

Evaluation of the *aroA* mutant of *Corynebacterium pseudotuberculosis* in cellular and murine models

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Abstract

Caseous lymphadenitis of small ruminants causes economic losses worldwide. To date, no effective vaccine has been developed against the causative agent of this disease, *Corynebacterium pseudotuberculosis*. The objective of the present work was to evaluate an *aroA* mutant gene strain of *C. pseudotuberculosis* in cellular and murine models, for attenuation and the ability to stimulate an immune response. The intracellular survival of the *aroA* mutant strain and the wild type strain (WT) of *C. pseudotuberculosis* was evaluated in J774A.1 murine macrophages using a multiplicity of infection (MOI) of 1:1 with the following infection times: 30 min, and 1, 2, 4, 8, 12, and 24 h. The largest difference in the intracellular survival of the mutant was observed 30 min post-infection. After subcutaneous skin vaccination, the subcutaneous lesion progression observed on the 14th day was more severe in those animals that were vaccinated with the WT strain. An analysis of the residual virulence in the murine model did not reveal any bacteria in mice vaccinated with the *aroA* strain on day 28 post-vaccination. Mice vaccinated with the mutant showed 50 % protection against the intraperitoneal challenge, exceeding that of the control group (41.67 %). We conclude that the virulence of the *aroA* mutant was significantly attenuated in both cellular and murine models according to the residual virulence detected in mice. However, vaccination with the mutant failed to confer at least 80 % protection, which is desirable for an immunogen. Hence, this study contributes to the knowledge of the immune response against *Corynebacterium pseudotuberculosis*.

Keywords: *Corynebacterium pseudotuberculosis*, *aroA* gene, vaccination, cellular model, murine model.

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Introduction

Corynebacterium pseudotuberculosis is a Gram-positive facultative intracellular bacterium and intracellular coccobacillus, capable of fermenting carbohydrates, producing urease, phospholipase D and catalase. Two biotypes are known of this species, a nitrate positive group infecting only horses and a nitrate negative group infecting only goats and sheep¹. *C. pseudotuberculosis* is the causative agent of

caseous lymphadenitis in small ruminants and typically produces abscesses with caseous necrosis in superficial lymph nodes, subcutaneous tissue, lungs, kidneys, liver, and spleen². In infected horses, there are three clinical manifestations: subcutaneous or muscular abscess, mainly in the pectoral and abdominal regions; internal infection causing visceral abscess (liver, kidney, lungs, and spleen); and ulcerative lymphangitis, which results in limb inflammation, cellulitis and severe lameness³. To control this disease, several bacterins and live vaccines have been applied and found to exert partial protective effects⁴. The use of a bacterin-toxoid prepared with the phospholipase D exotoxin increased the antibody levels against exotoxin in sheep, reducing the number and size of the lesions relative to those in non-immunized animals⁵. However, vaccination with this preparation did not protect the animals against experimental infection⁵.

The maintenance of a live strain of *C. pseudotuberculosis* for use as an immunogen has been attempted, but the results have not been satisfactory. One strategy that has been used to obtain attenuated strains of microorganisms is to perform several mutations in the genes affecting bacterial metabolism, such as those associated with aromatic amino acids⁶. In this regard, the *aro* genes have become a mutational target for modification and virulence attenuation of pathogenic strains of bacteria. The *aro* genes are involved in the shikimate biosynthetic pathway, in which 3-deoxy-D-heptulosonate-7-phosphate (DAPH) is converted to chorismate, which is required for the synthesis of aromatic amino acids that are important for bacterial metabolism. Disruption of genes associated with this pathway has been successfully used for the generation of live attenuated bacterial strains⁷. For example, several mutations have been achieved in the *aroA* gene of *Salmonella enterica*, subsp. *enterica* serovar Abortus equi, and it was observed that one major effect produced by these mutations corresponds to the inability of the bacteria to generate aromatic metabolites, such as phenylalanine, tyrosine, and tryptophan, which are essential for bacterial metabolism; thus, these mutants are incapable of intracellular replication⁶. Additionally, these mutations altered the regulation of the *murA* gene, which is involved in peptidoglycan synthesis, thereby causing several defects in the bacterial cell wall and the outer bacterial membrane and facilitating phagocytosis of the mutant^{6, 8}. In a study of the cp09 mutant of *C. pseudotuberculosis*, BALB/c mice were immunized and showed 50 % survival after being challenged, intraperitoneally (i. p.), with a virulent strain at a dose of 1×10^4 colony-forming units (CFU)⁹.

The *aroA* gene mutant of *Salmonella enterica*, subsp. *enterica* serovar Abortus equi administered by the oral and intramuscular routes induced 100 % protection against experimental infection of guinea pigs¹⁰. In contrast, attenuated mutant strains affecting the *aroB* and *aroQ* genes of *C. pseudotuberculosis* were used as immunogens in sheep but did not confer adequate protection against the experimental challenge¹¹. An attenuated live vaccine is needed to stimulate an adequate cellular immune response and protect animals against Caseous lymphadenitis. Based on this rationale, we developed an *aroA* mutant strain of *C. pseudotuberculosis* and tested its attenuation in a cellular and a murine model. Finally, we verified its ability to elicit an immune response to protect mice against a challenge with a virulent strain of *C. pseudotuberculosis* in the murine model.

Materials and methods

The experimental procedure was performed under the approval of the Animal Care and Use Ethics Committee (SICUAE) of Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México.

