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Should we be anesthetizing young children?

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In 2007, the FDA advisory committee on anesthesia recommended that «children less than 3 years of age not undergo elective procedures» because of concerns regarding anesthetic toxicity⁽¹⁾. The vast majority of the evidence upon which that recommendation was based on young rodents and primates. These studies implicated virtually every available anesthetic as a cause of apoptosis in the young. However, the positive predictive value of evidence from animals for apoptosis in humans is exceedingly small, 10-20%. In addition, socializing and exercise appear to mitigate the damage in young brains, possibly explaining why the gross anatomical changes in the brains of young animals are inconsistent with the experience in humans. Perhaps more importantly, interventions including preconditioning and caspase-3 inhibitors have substantially reduced the injury in these young animals. In this lecture, we review the available evidence that demonstrates that anesthetics (and other compounds) cause neuroapoptosis and neurocognitive dysfunction in young animals, address the limited external validity of these data and present strategies that prevent or limit anesthetic-associated injury in vulnerable animals.

WHAT ARE THE HISTOPATHOLOGICAL EFFECTS OF ANESTHETIC-ASSOCIATED INJURY?

The first pathological finding of anesthetic–induced brain injury was a dramatic increase in neuroapoptosis in newborn rodents who received NMDA receptor antagonists⁽²⁾. Although neuroapoptosis during rapid synaptogenesis and brain growth is a normal phenomenon, with loss of up to 50% of cells during the period of brain development, exposure to anesthetics (that antagonize NMDA receptors or stimulate GABA_A receptors) increased the severity of the apoptosis dramatically. The maximum damage in rodents occurred on postnatal day 7, with a corresponding interval in humans of the

3rd trimester of gestation to 3 years of age⁽³⁾. Furthermore, the severity of the apoptosis increased with combinations of drugs compared with individually administered drugs as well as with the duration of exposure^(4,5). The second notable finding was the special distribution of the neuroapoptosis. The apoptosis did not occur uniformly throughout the brain and differed among animal subspecies⁽⁶⁻⁹⁾. For example in newborn mice, apoptosis was detected after ketamine and midazolam in the cerebral cortex and caudate/putamen regions, in newborn rats in the frontal cortex, thalamus, and putamen, in guinea pigs in layers IV/V of the cortex and in monkeys in the cortex. Several different markers have been used to identify and quantify the apoptosis but the final common pathway has been caspase 3 levels as well as fluoro-Jade C and silver staining^(10,11). More recent evidence has further complicated these findings, noting that apoptosis peaks in some regions of the brain in adolescence, not neonatal rodents⁽¹²⁾. This raises questions regarding the period of vulnerability suggesting that perhaps it is greater than previously appreciated and involves many more areas of the brain than previously thought to be the case.

Not only is neuroapoptosis a significant concern after exposure to anesthetics, but anesthetics also appear to interfere with other facets of brain growth including the arborization of dendrites⁽¹³⁾, development of the neurocytoskeleton, neurogenesis and lastly, changes in mitochondrial function⁽¹⁴⁾.

The mechanism by which anesthetics induce apoptosis is poorly understood. How GABA_A agonists and NMDA receptor antagonists trigger apoptosis remains an area of active investigation. Currently, we know that neural cells undergo apoptosis by either the intrinsic or extrinsic pathway, the former dependent on mitochondrial pathways and the latter dependent on surface receptor activation^(3,15). These may be complements by involvement of neurotrophic factors and a neuronal cell dependent pathway⁽¹⁶⁾. During brain cell remodeling, cells that are active survive whereas those that are

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recognized as being inactive undergo apoptosis⁽¹⁴⁾. Similarly, synaptic development and dendritic growth are linked to the activity levels of the cells, resulting in a similar effect. It is possible that anesthetics induce a state of neuronal inactivity, rendering the cells vulnerable to self-destruction or apoptosis.

WHICH ANESTHETICS/DRUGS ARE INVOLVED AND HOW ARE THEY INVOLVED?

A series of experiments were performed in young rodents in which they received one or more anesthetics including ketamine, midazolam, propofol, inhalational anesthetics, and nitrous oxide (*Table I*) $^{(4-6,17-20)}$. All of these anesthetics are GABA, receptor agonists and/or NMDA receptor antagonists, although some are non-anesthetics (eg., dexamethasone). In fact, all anesthetics cause apoptosis and neurocognitive dysfunction when given in large enough doses for long enough periods except the alpha2 agonists (eg., dexmedetomidine), muscle relaxants and possibly opioids. Investigators confirmed that as in the case of alcohol toxicity, maximum apoptosis occurred in postnatal day 7 (PND 7) young rodents, although there is recent evidence that older rodents may develop apoptosis in other parts of the brain, heretofore not the focus of previous investigations (12). Most investigations have involved newborn rats or mice, although a number have studied fetal animals including non-human primates^(21,22). The severity of the apoptosis after propofol in fetal monkeys is comparable with that in the newborn monkey $^{(22)}$.

