulation of pro-inflammatory cytokine gene expression and the activation of Jak/STAT *signaling pathway* (Janus kinase-signal transducer and activator of transcription)²⁸. Markers of microglial phenotypes in human brains are still limited; the most widely used marker to describe activated microglia, particularly in diseased brains, has been HLA-DR, or major histocompatibility complex II protein. Ionized calcium binding adaptor molecule-1 (IBA1) and CD68 are generic markers of microglia and recruited monocytes²⁹. In patients with AD, Stages V-VI of Braak, degeneration of the microglia has been observed, especially in the dentate gyrus, probably due to the accumulation of toxic tau-soluble species³⁰. There are agents capable of increasing phagocytosis of A β in phagocytic cells, such as Lipoxin A4 (LXA4), an endogenous lipid mediator with anti-inflammatory properties. It has been shown in mice that the administration of aspirin (15 μ g/kg) twice a day, through the activation of LXA4, reduces the activation of NF- κ B and the levels of pro-inflammatory cytokines, and produces an increase in IL-10 with anti-inflammatory action and in TGF- β . This was translated to the brain level in the recruitment of microglia with a phenotype characterized by the upregulation of YM1 lectin protein, and arginase 1 and low-regulation of the inducible synthase expression of NO. With this phenotype, the microglia presented a better phagocytic function with an efficient clearance of A β , reduction of synaptotoxicity, and improvement of cognition³¹.

In transgenic mice bearing a mutated gene of tau (p301s MAPT), the elimination of astrocytic and microglial senescent cells (that accumulate p16^{INK4A}), with first-generation senolytics drugs (drugs that cause senescent cells to become susceptible to their own pro-apoptotic microenvironment)³², preserves the cognitive function; a therapy focused on senescent cells (with irreversible arrest of the cell cycle) could be useful in AD³³.

BBB

The BBB protects the central nervous system from the entry of substances that can be potentially harmful and maintains homeostasis and communication between the brain and the peripheral blood. In addition to affecting neurons, astrocytes, and microglia, AD also damages the vascular cells of the neurovascular unit

such as endothelial cells, pericytes, and vascular smooth muscle cells³⁴. The interaction of A β with endothelial cells produces structural and functional changes in the BBB. It is now known that C-reactive protein (CRP), a member of the pentraxin superfamily involved in innate immune response, acts as a direct mediator of inflammatory reactions. The inert circulating pentameric form (pCRP) is transformed into the pro-inflammatory isoform pCRP and finally into the monomeric form (CRPm) in the presence of amyloid beta and activated endothelial cells. It will then contribute to the inflammation in AD³⁵.

A β in CSF is rapidly cleared into the bloodstream. A β in the brain parenchyma is cleared across the BBB through the low-density lipoprotein receptor-related protein-1 (LRP-1)³⁶.

The luminal-to-abluminar transcytosis of A β is mediated by a transporter identified as the receptor for advanced glycation end products (RAGE). RAGE up-regulation has been observed in the CNS microvasculature of humans with AD by histochemical methods in autopsy material³⁷.

Decreased expression of low-density lipoprotein receptor-related to protein 1 (LPR-1) in the outer or abluminar part of cerebral capillary cells and decreased levels of multidrug transporter p-glycoprotein (P-gp) in the luminal plasma membrane of the capillaries lead to a decrease in the flow of A β from the brain to the blood³⁸.

On the other hand, it has been observed that Catalpol, an iridoid glycoside extracted from the root of *Rehmannia glutinosa* Libosch, has a neuroprotective effect in AD. The protective mechanism seems to depend on a decrease in the levels of metalloproteinases (MMPs), MMP-2, MMP-9, and RAGE, as well as an increase in the concentration of proteins of the narrow junctions (zonula occludens-1, occludin, and claudin-5), the LPR-1 and the glycoprotein P, so that the Catalpol could have utility in the early treatment of AD³⁹. The endothelial cells of the BBB can also be damaged by high concentrations of low-density proteins (LDL). Statins decrease the inflammatory effects of oxidized LDL in the microvasculature⁴⁰.