Bacterial strains, media, and culture conditions

The TOP'10 *E. coli* strain (F⁻ *mcrA* Δ (*mrr-hsdRMS-mcrBC*) ϕ 80/*lacZ* Δ M15 Δ *lacX74* *recA1* *araD139* Δ (*ara-leu*) 7697 *galU galK rpsL* (Str^R) *endA1 nupG* λ ; Invitrogen, Carlsbad, CA, USA) was cultivated in Luria Bertani (LB) medium (Difco, Detroit, MI, USA) with orbital shaking at 200 rpm for 16 h at 37 °C (Barnstead/Lab-line Max 4000, USA). The wild type strain of *C. pseudotuberculosis* (WT) isolated from a goat and the reference strain of ATCC 43924 *C. pseudotuberculosis* were grown at 37 °C in tryptic soy medium (TSB, Difco) for 20 h or in tryptic soy agar (TSA, Difco) for 48 h (Barnstead/Lab-line Max 4000, USA). The *aroA* mutant strain of *C. pseudotuberculosis* was cultivated under the same conditions as the WT, with 50 μ g/ml of kanamycin or 100 μ g/ml of ampicillin (Sigma Aldrich, St. Louis, MO, USA), added to the culture medium.

Construction of the *aroA* mutant

DNA extraction was performed using guanidine thiocyanate¹², and *aroA* gene amplification was achieved using the 5'-CCCGCTCGACAAAACCTGCA-3' and 5'-ATA-ACTGTTGCGCCCCGAGTC-3' primers designed from the genome sequence of *C. pseudotuberculosis* (NCBI-GenBank Accession No. NC014329), which amplified 2,138 bp, including the entire gene (1,413 bp) and the flanking sequences.

Polymerase chain reaction (PCR) reactions were performed in a final volume of 50 μ l using the following amplification conditions: one cycle of initial denaturation at 94 °C for 5 min; 33 cycles at 94 °C for 45 sec, 65 °C for 45 sec, and 72 °C for 2 min; and a final extension cycle at 72 °C for 10 min. The amplification product was cloned into the pCR2.1 TOPO plasmid (Invitrogen, Ventura, CA, USA)¹³. The plasmid containing the amplified product was digested with the *EcoRI* enzyme (Invitrogen) to verify the cloning and was denoted pIAroA. The pIAroA plasmid was then digested with *HpaI* enzyme (Fermentas, USA). *HpaI* is a site that is unique to the *aroA* gene; the 1222-bp kanamycin resistance gene was ligated at this site, generating the pIAroA-km plasmid, which was transformed by electroporation into the WT strain of *C. pseudotuberculosis* (2.5 kV, 200 Ω , 25 μ f, and 200 ng of plasmid)¹⁴. Transformant selection was performed based on the kanamycin resistance and ampicillin sensitivity of *C. pseudotuberculosis*. The level of chromosomal mutation was verified by PCR using the original primers; the PCR product was purified with a commercial QIAquick PCR Purification Kit (Qiagen, La Jolla, CA, USA). The nucleotide sequence in both directions was determined using Taq FS Dye Terminator Cycle Sequencing-Based on Fluorescence Sequencing and were analyzed on a DNA sequencer (Applied Biosystems 7 3730 DNA sequencing system). Sequences were verified with the Vector NTI program (Applied Biosystems, Foster City, California, USA) and Clone Manager software (Scientific & Educational Software, Morrisville, North Carolina, USA).

Evaluation in the cellular model

Prior to use the mouse model, attenuation of the mutant strain was established by comparing the *aroA* mutant with the virulent parental strain, and determining the time and the number of CFU surviving in infected cells. To evaluate the par-enteral and mutant strain survival of *C. pseudotuberculosis*, the J774A.1 murine macrophage cell line was used¹⁵. Cells were maintained in Dulbecco's modified Eagle's medium (DMEM, Invitrogen) containing 10 % fetal bovine serum (FBS), 1 % non-essential amino acids, and 2-mM L-glutamine (DMEMc). For maintenance and infection, the cells were incubated with 5 % CO₂ at 37 °C. Macrophages were cultivated in DMEM supplemented with 10 % FBS and 2-mM L-glutamine without antibiotics and then placed in polystyrene 24-well plates at a concentration of 5 x 10⁵ cells/ml at 37 °C with 5 % CO₂ for 18 h. Each experiment included two independent replications, and the infection assay was repeated three times.

Bacteria (*C. pseudotuberculosis*) were grown in brain-heart infusion (BHI) medium with orbital shaking at 200 rpm and 37 °C for 20 h until they reached the exponential phase. Prior to cell infection, they were diluted in DMEMc at 37 °C. Macrophages were infected at a multiplicity of infection (MOI) of 1: 1. The cell culture plates were centrifuged at 400xg for 5 min and incubated for 30 min at 37 °C with 5 % CO₂. The end of this incubation was considered time zero of the infection. The cells were washed three times with phosphate-buffered saline (PBS), and 100-mg/ml gentamycin was added to the DMEMc solution to remove extracellular bacteria. Then, the cells were incubated for 1 h at 37 °C with 5 % CO₂, at which point, the gentamycin concentration in the medium was decreased to 10 mg/ml for the remainder of the experiment. Intracellular bacterial survival was determined at 1, 2, 4, 8, 12, and 24 h after time zero. The number of viable intracellular bacteria was measured by lysing the infected cells with 0.5 ml of 0.1 % Triton-PBS. The CFU/ml value was then established by inoculating TSA plates, with 50 µg/ml kanamycin added for cells infected with the *aroA* mutant strain.

Evaluation in the murine model

To observe the appearance of skin lesions, establish the residual virulence of the *aroA* mutant strain, and evaluate *in vivo* mutant protection in terms of the immune response, BALB/c mice (6 to 7 weeks of age) were used. These mice were provided with food and water *ad libitum*.

Evaluation of the lesions present at the vaccination site

One problem associated with experimental live vaccines of *C. pseudotuberculosis* is the occurrence of lesions at the vaccination site^{9, 16}. To demonstrate the *in vivo* mutant attenuation, the degree of the lesion at the vaccination site was evaluated. Five groups, each with three BALB/c mice, were used as follows: The first group was subcutaneously vaccinated in the midline of the back with the WT strain using 2 x 10⁵ CFU in 50 µl of PBS. The second group was vaccinated with the mutant strain through the same route and using the same dosage as for the first group. The third and fourth groups were vaccinated in the footpad with 2 x 10⁵ CFU in 50 µl of PBS, one with the WT and the other with the mutant strain. A control group was included and was injected with PBS in the footpad and midline of the back. The

animals were monitored daily for 21 days to assess the degree of injury, abscess formation, and presence of fistula or exudate. They were euthanized on day 21. The mice footpad and subcutaneous tissues were kept in 10 % formalin for histopathological analysis¹⁷.