Combinations of anesthetics cause more apoptosis than individual anesthetics. In fact, N_2O does not cause apoptosis alone, but the combination such of N_2O and ketamine resulted in apoptosis in specific parts of the brain that exceeded the additive effects of apoptosis by the individual anesthetics⁽²³⁾. Hence, it appears that combinations of anesthetics augment the severity of the apoptosis to a greater extent than that explained by the sum of the apoptosis of the individual anesthetics. Several anesthetics have been shown to be anti-apoptotic

Table I. Anesthetics anti-apoptotic.

Pro-apoptotic:

- Isoflurane, sevoflurane, desflurane, N2O, ± xenon
- Propofol, thiopental, ketamine, midazolam, diazepam, MgSO₄, dexamethasone
- CO₂, acetaminophen

Anti-apoptotic:

Lithium, melatonin, clonidine

Non-apoptotic:

- Dexmedetomidine, acute exposure to opioids Unknown:
 - Muscle relaxants

(*Table I*). Lithium, melatonin, dexmedetomidine and clonidine prevent apoptosis-induced by other agents. Indeed, they have been shown to prevent ketamine-induced neuroapoptosis in the newborn rodent models^(24,25).

Beyond the histopathology associated with administering anesthetics to young infants, administering an anesthetic impairs neurocognitive development in these young animals. The evidence is unequivocal that the neurocognitive development in young animals exposed to anesthetics is impaired compared to those not exposed and that the dysfunction is not transient but continues into adulthood⁽²⁶⁻²⁸⁾. A variety of cognitive tests including memory, spatial orientation and learning are impaired suggesting substantially impaired cognitive function.

Numerous editorials and letters suggested that methodological errors limit the external validity of the data. Although subsequent studies confirmed reasonably normal physiologic conditions, blood gases and glucose concentrations, the effects of other factors such as pain and surgery remained conflicting.

Current animal evidence points to a direct effect of anesthetics on the immature brain resulting in neuroapoptosis, changes in brain development and long-term cognitive impairment in newborn animals. Most anesthetics that we use today have been implicated in this pathology, although some have been shown to confer anti-apoptotic or no apoptotic properties. Exercise and socializing and several specific drugs and interventions have been shown to attenuate both the histopathological impairment associated with anesthetic exposure in newborn animals⁽²⁹⁾. However, what is the positive predictive value of animal data to humans? The answer is ~10%. Hence, we can continue to perform animal studies but until there is confirmation in humans, these data remain of interest in animals only.

Studies have attempted to link the above animal data to humans. To what end has a link been found? First, a host of retrospective, flawed studies have been published, most of which claimed that a link exists between anesthetic exposure multiple times before the age of 3 years and subsequent learning disabilities and other deficits⁽³⁰⁾. A large database in identical twins in Denmark however found no difference in school-tested performance in twins who were discordant for anesthetics⁽³¹⁾. Second, two prospective studies now have failed to identify any difference in cognitive function between general anesthetic exposure in young children and precise neurocognitive testing at a later age^(32,33). These two recent studies are consistent with our decades of anecdotal observations as a collective pediatric anesthesia body that general anesthetics are UNLIKELY to induce cognitive dysfunction in children. Additional studies will assist in closing the door on this interesting roller coaster ride of questioning the safety of anesthesia in children.

REFERENCES

- Rappaport B, Mellon RD, Simone A, Woodcock J. Defining safe use of anesthesia in children. NEJM. 2011;364:1387-1390.
- Ikonomidou C, Bosch F, Miksa M, et al. Blockade of NMDA receptors and apoptotic neurodegeneration in the developing brain. Science. 1999:283:70-74
- Yon JH, Daniel-Johnson J, Carter LB, Jevtovi-Todorovic V. Anesthesia induces neuronal cell death in the devveloping rate brain via the intrinsic and extrinsic apoptotic pathways. Neuroscience. 2005;135:815-827.
- Young C, Jevtovic-Todorovic V, Qin YQ, et al. Potential of ketamine and midazolam, individually or in combination, to induce apoptotic neurodegeneration in the infant mouse brain. Br J Pharmacol. 2005;146:189-197.
- Jevtovic-Todorovic V, Benshoff N, Olney JW. Ketamine potentiates cerebrocortical damage induced by the common anaesthetic agent nitrous oxide in adult rats. Br J Pharmacol. 2000;130:1692-1698.
- Rudin M, et al. Single-dose ketamine administration induces apoptosis in neonatal mouse brain. J Basic Clin Physiol Pharmacol. 2005;16:231-243
- Scallet AC, et al. Developmental neurotoxicity of ketamine: morphometric confirmation, exposure parameters, and multiple fluorescent labeling of apoptotic neurons. Toxicol Sci. 2004;81:364-370.
- 8. Rizzi S, et al. Clinical anesthesia causes permanent damage to the fetal guinea pig brain. Brain Pathol. 2008;18:198-210.
- Brambrink AM, Evers AS, Avidan MS, et al. Isoflurane-induced neuroapoptosis in the neonatal rhesus macaque brain. Anesthesiology. 2010;112:834-841.
- Hengartner MO. The biochemistry of apoptosis. Nature. 2000;407:770-776.
- Olney JW, Wozniak DF, Jevtovic-Todorovic V, et al. Drug-induced apoptotic neurodegeneration in the developing brain. Brain Pathol. 2002;12:488-498.
- Deng M, Hofacer RD, Jiang C, et al. Brain regional vulnerability to anaesthesia-induced neuroapoptosis shifts with age at exposure and extends into adulthood for some regions. Br J Anaesth. 2014;113:443-451.
- Head BP, Patel HH, Nissman IR, et al. Inhibition of p75 neurotrophin receptor attenuates isoflurane-mediated neuronal apoptosis in the neonatal central nervous system. Anesthesiology. 2009;110:813-825.
- Davidson A, Flick RP. Neurodevelopmental implications of the use of sedation and analgesia in neonates. Clin Perinatol. 2013;40:559-573.
- Reddy SV. Effect of general anesthetics on the developing brain. J Anaesth Clin Pharm. 2012;28:6-10.
- Olney JW, Chainllie Y, Wozniak DF, et al. Anesthesia-induced developmental neuroapoptosis: does it happen in humans? Anesthesiology. 2004;101:273-275.
- Cattano D, Young C, Straiko MMW, et al. Subanesthetic doses of propofol induce neuroapoptosis in the infant mouse brain. Anesth Analg. 2008;196:17112-17114.