Lymphocytes

AD appears to be a systemic pathology in which some of the dysfunctions found in the brain are present in peripheral tissues. Lymphocytes from patients with AD have an increased susceptibility to death induced by hydrogen peroxide (H₂O₂) that is related to

the severity of dementia and appears to depend on deregulation of the p53 pathway with increased expression of p53⁴¹. Various alterations in the lymphocytes of patients with AD have been described, including a systemic decrease in B and T lymphocytes. The peripheral CD4 + and CD19 + lymphocytes in the early stages of AD show mitochondrial depletion. Lymphocytes T-helper 17 are found in the brain parenchyma of AD and IL-17A is located around the A β deposits. Overexpression of IL-17 improves glucose metabolism, amyloid angiopathy and learning in rats exposed to ozone, a murine model of oxidative stress and decreases soluble A β in the hippocampus and the CSF⁴². More recently, it has been found that micro-RNA Let-7b levels increase with the progression of the disease and this is parallel to the increase in the number of CD4 + T lymphocytes in the CSF. This microRNA correlates positively with the expression of t-Tau and p-Tau and is being studied as a possible biomarker of progression⁴³.

In the late stages of the disease, CD8 + T lymphocytes are increased in number in the hippocampus of subjects with AD compared to subjects without dementia. The numerical density of T-lymphocytes correlates with the tau pathology (AT8 staining)⁴⁴.

It is known that Vitamin D deficiency is a risk factor for cognitive deterioration and that this vitamin is involved in the clearance of A β from the brain. In subjects with mild cognitive impairment, their lymphocytes are more susceptible to oxidative damage, which improves with treatment with Vitamin D for 6 months as well as the plasma concentration of A β and cognitive function⁴⁵.

It has been proposed that the T cell profile may change depending on the stage of the evolution of AD, with an increase in pro-inflammatory activity as the disease progresses. Neurodegeneration, in general, has been linked to an imbalance between effector T cells that release IFN γ or IL17 and T-lymphocytes reg, which leads to a decrease in neuroprotection and increases neuronal damage⁴⁶.

Mechanisms of Cell Death

Apoptosis and necrosis

In the context of AD, although death due to apoptosis seems to prevail, necrosis also contributes to neurodegeneration. In familial Alzheimer's, some genes involved (presenilin 1 and 2) make neurons more susceptible to apoptosis².

On the other hand, greater expression of Bak and Bad pro-apoptotic proteins, activation of caspases and decreased expression of the antiapoptotic gene NCK-associated protein 1 has been demonstrated in affected brains².

Neuronal autophagy

Excessive accumulation of autophagic vacuoles and toxic substances such as misfolded proteins or damaged organelles can lead to cell death by self-destruction. Autophagosomes are frequent in AD. The beclin-1 protein plays an important role in autophagy and is diminished in AD. In fact, patients with AD show accumulation of autophagy markers such as sequestosome 1/p62 (ubiquitin-binding protein) and LC3 (Microtubule-associated protein 1A/1B-light chain 3) and these markers colocalize with the A β marker-6E10 and hyperphosphorylated tau⁴⁷.

The intracellular A β alters the retrograde transport mediated by dynein. The neurons must transport the autophagosomes generated in the distal axons to the soma or neuronal body. This retrograde transport starts with the recruitment of the late endosome complex (LE)-loaded with dynein and with SNAPIN (SNAP-associated protein) after fusion of LE with autophagic vacuoles to form amphisomes. However, in AD, autophagic vacuoles accumulate massively within dystrophic neurites. A β is associated with an increased density of LEs or multivesicular bodies. It has been shown in animal models that Dynein intermediate chain 1, axonemal interacts with A β _ interacts with A β . This binding competitively disrupts the formation of dynein-SNAPIN complexes, and therefore the recruitment of dynein motors to LEs-loaded with SNAPIN (amphisomes), is markedly reduced⁴⁸.