Residual virulence of the aroA mutant strain

Two groups, each with 27 BALB/c mice, were used as follows: Animals in the first group were vaccinated subcutaneously with the *aroA* mutant strain (2×10^5 CFU/0.1 ml of PBS), and in the second group (control) animals were vaccinated with wild type bacteria (WT), using the same route and the same dose as that used in the first experimental group. The animals were euthanized in groups of three on days 0, 3, 6, 10, 14, 21, 28, 35, and 42 post-vaccination. Mice spleen and liver tissues were obtained and macerated in 1 ml of PBS. The homogenized organs were diluted at 1:10 in PBS for CFU counting, and then vaccinated (in triplicate) in TSA plates or 50- μ g/ml kanamycin TSA plates, and incubated for 48 h at 37 °C.

Evaluation of the in vivo mutant protection

Twenty-four mice were vaccinated with *aroA* mutant strain at a concentration of 2×10^5 CFU/ml via subcutaneous injection in the withers region. The control group was injected with PBS. The challenge was performed on day 21 post-vaccination using the ATCC 43924 *C. pseudotuberculosis* strain at a concentration of 1×10^4 /0.1 ml in PBS, by intraperitoneal route⁹. The animals were monitored daily, and on day 28, they were euthanized and underwent necropsy for bacteriological studies in the liver and spleen.

Evaluation of the humoral and cellular immune responses induced by the mutant

An indirect commercially available enzyme-linked immunosorbent assay (ELISA) system (mouse monoclonal antibody isotyping reagent, Sigma–Aldrich) was used to determine the presence of immunoglobulins (Igs) of the IgG1, IgG2a, IgG2b, and IgM isotypes induced by *C. pseudotuberculosis*. Two groups of 40 BALB/c mice were used as follows: One group was subcutaneously immunized with the *aroA* mutant strain at a concentration of 2×10^5 CFU/0.1 ml, and the other group (control) was injected with PBS using the same subcutaneous route. Serum was obtained on days 0, 3, 5, 7, 9, 12, 14, 21, 28, 35, and 42 post-vaccination. A protein extract of the ATCC 43924 *C. pseudotuberculosis* strain was used as the antigen. For this purpose, the bacteria were grown in BHI medium for 24 h at 200 rpm. They were then sonicated for 12 min at 60 Hz, and the obtained mass was centrifuged at 960 xG for 10 min. The material was diluted to a concentration of 4 μ g/ml, as quantified by the Bradford technique, diluted again in 0.1-M sodium bicarbonate-carbonate solution (pH 9.6), and incubated for 24 h at 37 °C in 24-well polystyrene plates. Finally, the samples were washed three times with PBS-T (PBS 1X, pH 7.4, 0.05 % Tween 20). A solution of 1 % skim milk was used to block the wells, and the plates were incubated for 1 h at 37 °C. The varied serum

samples were diluted 1:200 in PBS and added to the plates in duplicate¹⁸. The plates were subsequently incubated for 1 h at 37 °C and washed three times with PBS-T. Then, 100 µl of the corresponding isotype (anti-IgG 1, anti-IgG2a, anti-IgG2b, or anti-IgM) was added and diluted to 1:1,000 in PBS. The plates were incubated at room temperature for 30 min and washed three times with PBS-T. Peroxidase-conjugated Anti-IgG (obtained from a goat) was used as the secondary antibody (Sigma-Aldrich) at a concentration of 1:5,000 in PBS-T. The plates were washed with PBS-T, and 5-aminosalicylic acid was added at a concentration of 1 mg/ml. Then, the plates were incubated for 15 min at room temperature, and 50 µl of stopping solution (3-N NaOH) was added. Finally, the plates were examined at a wavelength of 490 nm.

To determine the cytokine production induced by the *aroA* mutant, we tested and compared two groups of 40 BALB/c mice. On day 0, one group was subcutaneously vaccinated with 2×10^5 CFU/0.1 ml of the *aroA* mutant strain; and the control group was vaccinated with PBS. Mice were sacrificed in groups of four on days 0, 3, 5, 7, 9, 12, 14, and 21¹¹. The mice spleens were obtained under sterile conditions and washed three times with Hank's balanced salt solution, and placed in a Petri dish containing 5 ml of DMEM (Gibco, Invitrogen, USA) with 100 IU/ml penicillin and 100 µg/ml streptomycin. The spleens were wrapped in sterile gauze pads to retain the tissue debris and macerated, and the cell suspension was transferred into a conical 15 ml plastic tube containing 5 ml of the same cell culture medium. They were then centrifuged at 400 x G for 3 min. To lyse the erythrocytes, the pellet was re-suspended in 1 ml of 0.17 M ammonium chloride per spleen for 5 min at 4 °C. Three more washes with DMEM supplemented with 10 % FBS, 2 mM L-glutamine, and 0.1 mM non-essential amino acids (Gibco) were performed¹⁹. Subsequently, 5×10^6 lymphocyte cells were extracted from the mutant-vaccinated and control mice spleens and placed in 24-well polystyrene plates. To each well, 50 µg of a protein extract of the ATCC 43924 reference strain of *C. pseudotuberculosis* diluted in PBS was added. The cells were then cultivated until the log phase for 24 h in BHI medium. Then, they were centrifuged, washed three times with PBS, and finally, sonicated for 12 min at 60 Hz¹⁸. The protein concentration was determined using the Bradford method and adjusted as required.