- Istaphanous GK, Howard J, Nan X, et al. Comparison of the neuroapoptotic properties of equipotent anesthetic concentrations of desflurane, isoflurane, or sevoflurane in neonatal mice. Anesthesiology. 2011;114:578-587.
- Kodama M, Satoh Y, Otsubo Y, et al. Neonatal desflurane exposure induces more robust neuroapoptosis than do isoflurane and sevoflurane and impairs working memory. Anesthesiology. 2011;115:979-991.
- Ramage TM, Chang FL, Shih J, et al. Distinct long-term neurocognitive outcomes after equipotent sevoflurane or isoflurane anaesthesia in immature rats. Br J Anaesth. 2013;110(S1):i39-i46.
- 21. Creeley C, Dikranian K, Dissen G, et al. Propofol-induced apoptosis of neurones and oligodendrocytes in fetal and enonatal rhesus macaque brain. Br J Anaesth. 2013;110(S1):i29-i38.
- Brambrink AM, Evers AS, Avidan MS, et al. Ketamine-induced neuroapoptosis in the fetal and neonatal rhesus macaque brain. Anesthesiology. 2012;116:372-384.
- Young C, Jevtovic-Todorovic V, Qin YQ, et al. Potential of ketamine and midazolam, individually or in combination, to induce apoptotic neurodegeneration in the infant mouse brain. Br J Pharmcol. 2005;1246:189-197.
- Pontén E, Viberg H, Gordh T, et al. Clonidine abolishes the adverse effects on apoptosis and behaviour after neonatal ketamine exposure in mice. Acta Anaesthesiol Scand. 2012;56:1058-1065.
- 25. Turner CP, Gutierrez S, Liu C, et al. Strategies to defeat ketamine-induced neonatal brain injury. Neuroscience. 2012;210:384-392.
- Stratmann G, Sall JW, May LDV, et al. Beyond anesthetic properties: the effects of isoflurane on brain cell death, neurogenesis, and long-term neurocognitive function. Anesth Analg. 2010;110:431-437.
- Stratmann G, Sall JW, May LDV, et al. Isoflurane differentially affects neurogenesis and long-term neurocognitive function in 60-day-old and 7-day-old rats. Anesthesiology. 2009;110:834-848.
- Paule MG, Li M, Allen RR, et al. Ketamine anesthesia during the first week of life can cause ong-lasting cognitive deficits in rhesus monkeys. Neurotoxicol Teratol. 2011;33:220-230.
- Shih J, May LDV, Gonzalez HE, et al. Delayed environmental enrichment reverses sevoflurane-induced memory impairment in rats. Anesthesiology. 2012;116:586-602.
- Wilder RT, Flick RP, Sprung J, et al. Early exposure to anesthesia and learning disabilities in a population-based birth cohort. Anesthesiology. 2009;110:796-804.
- Bartels M, Althoff RR, Boomsma DI. Anesthesia and cognitive performance in children: no evidence of a causal relationship. Twins Research Human Genetics. 2009;12:246-253.
- 32. Davidson AJ, Disma N, de Graaff JC, et al. Neurodevelopmental outcome at 2 years of age after general anaesthesia and awake-regional anaesthesia in infancy (GAS): an international multicenter, randomized controlled trial. Lancet. 2016;387:239-250.
- Sun LS, Li G, Miller TLK, et al. Association between a single general anesthesia exposure before age 36 months and neurocognitive outcomes in later childhood. JAMA. 2016;315:2312-2320.

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