Inflammation

In the development of AD, A β , and Tau protein can act as pro-inflammatory factors. In fact, the deposition of A β in the brain is associated with the activation of astrocytes and microglia. The soluble isoforms of A β are phagocytosed by microglia while the insoluble deposits activate microglia by binding to TLRs. In these cells, A β activates MAPKs and favors the expression of a pro-inflammatory profile with the secretion of cytokines and chemokines that amplify the inflammatory process⁴⁹. The misfolded protein aggregates activate the NLRP3 (cryopyrin) inflammasome. NLRP3 inflammasome is a multiprotein complex expressed in

myeloid cells and has an important role in the activation of caspase-1 by A β ⁵⁰.

Glia maturation factor (GMF) is a regulator of the actin cytoskeleton with a unique role in remodeling actin network architecture; it does not bind actin but instead binds the Arp2/3 complex. GMF catalyzes the debanching of actin filament networks and inhibits actin nucleation by Arp2/3 complex. GMF is also a pro-inflammatory molecule present in glial cells and some neurons. Its over-expression causes inflammation. It is localized and expressed in the neighborhood of A β and tau in the temporal cortex of patients with AD. GMF could be a therapeutic target in AD⁵¹.

Dendritic cells (DC) appear to regulate the entry of T-lymphocytes into the perivascular and leptomeningeal spaces. Their protective properties in AD are related to their ability to clear A β . A β significantly decreases the expression of brain-derived neurotrophic factor (BDNF) in DCs derived from AD patients but not from control subjects, AD-linked dysregulated immune mechanisms lead to dendritic cell-mediated over-activation of inflammation and impaired antigen presentation, thus supporting the idea that immune cell activation could play an important role in AD pathogenesis⁵².

Recently, elevated levels of CSF biomarkers such as heparin and chitin-binding glycoprotein (YKL-40), intercellular adhesion molecule-1, vascular adhesion molecule-1, IL-15, and fms-related tyrosine kinase-1 have been described, both during the preclinical phase and in the dementia phase of AD. These levels correlate with the total concentration of tau in the CSF⁵³.

Chemokines

Chemokines, belonging to the cytokine family, are small proteins that bind to heparin and are chemoattractants; some are pro-inflammatory while others control the migration of cells during development. Chemokines are classified according to their primary protein structure that is based on the number of amino acids that separate two cysteine residues; thus, four groups are recognized, α (CXC), β (CC), γ (CX3C), and δ (C). Chemokine receptors are designated CXCR1-CXCR6, CCR1-CCR11, CX3CR1, and XCR. Chemokines and their receptors represented by MCP-1 (also called chemokine (C-C motif) ligand 2 [CCL²] and its receptor (CCR2) are considered biomarkers to monitor progression in AD since the progression of the disease seems to be related to the expression of chemokines¹⁴.

In studies of clinical follow-up of patients with a cognitive neurological deficit, it has been seen that those with the highest tertile of MCP-1 in CSF showed a significant cognitive decrease and developed dementia in a shorter time than those in the lowest tertile¹⁴.

The chemotactic cytokines stimulate and control the movement of leukocytes from the blood to the tissues. In the context of AD, the most studied chemokine is CCL5 (RANTES) that regulates the expression and secretion of T cells¹⁴. Curcumin increases neuronal survival in the toxicity model induced by N-methyl-d-aspartic acid by inducing the expression of RANTES in astrocytes through the phosphatidylinositol 3-kinase and MAPK pathways⁵⁴.

In AD increased concentrations of CCL5 of astroglial origin have been described in the cerebral microcirculatory system in response to the increase in reactive oxygen species-mediated by cytokines.

It has been described in patients with AD that levels of MCP-1 and IL8 are increased in serum, CSF and parenchyma; on the other hand, levels of fractalkine and stromal cell-derived factor 1 are decreased in serum. Fractalkine (CX3CL1) is made by neurons, and its receptor (CX3CR1) is expressed by microglia; therefore, fractalkine/receptor interactions are a neuron–microglial signaling system⁵⁵.