A positive control containing 10 µg/ml pokeweed (Invitrogen) and a negative control consisting of DMEM (Gibco) were used to compare to the experimental group. The plates were incubated at 37 °C with 5 % CO₂; and the supernatant was collected 24 h after vaccination and frozen at -20 °C until use. Commercial cytokine detection systems for mouse IFN-γ, TNF-α, IL-4, and IL-10 were used following the manufacturer's instructions (Duoset ELISA; R&D Systems, Minneapolis, MN, USA).

Statistical analysis

General mixed models of repeated observations were used to evaluate the *aroA* mutant experiments in the cellular model, residual virulence in the liver and spleen, and the immune response²⁰. For the protection experiment, the survival rates of the vaccinated and control groups were compared after the challenge. Statistical significance was considered at $P < 0.05$, and analyses were performed using SAS software.

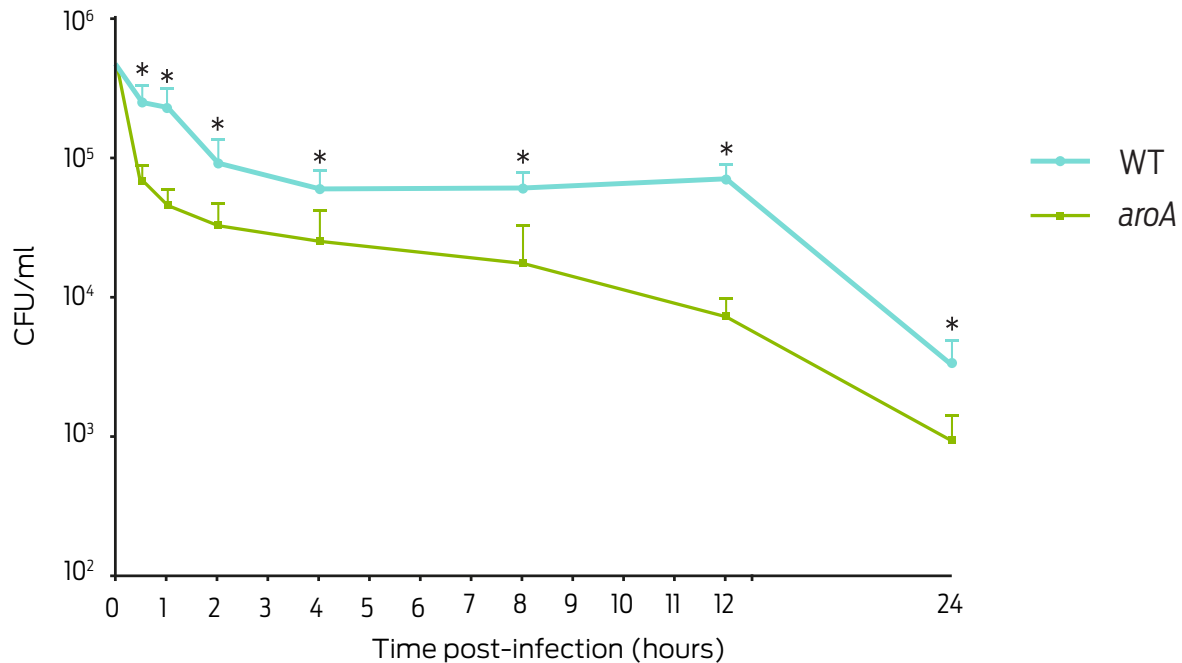


Figure 1. Intracellular survival of *C. pseudotuberculosis* in J774A.1 macrophages with a Multiplicity of infection (MOI) of 1:1. The results are presented as the mean \pm standard deviation in CFU/ml after different post-infection times. Significant differences ($P < 0.05$) between the strains are indicated by an asterisk.

Results and discussion

Evaluation in the cellular model

The J774A.1 macrophage infections performed with the WT and *aroA* strains showed a significant difference 30 min post-infection as follows: $273,333.30 \pm 40,221.61$ CFU/ml for the WT strain and $75,000.00 \pm 8,062.26$ CFU/ml for the *aroA* strain ($P < 0.05$). For the following infection times, a gradual decrease in the number of intracellular bacteria was observed over time between the two bacteria strains: At 12 h, one log of difference between the two strains was observed. At the end of the experiment (24 h), the WT strain exhibited a value of $3,566 \pm 691$ CFU/ml and that of the *aroA* mutant strain was 966.67 ± 221.61 CFU/ml (Figure 1).

Evaluation of the lesion formation at the vaccination site

Vaccinating mice with the WT and mutant strains caused injuries on the footpad and in the subcutaneous tissue. The progression of the lesion was more rapid and severe in the mice vaccinated with the WT strain than in those vaccinated with the mutant strain. In the three WT-vaccinated mice, a well-defined nodular lesion with a diameter of approximately 1.5 cm was formed within the zones showing alopecia. A continuing loss of hair was compatible with the formation of an ulcer. A severe inflammatory reaction comprising macrophages and lymphocytes, which primarily affected the hypodermis with a slight invasion of the dermis, was observed microscopically. This lesion also presented a suggestive loss of cell continuity to edema. In contrast, the mice that were vaccinated with the *aroA* strain in their backs via subcutaneous injection exhibited a slight inflammatory reaction consisting of lymphocytes in the hypodermis region.

At the dermis level, a suggestive loss of cell continuity to edema was observed but was less intense compared to that resulting from the WT strain. The application of the *aroA* mutant in the footpad caused less-severe injuries than WT strain. One mouse vaccinated via the footpad died, and others suffered liver abscesses. Microscopically, a discrete inflammatory reaction involving localized lymphocytes in the hypodermis region was observed in the *aroA* mutant-vaccinated mice, and at the dermis level, thus suggesting of the loss of cell continuity to edema.

In the WT-vaccinated animals, a severe inflammatory reaction comprising macrophages and lymphocytes occurred, affecting the hypodermis with a slight invasion of the dermis. A loss of cell continuity to edema was also observed in this case. In the *aroA* mutant-vaccinated group, an additional discrete inflammatory reaction involving mononuclear cells (lymphocytes) in the hypodermis region was observed accompanied by a loss of cell continuity to edema at the dermis level.

Evaluation of the residual virulence

Throughout the experiment, animals vaccinated with the WT strain showed 12.5 % (3/24) mortality, whereas mice that were vaccinated with the *aroA* strain exhibited no mortality. Regarding the number of bacteria isolated from the spleen, a significant difference between the WT and *aroA* strains was observed on days 3; (405.55 ± 89.58 CFU/ml vs 7.41 ± 2.93 CFU/ml), 10 (11,175.91 ± 74.07 CFU/ml vs 5.50 ± 2.77 CFU/ml), and 14 (3,444.44 CFU/ml ± 430.56 vs 3.07 ± 2.45 CFU/ml). No bacteria were recovered on day 28 post-vaccination in the *aroA* mutant-vaccinated mice (Figure 2). On days 0, 3, and 6 post-vaccinations, there was no significant difference in the CFU numbers recovered from mice livers. In contrast, on day 14, the liver bacterial load of mice infected with the WT strain was significantly higher ($7.03 \times 10^7 \pm 1.63 \times 10^7$ CFU/ml) than that of the *aroA* mutant-vaccinated mice (3.70 ± 1.69 CFU/ml). On day 21, the livers of animals infected with the WT strain had a bacterial count of $2,490.74 \pm 305.70$ CFU/ml, whereas no bacteria was isolated from those infected with the *aroA* strain. However, on day 28, values of 472.22 ± 164.94 CFU/ml and 7.41 ± 4.90 CFU/ml were determined in the livers of WT- and *aroA*-infected mice (Figure 2).

Evaluation of the in vivo mutant protection

To evaluate the protective ability of the *aroA* mutant strain against challenge, the animals were immunized and subsequently challenged with the ATCC43924 *C. pseudotuberculosis* strain. During the first post-challenge week, a marked significant difference ($P < 05$) was observed between the survival of the control group (41.67 %; 10/24 animals) and the *aroA* mutant-vaccinated group (83.33 %; 20/24 animals). However, by day 28 of the experiment, survival in control group 40 % was not significantly different than survival in the mutant-vaccinated mice 50 % (Figure 3).

Humoral immune response

The production of IgM, IgG1, IgG2a, and IgG2b before exposure to the *aroA* mutant strain was evaluated, *aroA* mutant-vaccinated mice showed a statistically significant

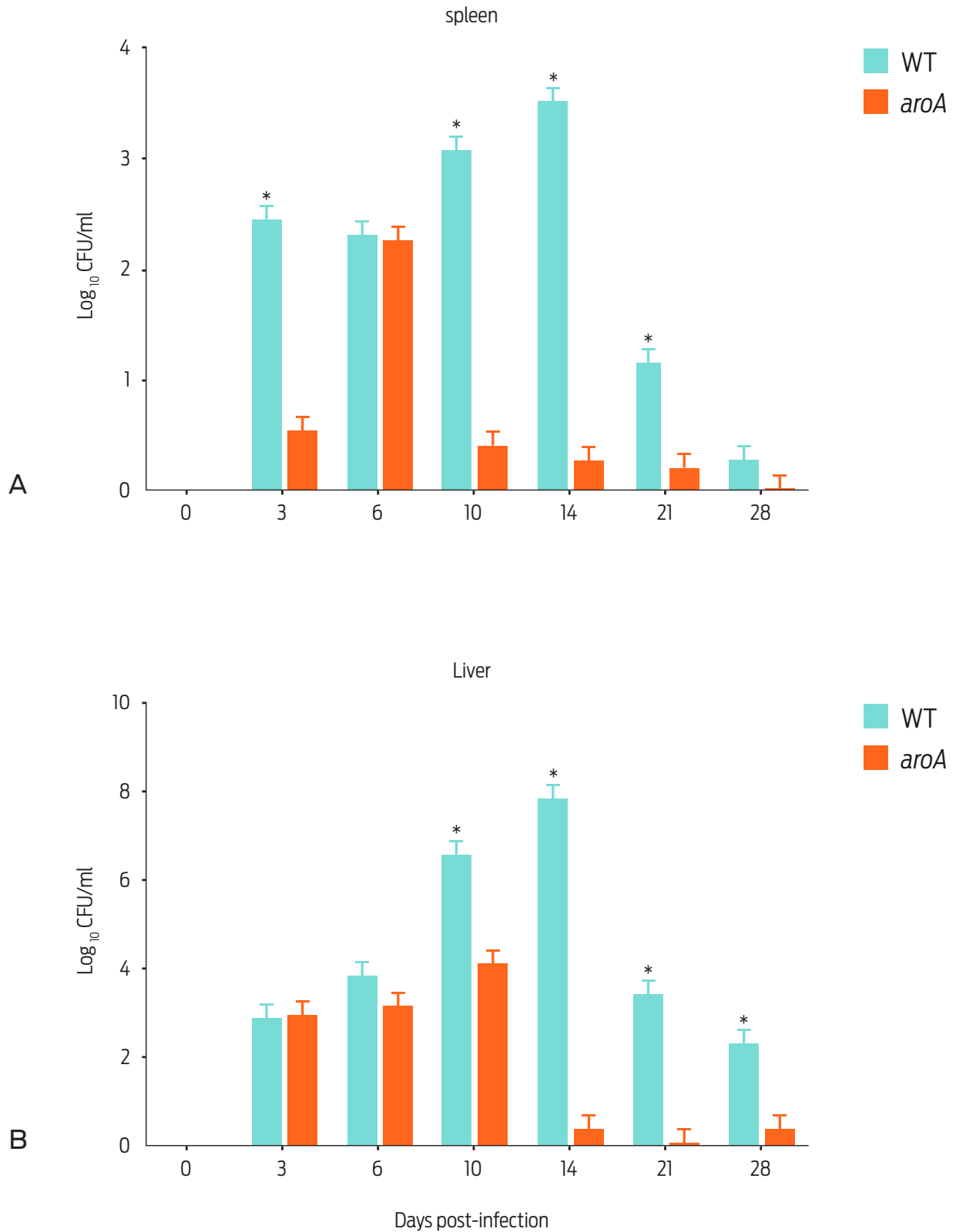


Figure 2. Recovery of the WT and *aroA* strains of *C. pseudotuberculosis* from the spleens and livers of mice that were subcutaneously vaccinated with 2×10^6 CFU/0.1 ml. The animals were euthanized in groups of three on days 0, 3, 6, 10, 14, 21, 28, 35 and 42 post vaccination. The data represented as the mean \pm standard deviation. On days 35 and 42, no bacteria could be recovered. Significant differences ($P < 0.05$) between mutant and parental strain are indicated by an asterisk.