MIP-1 and RANTES levels are elevated in the brain parenchyma. MCP-1, IL-6, and IL-8 are over-expressed in brain tissue in AD. Immunohistochemical studies have confirmed the increase and localization of these three factors in neurons⁵⁶, whereas in astrocytes MCP-1 and IL-6 were detected. MCP-1 and IL-8 have been observed in senile plaques.

Interferons (IFN)

IFN are cytokines made up of glycoproteins used for communication between cells; they activate the immune system in case of aggression. They are divided into three classes: types I, II, and III. Type I IFN in humans is IFN- α , IFN- β , IFN- ϵ , IFN- κ , and IFN- ω ; produced by fibroblasts and monocytes; they are activated in viral infections. Type II or IFN- γ in humans is activated by IL-12 and produced by helper T-lymphocytes type 1 (Th1) and natural killer cells. Type III IFNs are characterized by having CRF2-4 and CRF2-12 receptors. The expression of IFNs Type I and III can be induced in virtually all cell types after recognition of viral components, while the production of IFN Type II is restricted to immune cells.

Type I IFNs are pleiotropic cytokines that control the secretion of pro-inflammatory cytokines and regulate the immune response that contributes to progression in AD. IFN beta 1a has been used in patients with AD in early stages; an improvement in the instrumental activities of daily life has been observed in these patients⁵⁷.

TLRs

TLRs are Type I transmembrane proteins with ectodomains that contain leucine-rich repeats that mediate the recognition of molecular PAMP; they are homologous to toll, a receptor found in insects, which participate both in the establishment of dorsoventral polarity during embryogenesis and in the immune response against fungal infections. 12 TRLs have been described in mice and 10 in humans; TLR1-9 are conserved in both. Fibrillar A β can interact directly with TLR2, TLR4, and CD14 to induce phagocytosis of A β by microglia in the early stages and neuroinflammatory responses in the advanced stages. In the early stages, the signal mediated by TLR3 increases the autophagy of A β ; it increases neuronal apoptosis in the late stages. Furthermore, TLR7, TLR8, and TLR9 can increase phagocytosis of A β early, to later contribute to neuroinflammation. TLR2 and TLR4 can be a target of therapeutic intervention in AD⁵⁸. It has also been described that the polymorphism of TLR2 -196-174del is a risk factor for late-onset AD in some populations⁵⁹.

It seems today well established that chronic inflammatory reactions are present in Alzheimer disease and are important factors that accelerate the progression of the disease. Receptors of innate immunity such as TLRs and RAGE play a central role in the perpetuation of inflammation. RAGE activation is a primary mechanism which determines self-perpetuated chronic inflammation, and RAGE cooperation with TLRs amplifies inflammatory signaling⁴⁹.

Conclusions

AD is multifactorial neurodegenerative pathology. Neuroinflammation is an early event of the presymptomatic stages in AD and contributes to the progression of the disease⁵³.

To identify the different actors participating in the inflammation and the mechanisms of damage involved can allow the development of treatments and preventive strategies in the fight against this disease.

Conflicts of interest

There are no potential conflicts of interest for any of the authors in this scientific report.