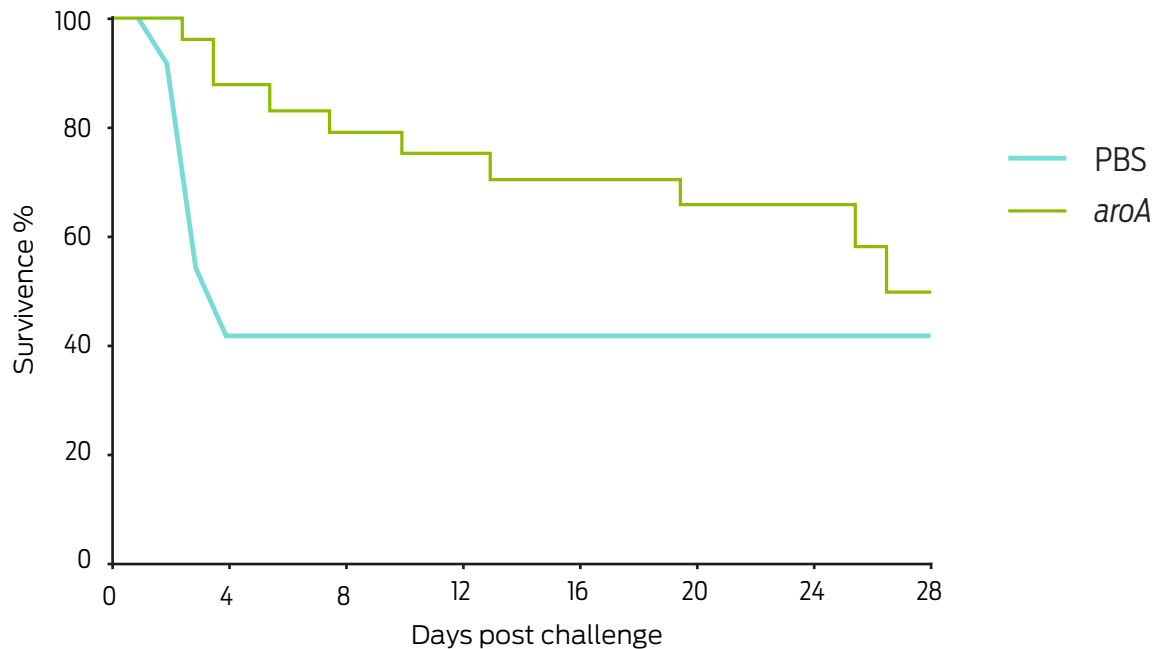


Figure 3. Survival percentage of Balb/c mice inoculated with the *aroA* mutant and challenged on day 21 with 1×10^4 CFU of the ATCC *C. pseudotuberculosis* strain, intraperitoneally inoculated. In the first week post-challenge, a marked survival difference between the control group (41.67 %) and the *aroA* mutant the vaccinated group (83.33 %) was observed ($P < 0.05$); however at the end of the experiment (day 28), 50 % survival was observed in mutant vaccinated mice, whereas the WT-vaccinated group survival did not change.

difference relative to the control group regarding the production of the IgG1 isotype between day 28 and day 42 post-vaccination ($P < 0.05$). For IgG2a, a statistically significant difference between the control and vaccinated groups was only noted on day 28 (Figure 4). For IgM and IgG2b, no significant difference was observed.

Cytokine measurement induced by vaccination

Measuring the cytokines induced by the *aroA* mutant in our murine model revealed that IL-10 exhibited a significant difference between day 7 and day 12 post-vaccination (Figure 5). A study of the IFN- γ , TNF- α and IL-4 responses revealed no statistically significant differences during the experiment.

This study was performed because of the need for a live vaccine to prevent caseous lymphadenitis. To date, the mutant strains that have been evaluated as possible *C. pseudotuberculosis* immunogens have failed to confer adequate protection to susceptible animals^{9, 21, 22}. Here, the *aroA* mutant of *C. pseudotuberculosis* was chosen based on the good results that were obtained by applying the *aroA* mutant of *Salmonella enterica*, subsp. *enterica* serovar Abortus equi, to guinea pigs through oral and intramuscular routes. This mutant provided the vaccinated animals with 100 % protection against an experimental challenge with the virulent strain¹⁰.

The observed difference between the recovered CFU/ml values of the WT and *aroA* mutant strains after 30 min p.i. in J774A murine macrophages (Figure 1) can be attributed to the *aroA* gene mutation, which decreases the regulation of the *murA* gene and, in turn, increases peptidoglycan synthesis. The resulting defects in the bacterial wall causes phagocytosis and bacterial elimination⁶.