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References

- Lewis J, Dickson DW. Propagation of tau pathology: hypotheses, discoveries, and yet unresolved questions from experimental and human brain studies. *Acta Neuropathol.* 2016;131:27-48.
- Balin BJ, Hudson AP. Etiology and pathogenesis of late-onset alzheimer's disease. *Curr Allergy Asthma Rep.* 2014;14:417.
- Gatz M, Reynolds CA, Fratiglioni L, et al. Role of genes and environments for explaining alzheimer disease. *Arch Gen Psychiatry.* 2006;63:168-74.
- International Genomics of Alzheimer's Disease Consortium (IGAP). Convergent genetic and expression data implicate immunity in alzheimer's disease. *Alzheimers Dement.* 2015;11:658-71.
- Heppner FL, Ransohoff RM, Becher B. Immune attack: the role of inflammation in alzheimer disease. *Nat Rev Neurosci.* 2015;16:358-72.
- Liu C, Cui G, Zhu M, Kang X, Guo H. Neuroinflammation in alzheimer's disease: chemokines produced by astrocytes and chemokine receptors. *Int J Clin Exp Pathol.* 2014;7:8342-55.
- Birch AM. The contribution of astrocytes to alzheimer's disease. *Biochem Soc Trans.* 2014;42:1316-20.
- Lee WJ, Liao YC, Wang YF, et al. Plasma MCP-1 and cognitive decline in patients with alzheimer's disease and mild cognitive impairment: a two-year follow-up study. *Sci Rep.* 2018;8:1280.
- Zhao H, Wang Q, Cheng X, et al. Inhibitive effect of resveratrol on the inflammation in cultured astrocytes and microglia induced by A β_{1-42} . *Neuroscience.* 2018;379:390-404.
- Bahniwal M, Little JP, Klegeris A. High glucose enhances neurotoxicity and inflammatory cytokine secretion by stimulated human astrocytes. *Curr Alzheimer Res.* 2017;14:731-41.
- Parkhurst CN, Yang G, Ninan I, et al. Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor. *Cell.* 2013;155:1596-609.
- Melchior B, Puntambekar SS, Carson MJ. Microglia and the control of autoreactive T cell responses. *Neurochem Int.* 2006;49:145-53.
- Kierdorf K, Erny D, Goldmann T, et al. Microglia emerge from erythromyeloid precursors via pu.1- and irf8-dependent pathways. *Nat Neurosci.* 2013;16:273-80.
- Guedes JR, Lao T, Cardoso AL, El Khoury J. Roles of microglial and monocyte chemokines and their receptors in regulating alzheimer's disease-associated amyloid- β and tau pathologies. *Front Neurol.* 2018;9:549.
- Lanni C, Fagiani F, Racchi M, et al. Beta-amyloid short-and long-term synaptic entanglement. *Pharmacol Res.* 2019;139:243-60.
- Gay M, Evrard C, Descamps F, et al. A phenotypic approach to the discovery of compounds that promote non-amyloidogenic processing of the amyloid precursor protein: toward a new profile of indirect β -secretase inhibitors. *Eur J Med Chem.* 2018;159:104-25.
- Kirouac L, Rajic AJ, Cribbs DH, Padmanabhan J. Activation of ras-ERK signaling and GSK-3 by amyloid precursor protein and amyloid beta facilitates neurodegeneration in alzheimer's disease. *ENeuro.* 2017;4:ENEURO.0149-16.2017.
- Kim Y, Kim C, Jang HY, Mook-Jung I. Inhibition of cholesterol biosynthesis reduces γ -secretase activity and amyloid- β generation. *J Alzheimers Dis.* 2016;51:1057-68.
- Neumann U, Ufer M, Jacobson LH, et al. The BACE-1 inhibitor CNP520 for prevention trials in alzheimer's disease. *EMBO Mol Med.* 2018;10:e9316.
- Zhang H, Liu D, Huang H, Zhao Y, Zhou H. Characteristics of insulin-degrading enzyme in alzheimer's disease: a meta-analysis. *Curr Alzheimer Res.* 2018;15:610-7.
- Kazkayasi I, Burul-Bozkurt N, Ismail MA, et al. Insulin deprivation decreases insulin degrading enzyme levels in primary cultured cortical neurons and in the cerebral cortex of rats with streptozotocin-induced diabetes. *Pharmacol Rep.* 2018;70:677-83.
- Shi Y, Holtzman DM. Interplay between innate immunity and alzheimer disease: APOE and TREM2 in the spotlight. *Nat Rev Immunol.* 2018;18:759-72.
- Arboleda-Bustos CE, Ortega-Rojas J, Mahecha MF, et al. The p.R47H variant of TREM2 gene is associated with late-onset alzheimer disease in colombian population. *Alzheimer Dis Assoc Disord.* 2018;32:305-8.
- Wang Y, Ulland TK, Ulrich JD, et al. TREM2-mediated early microglial response limits diffusion and toxicity of amyloid plaques. *J Exp Med.* 2016;213:667-75.
- Li JT, Zhang Y. TREM2 regulates innate immunity in alzheimer's disease. *J Neuroinflammation.* 2018;15:107.