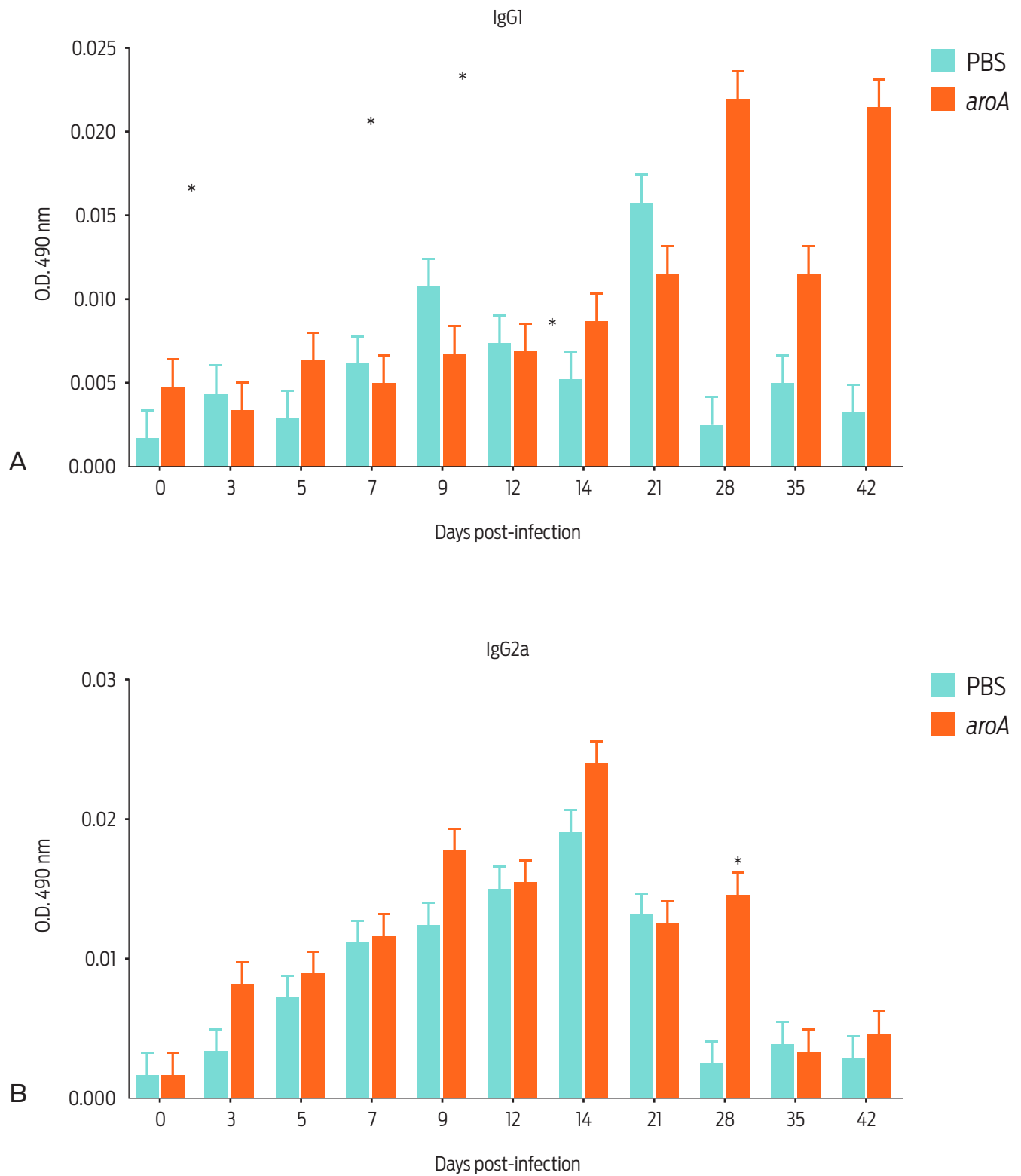


Figure 4. Humoral immune response of mice vaccinated with the *aroA* mutant strain after different post vaccination times: IgG1 immune response (a), and IgG2a immune response (b). Data are expressed as the mean \pm standard deviation of the sera from four mice vaccinated with *aroA* mutant strain and a control group injected with PBS. Statistically significant differences ($P < 0.05$) are indicated with *.

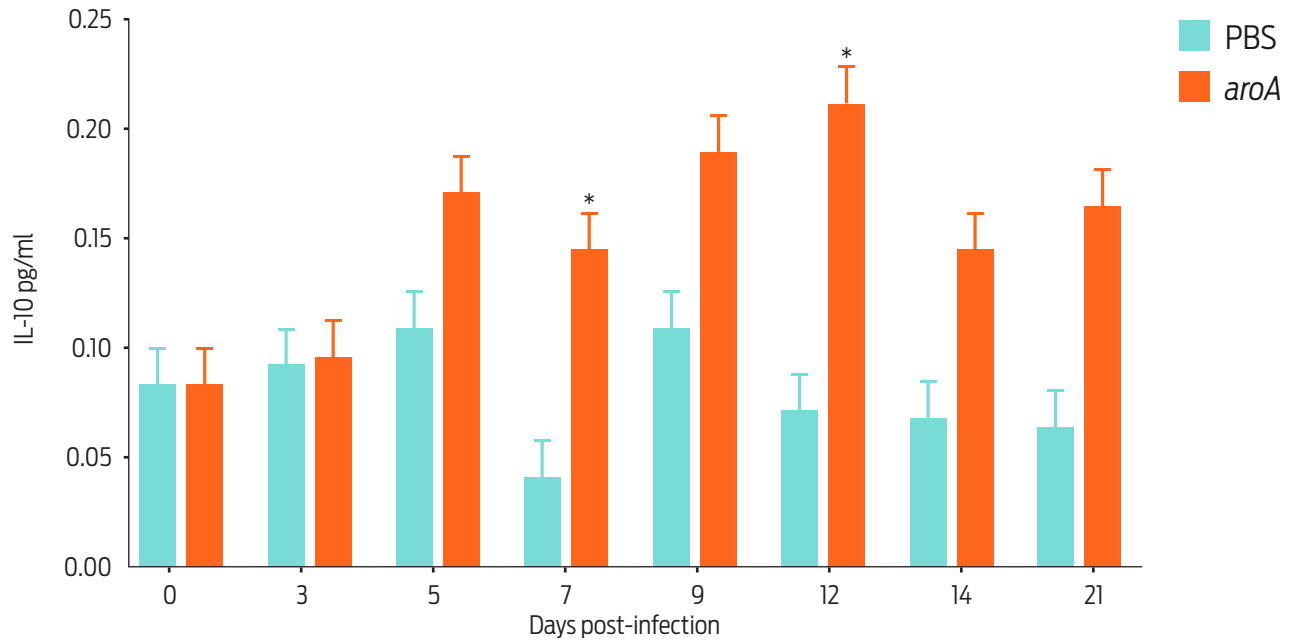


Figure 5. Cytokine detection in the culture supernatant of spleen lymphocytes from *aroA* mutant-vaccinated and control (PBS) BALB/c mice. The data are shown as the mean \pm standard deviation of groups of five mice on each post-vaccination day. Statistically significant differences ($P < 0.05$) are indicated with *.

One limitation on the use of immunogens to prevent caseous lymphadenitis is that they cause liquefactive necrosis at the vaccination site²³. Although some degree of inflammation is desirable because it is associated with macrophage recruitment and, thus, antigen presentation, the damage should not be extensive and should not produce necrosis. We suggest that if the bacterium's virulence was reduced, it would be efficiently phagocytosed by macrophages, which would then present the antigenic components of the bacteria and facilitate the recruitment of inflammatory cells⁶. The application of the *aroA* mutant in the footpads of mice resulted in injuries that were less severe than those caused by the WT strain. Although the *aroA* mutant's virulence was attenuated, it still had the potential to cause abscess at the application site. Such injuries are typical of actinomycetes because these bacteria have lipids in their cell walls that induce inflammation and the modulation of dendritic cells and macrophages¹⁷. However, the phospholipase D action must also be considered in this context because it is the main virulence factor causing injury and damage to cell membranes. Activating this gene is undesirable because the virulence of the resulting mutants was abolished and did not stimulate the immune system²⁴.

The reduced CFU/ml recovery in the liver and spleen of *aroA* mutant-vaccinated mice indicates that the host controlled this strain more effectively than the WT strain. Similar results have been reported for the *aroQ* mutant of *C. pseudotuberculosis*¹¹ and the *aroA* mutants of *S. enterica*⁶ and *Listeria monocytogenes*⁶. In these studies, it was observed that the mutant strain was recovered in lower amounts from the liver and spleen samples of BALB/c mice on days 7 and 14 post-vaccination.

Although the attenuation of the *aroA* mutant strain of *C. pseudotuberculosis* was observed relative to the WT strain in the cellular model and less injury and

residual virulence were observed, the survival rate of the vaccinated group was only 50 % by day 28 post-challenge (Figure 4). Thus, the mutant failed to confer at least 80 % protection, which is desirable for an immunogen, in the vaccinated mice¹⁹. Previously, an immunogen against caseous lymphadenitis was developed by deleting the *pld* gene of *C. pseudotuberculosis*, which is related to the production of phospholipase D, an exotoxin that is considered the major bacterial virulence factor²⁵. Studies in sheep immunized with a *pld* mutant showed 44 % protection after being challenged with a WT strain²¹. An Hsp60 recombinant protein was used in another study, but it did not provide sufficient protection in BALB/c mice that were challenged with a 1×10^6 CFU i. p. Actually, 100 % mortality was observed on day 11 post-vaccination.

In our assay, the humoral immune response of the mutant-vaccinated group involved relatively high amounts of IgG1 after day 28 post-vaccination. This type of response contributes to the Type Helper 2 (TH2) stimulation response by activating antigen-presenting cells and cloning B cells to produce neutralizing antibodies²⁶. The application of the *aroA* mutant in mice only stimulated IL-10 after day 7 post-vaccination and induced a poor cellular response as the IFN- γ levels only increased on day 5 post-vaccination in the vaccinated mice. This finding was related to the poor IgG2a production associated with the Type Helper 1 (TH1) response. The behavior of this cytokine was very similar to that reported by Simmons *et al.*¹¹, who immunized BALB/c mice with the *aroQ* mutant of *C. pseudotuberculosis*. The authors found that the levels of this cytokine increased significantly in the vaccinated animals only after day 7 post-vaccination and determined that the TH1 response was poor. The cellular immune response is known to be important for the resolution (or control) of diseases caused by intracellular bacteria, such as *C. pseudotuberculosis*, but its *aroA* mutant bacteria failed to elicit an adequate TH1 response. This was reflected by the *aroA* mutant's poor ability to adhere to and invade the murine cells, thereby reducing the bacterial antigen presentation by the main histocompatibility complex type I because these antigens were not found in adequate levels in the intracellular compartments⁹.

Conclusions

This study intended to attenuate *Corynebacterium pseudotuberculosis* virulence by affecting the process by which their cell wall is synthesized. This outcome was achieved because the *aroA* mutant showed lower multiplication in the cell model, and mice inoculated with the mutant showed less severe lesions than those inoculated with the wild type bacteria. However, this bacterium exhibited other virulence factors that were similar to the PLD exotoxin that must be considered when developing live vaccines.

The *aroA* mutant of *C. pseudotuberculosis* exhibited attenuation in the cellular model and its residual virulence and successfully stimulated antibody production. Nevertheless, it failed to generate a sufficient cellular response and did not adequately protect the vaccinated BALB/c mice against the experimental challenge. In the mouse model, mutant *aroA* only had the capacity to protect 50 % of Balb/C mice at 28 days' post- challenge, so the *aroA* mutant of *C. pseudotuberculosis* not it is a good candidate as an immunogen.

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Conflicts of interest

The authors declare that they have no competing interests.

Author contributions

Cristina Ibarra Zazueta: Conducted the experimental work and wrote the manuscript.

Efrén Díaz Aparicio: Designed the research and wrote the manuscript.

Beatriz Arellano Reynoso, Rigoberto Hernández Castro, and Erika Gabriela Palomares

Resendiz: Supervised the research and wrote the manuscript.

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