- Zheng H, Jia L, Liu CC, et al. TREM2 promotes microglial survival by activating wnt/ β -catenin pathway. *J Neurosci.* 2017;37:1772-84.
- Jendresen C, Årskog V, Daws MR, Nilsson LN. The alzheimer's disease risk factors apolipoprotein E and TREM2 are linked in a receptor signaling pathway. *J Neuroinflammation.* 2017;14:59.
- Tam WY, Ma CH. Bipolar/rod-shaped microglia are proliferating microglia with distinct M1/M2 phenotypes. *Sci Rep.* 2014;4:7279.
- Krbot K, Hermann P, Skorić MK, et al. Distinct microglia profile in creutzfeldt-jakob disease and alzheimer's disease is independent of disease kinetics. *Neuropathology.* 2018;38:591-600.
- Streit WJ, Braak H, Xue QS, Bechmann I. Dystrophic (senescent) rather than activated microglial cells are associated with tau pathology and likely precede neurodegeneration in alzheimer's disease. *Acta Neuropathol.* 2009;118:475-85.
- Medeiros R, Kitazawa M, Passos GF, et al. Aspirin-triggered lipoxin A4 stimulates alternative activation of microglia and reduces alzheimer disease-like pathology in mice. *Am J Pathol.* 2013;182:1780-9.
- Kirkland JL, Tchkonja T. Cellular senescence: a translational perspective. *EBioMedicine.* 2017;21:21-8.
- Bussian TJ, Aziz A, Meyer CF, et al. Clearance of senescent glial cells prevents tau-dependent pathology and cognitive decline. *Nature.* 2018;562:578-82.
- Snyder HM, Corriveau RA, Craft S, et al. Vascular contributions to cognitive impairment and dementia including alzheimer's disease. *Alzheimers Dement.* 2015;11:710-7.
- McFadyen JD, Kiefer J, Braig D, et al. Dissociation of C-reactive protein localizes and amplifies inflammation: evidence for a direct biological role of C-reactive protein and its conformational changes. *Front Immunol.* 2018;9:1351.
- Shibata M, Yamada S, Kumar SR, et al. Clearance of alzheimer's amyloid-ss(1-40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *J Clin Invest.* 2000;106:1489-99.
- Jeynes B, Provias J. Evidence for altered LRP/RAGE expression in alzheimer lesion pathogenesis. *Curr Alzheimer Res.* 2008;5:432-7.
- Erickson MA, Banks WA. Blood-brain barrier dysfunction as a cause and consequence of alzheimer's disease. *J Cereb Blood Flow Metab.* 2013;33:1500-13.
- Liu C, Chen K, Lu Y, Fang Z, Yu G. Catalpol provides a protective effect on fibrillary A β_{1-42} -induced barrier disruption in an *in vitro* model of the blood-brain barrier. *Phytother Res.* 2018;32:1047-55.
- Wong WB, Lin VW, Boudreau D, Devine EB. Statins in the prevention of dementia and alzheimer's disease: a meta-analysis of observational studies and an assessment of confounding. *Pharmacoeconomics Drug Saf.* 2013;22:345-58.
- Salech F, Ponce DP, SanMartín CD, et al. PARP-1 and p53 regulate the increased susceptibility to oxidative death of lymphocytes from MCI and AD patients. *Front Aging Neurosci.* 2017;9:310.
- Solheiro-Villavicencio H, Rivas-Arancibia S. Systemic th17/IL-17A response appears prior to hippocampal neurodegeneration in rats exposed to low doses of ozone. *Neurologia.* 2017. pii: S0213-4853(17)30194-9.
- Derkow K, Rössling R, Schipke C, et al. Distinct expression of the neurotoxic microRNA family let-7 in the cerebrospinal fluid of patients with alzheimer's disease. *PLoS One.* 2018;13:e0200602.
- Merlini M, Kirabali T, Kulic L, Nitsch RM, Ferretti MT. Extravascular CD3+ T cells in brains of alzheimer disease patients correlate with tau but not with amyloid pathology: an immunohistochemical study. *Neurodegener Dis.* 2018;18:49-56.
- SanMartín CD, Henríquez M, Chacon C, et al. Vitamin D increases a β 140 plasma levels and protects lymphocytes from oxidative death in mild cognitive impairment patients. *Curr Alzheimer Res.* 2018;15:561-9.
- Sommer A, Winner B, Prots I. The trojan horse-neuroinflammatory impact of T cells in neurodegenerative diseases. *Mol Neurodegener.* 2017;12:78.
- Ahmed ME, Iyer S, Thangavel R, et al. Co-localization of glia maturation factor with NLRP3 inflammasome and autophagosome markers in human alzheimer's disease brain. *J Alzheimers Dis.* 2017;60:1143-60.
- Tammini P, Cai Q. Defective retrograde transport impairs autophagic clearance in alzheimer disease neurons. *Autophagy.* 2017;13:982-4.

49. Navarro V, Sanchez-Mejias E, Jimenez S, et al. Microglia in alzheimer's disease: activated, dysfunctional or degenerative. *Front Aging Neurosci.* 2018;10:140.
50. Shao BZ, Cao Q, Liu C. Targeting NLRP3 inflammasome in the treatment of CNS diseases. *Front Mol Neurosci.* 2018;11:320.
51. Raikwar SP, Thangavel R, Dubova I, et al. Targeted gene editing of glia maturation factor in microglia: a novel alzheimer's disease therapeutic target. *Mol Neurobiol.* 2019;56:378-93.
52. Ciaramella A, Salani F, Bizzoni F, et al. The stimulation of dendritic cells by amyloid beta 1-42 reduces BDNF production in alzheimer's disease patients. *Brain Behav Immun.* 2013;32:29-32.
53. Janelidze S, Mattsson N, Stomrud E, et al. CSF biomarkers of neuroinflammation and cerebrovascular dysfunction in early alzheimer disease. *Neurology.* 2018;91:e867-77.
54. Lin MS, Hung KS, Chiu WT, et al. Curcumin enhances neuronal survival in N-methyl-D-aspartic acid toxicity by inducing RANTES expression in astrocytes via PI-3K and MAPK signaling pathways. *Prog Neuropsychopharmacol Biol Psychiatry.* 2011;35:931-8.
55. Ransohoff RM, El Khoury J. Microglia in health and disease. *Cold Spring Harb Perspect Biol.* 2015;8:a020560.
56. Xia MQ, Qin SX, Wu LJ, Mackay CR, Hyman BT. Immunohistochemical study of the beta-chemokine receptors CCR3 and CCR5 and their ligands in normal and alzheimer's disease brains. *Am J Pathol.* 1998;153:31-7.
57. Grimaldi LM, Zappalà G, Iemolo F, et al. A pilot study on the use of interferon beta-1a in early alzheimer's disease subjects. *J Neuroinflammation.* 2014;11:30.
58. Rubio-Araiz A, Finucane OM, Keogh S, Lynch MA. Anti-TLR2 antibody triggers oxidative phosphorylation in microglia and increases phagocytosis of β -amyloid. *J Neuroinflammation.* 2018;15:247.
59. Sohrabifar N, Gharesouran J, Talebi M, Ghojaziadeh M, Mohaddes Ardebili SM. Association of CLU and TLR2 gene polymorphisms with late-onset alzheimer disease in a Northwestern Iranian population. *Turk J Med Sci.* 2015;45:1082-6.